Effect of Evaporation Losses on Experimental Continuous Culture Results

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(Accepted for publication 8 January 1972)

SUMMARY

Evaporation can lead to errors in the measurement of parameters such as yield and maintenance coefficient, using continuous cultures at dilution rates below 0.05 h⁻¹.

INTRODUCTION

This paper shows how simple practical details, if ignored, can lead to false conclusions regarding the kinetic model used for a continuous culture system. The need to humidify the air, when operating a continuous culture system at low dilution rates, can be assessed by measuring the function

\[ \frac{Q_b}{Z_e/X} = \frac{X_e}{X} \]

where \( X \) and \( X_e \) are the cell concentrations, which would be observed for evaporating and non-evaporating conditions, respectively. As shown later, this function becomes significant at dilution rates below 0.05 h⁻¹, values which were used in work reported for example by Marr, Nilson & Clark (1963) to calculate a maintenance energy constant (\( a \)), by Schulze & Lipe (1964) and by Tempest, Herbert & Phipps (1967). This error can be avoided by fixing a condenser on the gas outlet of the fermenter.

The rate of evaporation, \( D_a \), will be a function of several parameters, including the stirrer speed, the temperature and humidity of the inlet air, the temperature of the liquid medium, the flow rate of air and the design of the vessel. The effect of evaporation has been calculated for three different theoretical models (Fig. 2 and 3).

THEORY

In Fig. 1, \( F_0 \) is the inlet liquid flow rate, \( F_1 \) the exit liquid flow rate and \( F_2 \) the evaporation rate of water into the air stream, all in units of litres of liquid per hour. The basic assumptions of a completely stirred tank reactor (C.S.T.R.) are assumed to hold for the models developed (i.e. the organism and substrate concentrations in the vessel are completely homogeneous and equal to their exit concentrations).

Monod's model. This is based on Monod's model (Monod, 1942, 1950) for the growth rate, \( \mu \).

Organism balance:

\[ V \frac{dx_e}{dt} = \frac{V\mu_m x_e S_e}{K_s + S_e} - F_1 x_e. \]  \hspace{1cm} (1)

Accumulation = Input - Output.

Substrate balance:

\[ V \frac{dS_e}{dt} = F_0 S_e - F_1 S_e - \frac{V\mu_m x_e S_e}{Y(K_s + S_e)}. \]  \hspace{1cm} (2)

Overall balance:

\[ F_0 = F_1 + F_2. \]  \hspace{1cm} (3)
At steady state, \( \frac{dx}{dt} = \frac{dS}{dt} = 0 \).

From equation (1):

\[
0 = \frac{\mu_m \bar{x}_e \bar{S}_e}{K_s + \bar{S}_e} - D_1 \bar{x}_e.
\]

From equation (2):

\[
0 = D_0 \bar{S}_e - D_1 \bar{S}_e - \frac{\mu_m \bar{x}_e \bar{S}_e}{Y(K_s + \bar{S}_e)}.
\]

From equation (3):

\[
D_1 = D_0 - D_2,
\]

where \( D_1 = \frac{F_i}{V} \), therefore \( \bar{S}_e = K_s D_1 (\mu_m - D_1) \) and \( \bar{x}_e = \frac{Y}{D_1(D_0 \bar{S}_e - D_1 \bar{S}_e)} \).

If \( D_0 = D_1 \), \( \bar{x} = Y(\bar{S}_e - \bar{S}) \), giving the steady-state cell concentration of the normal Monod model without evaporation.

Therefore, the expression for the parameter, \( \phi \), becomes

\[
\phi = \frac{\bar{x}_e}{\bar{x}} = \frac{D_0 \bar{S}_e - D_1 \bar{S}_e}{D_1(\bar{S}_e - \bar{S})}.
\]

At low values of \( D, \bar{S} \) and \( \bar{S}_e \approx \bar{S}_e \), therefore \( \phi \approx \frac{D_0}{D_0} = \frac{D_0}{(D_0 - D_2)} \).

**Endogenous model.** The second model is based on Herbert's formulation of endogenous metabolism (Herbert, 1958), in which the constant, \( k \), accounts for the oxidation of cell substance to \( CO_2 \). This constant endogenous metabolism added to the anabolic metabolism of the culture gives the modified equation set for evaporation.

