Serine/threonine/tyrosine phosphorylation regulates DNA binding of bacterial transcriptional regulators

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Reversible phosphorylation of bacterial transcriptional regulators (TRs) belonging to the family of two-component systems (TCSs) is a well-established mechanism for regulating gene expression. Recent evidence points to the fact that reversible phosphorylation of bacterial TRs on other types of residue, i.e. serine, threonine, tyrosine and cysteine, is also quite common. The phosphorylation of the ester type (phospho-serine/threonine/tyrosine) is more stable than the aspartate phosphorylation of TCSs. The kinases which catalyse these phosphorylation events (Hanks-type serine/threonine protein kinases and bacterial protein tyrosine kinases) are also much more promiscuous than the TCS kinases, i.e. each of them can phosphorylate several substrate proteins. As a consequence, the dynamics and topology of the signal transduction networks depending on these kinases differ significantly from the TCSs. Here, we present an overview of different classes of bacterial TR phosphorylated and regulated by serine/threonine and tyrosine kinases. Particular attention is given to examples when serine/threonine and tyrosine kinases interact with TCSs, phosphorylating either the histidine kinases or the response regulators. We argue that these promiscuous kinases connect several signal transduction pathways and serve the role of signal integration.

Many effector domains of bacterial RRs have DNA-binding capacity. This allows RRs to function as transcriptional regulators (TRs) and consequently change gene transcription when they become phosphorylated. In Escherichia coli, osmoregulation of porin proteins OmpF and OmpC is under transcriptional control of the TCS EnvZ/OmpR. The phosphorylation of OmpR by EnvZ changes its affinity for the promoter region of ompF and ompC, resulting in different transcriptional levels of these genes (Forst et al., 1989; Rampersaud et al., 1994). The effector domains can also perform enzymic activities, bind RNA or engage in protein–protein interactions (Gao et al., 2007; Galperin, 2010). The TCS that consists of the HK CheA and two RRs, CheY and CheB, is responsible for regulating the chemotaxis in E. coli. Phosphorylated CheY binds to flagellar motor switches to increase the tumble frequency, resulting in different swimming behaviour of the cell (Alon et al., 1998), whilst phosphorylated CheB has higher methyltransferase activity, which diminishes the activity of chemotaxis receptors (Alon et al., 1999). Although the effector domains are diverse, the majority of them are DNA-binding domains. The classification of the effector domains...
from >9000 bacterial RRs suggests that 65% of them have DNA-binding capacity and act as TRs (Gao et al., 2007).

In addition to kinase activity, many HKs also possess phosphatase activity, which allows them to dephosphorylate their cognate RRs. An in vitro phosphorylation assay suggested that EnvZ transfers the phosphate group to OmpR within 30 s and then rapidly dephosphorylates OmpR (the half-life of phosphorylated OmpR is <30 s) (Aiba et al., 1989). The Rhizobium meliloti HK FixL also can dephosphorylate its RR FixJ (Lois et al., 1993). In addition to the short half-life of phospho-histidine, the phosphatase activity of some HKs also ensures that their signal is quickly extinguished.

One of the most impressive feature of TCSs is the specificity of HKs for their RRs (Hoch & Varughese, 2001; Szurmant et al., 2007). A HK usually only phosphorylates its own cognate RR and discriminates against all other RRs. This specificity is determined by the amino acid residues involved in the interaction between the histidine phosphotransfer domain of the HK and the regulatory domain of the RR (Laub & Goulian, 2007; Szurmant & Hoch, 2010). This has been elegantly demonstrated with HK subdomain chimaeras. The histidine phosphotransfer domain of E. coli EnvZ was replaced by the corresponding sequences of RstB and CpxA, two other HKs from E. coli. The resulting chimaeric EnvZ could phosphorylate only RstA and CpxR, the cognate RRs of RstB and CpxA, respectively. The chimaeric EnvZs efficiently discriminated against all the remaining 32 RRs present in E. coli (Skerker et al., 2008).

Based on the above-mentioned features, the prototype bacterial TCS could be described as a linear device for very rapid signal transduction, soliciting a quick response to one particular environmental stimulus. Having said that, there exists a large diversity amongst bacterial TCSs, in terms of speed, complexity of the signalling pathway and ability to engage in cross-talk with other regulators. Some of these features will be highlighted in the following sections, as we discuss the interaction of TCSs with other bacterial phosphorylation-based signalling devices.

**Bacterial TRs can be regulated by other types of phosphorylation**

Examination of the available genome sequences reveals that TCSs are widely distributed in bacteria, and are present in archaea and some eukarya. As ~65% of all bacterial RRs are TRs, TCSs typically account for 5–25% of all TRs in a bacterial cell (this fraction varies considerably amongst different bacterial phyla). Whilst TCS TRs are regulated by reversible phosphorylation, it is widely presumed that other bacterial TRs are regulated by reversible ligand binding and not by phosphorylation. However, recent phosphoproteomics analyses have revealed that many bacterial TRs can be phosphorylated on serine, threonine and tyrosine residues (Macek et al., 2007; Prisic et al., 2010; Soufi et al., 2010; Derouiche et al., 2013). Moreover, some recent studies suggest that non-TCS bacterial TRs can also be phosphorylated on arginine (Schmidt et al., 2014), histidine (Hammerstrom et al., 2015) and cysteine residues (Sun et al., 2012).

**Serine/threonine phosphorylation of bacterial TRs**

In the past few years, it has been established that Hanks-type serine/threonine kinases (STKs) can phosphorylate TRs in many bacteria and regulate different functions, such as antibiotic resistance, virulence, capsule synthesis and sporulation (