**Organism balance:**

\[
V \frac{dx}{dt} = V \left( \frac{(\mu_m + k)S_e}{K_s + S_e} - k \right) x_e - F_1 x_e.
\]

**Substrate balance:**

\[
V \frac{dS}{dt} = F_0 S_e - F_1 S_e - \frac{V(\mu_m + k) x_e S_e}{Y(K_s + S_e)}.
\]

As before, the steady-state equations are solved to give:

\[
\bar{S}_e = \frac{K_s(D_1 + k)}{\mu_m - D_1},
\]

\[
\bar{x}_e = \frac{Y_e(D_0 S_e - D_1 \bar{S}_e)}{D_1 + k},
\]

\[
D_0 = D_1 + D_2.
\]

For \( D_0 = D_1 \), we obtain the cell concentration under non-evaporating conditions.

\[
\bar{x} = \frac{Y_e D_0 (S_e - \bar{S})}{D_0 + k}.
\]

Therefore

\[
\bar{x}_e = \frac{D_0 S_e - D_1 \bar{S}_e}{D_0 + k} \frac{D_0 + k}{(D_1 + k)(S_e - \bar{S})} D_0,
\]

and at low dilution rates, \( \phi \approx (D_0 + k)(D_0 + k - D_2) \).

**Maintenance model.** This was suggested by Pirt (1965), who included a specific maintenance term (\( a \)) accounting for the consumption of the limiting substrate to provide energy to maintain integrity and viability, but no biomass.

This maintenance model, when modified for evaporation, gives the equation set.

**Organism balance:**

\[
\frac{dx}{dt} = \frac{\mu_m S_e}{K_s + S_e} x_e - D_1 x_e.
\]
Evaporation losses in continuous cultures

\[
F_0 S_R \quad \text{due to evaporation} \quad F_2
\]

Fig. 1. Evaporating model.

Table 1.

<table>
<thead>
<tr>
<th>Type and reference</th>
<th>Air flow rate (l liquid/h)</th>
<th>Temp. (°C)</th>
<th>Agitator speed (rev./min)</th>
<th>( D_2 ) (h(^{-1}))</th>
</tr>
</thead>
<tbody>
<tr>
<td>Continuous culture (Pirt, 1957)</td>
<td>2.0</td>
<td>37</td>
<td>1030</td>
<td>0.003</td>
</tr>
<tr>
<td>Continuous culture (this work)</td>
<td>0.7</td>
<td>25</td>
<td>250</td>
<td>0.0013</td>
</tr>
<tr>
<td>Continuous culture (this work)</td>
<td>1.2</td>
<td>45</td>
<td>450</td>
<td>0.00275</td>
</tr>
<tr>
<td>250 ml shake flask with 100 ml of culture (this work)</td>
<td>-</td>
<td>30</td>
<td>125</td>
<td>0.00034</td>
</tr>
</tbody>
</table>

Substrate balance:

\[
\frac{dS_e}{dt} = D_0 S_R - D_1 S_e - \frac{\mu_m x_e S_e}{Y_e (K_s + S_e)} - \frac{a x_e}{Y_g}.
\]

As before, the steady-state equations are solved to give

\[
\bar{S}_e = \frac{K_s D_1}{\mu_m - D_1},
\]

\[
\bar{x}_e = \frac{Y_e (D_0 S_R - D_1 \bar{S}_e)}{D_1 + a},
\]

\[
D_0 = D_1 + D_2.
\]

For non-evaporating conditions,

\[
\bar{x} = \frac{Y_e D_0}{D_0 + a} \left( S_R - \bar{S} \right),
\]

and therefore

\[
\phi = \frac{\bar{x}_e}{\bar{x}} = \frac{D_0 S_R - D_1 \bar{S}_e}{(D_1 + a)(S_R - \bar{S})} \frac{D_0 + a}{D_0};
\]

and at low dilution rates, \( \phi = (D_0 + a)/(D_0 + a - D_2) \), which is identical in form to the simplified expression for the endogenous metabolism models.
Fig. 2. Monod’s model errors. Curve 1, $D_1 = 0.001$; curve 2, $D_2 = 0.002$; curve 3, $D_3 = 0.003$.

Fig. 3. Endogenous metabolism or maintenance model errors. --, $k$ (or $a$) = 0.03; ---, $k$ or $(a) = 0.01$; curve 1, $D_1 = 0.001$; curve 2, $D_2 = 0.002$; curve 3, $D_3 = 0.003$. 
Evaporation losses in continuous cultures

EXPERIMENTAL

Realistic values of the parameters in these expressions for $\phi$ may be obtained from the literature, except for the effective evaporation rate ($D_e$). Pirt (1957) has published a value for evaporation in vigorously agitated laboratory-scale continuous culture. This is included together with some experimental values for $D_e$ obtained by us (Table 1).

The parameter, $\phi$, was calculated from the simplified expression derived in the theory section and plotted against $D_0$ for a practical range of $D_0$, as shown in Fig. 2 and 3. The expression for $\phi$ is identical for the endogenous and maintenance models and therefore the effect of different values of $'k'$ and $'a'$, respectively, will be identical.

DISCUSSION

The figures indicate that, for negligible endogenous metabolism, the accepted formulae connecting $D$, $\bar{x}_e$ and $S_e$ need modification at low dilution rates and Herbert's model should read,

$$
\bar{x}_e = \frac{Y_p D_0 S_k - D_1 S_e}{D_1 + k} \\
= \frac{D_1}{D_1 + k} \left( \frac{D_0}{D_1} S_k - S_e \right).
$$

The culture behaves exactly as if the dilution rate were $D_1$, as indeed it is as far as the organism is concerned, with the ingoing substrate concentration of $D_0 S_k / D_1$. Even where the endogenous metabolism or maintenance energy constant is relatively large, the errors are not negligible. Steady state can be obtained at dilution rates down to $0.004$ h$^{-1}$ (Postgate & Hunter, 1962; Tempest et al., 1967) in bacterial cultures and that fungal cultures are operated at dilution rates as low as $0.0025$ h$^{-1}$ (Holme & Zacharias, 1965) with an endogenous metabolism constant $k$ of nearly zero.

The low value for evaporation in shake flasks (Table 1; $D_2 = 0.0034$ h$^{-1}$) must be considered in the context of the period of time the shake flask is on the rotary incubator. At this rate of evaporation the error in the measurement of organism concentration rises linearly with time and would be approximately 4% after five days.

We conclude therefore that errors in organism concentrations at low dilution rates can be significant especially for systems with low endogenous metabolism rates, when no special precautions are taken to reduce evaporation losses. When evaluating the endogenous metabolism constant ($k$) by plotting $1/x$ versus $1/D_0$, the slope of the line is equal to $(k - D_2)/Y_p S_k$ and not $k/Y_p S_k$. Thus, if evaporation were not taken into account the value of $'k'$ (or $'a'$) would be underestimated by an amount $D_2$ h$^{-1}$.

Symbols

- $a$: Substrate utilization constant due to maintenance requirements h$^{-1}$
- $D$: Dilution rate h$^{-1}$
- $F_0$: Inlet liquid flow rate l/h
- $F_1$: Exit liquid flow rate l/h
- $F_2$: Evaporation rate of water l/h
- $k$: Substrate utilization constant due to endogenous metabolism h$^{-1}$
- $K_s$: Monod's substrate constant, numerically equal to $S$ at which the rate is $\frac{1}{2} \mu_m$ g/l
- $S$: Substrate concentration under non-evaporating conditions g/l
Substrate concentration under evaporating conditions

Inlet substrate concentration

Time

Organism concentration under non-evaporating conditions

Organism concentration under evaporating conditions

Potential yield constant for the Monod model

Potential yield constant for the endogenous metabolism model

Potential yield constant for the maintenance model

Specific growth rate

Maximum growth rate

\[ \mu(S) = \frac{\dot{x}}{x} \]

\[ \phi = \frac{\dot{x}}{\ddot{x}} \]

(-) Indicates steady-state values

REFERENCES


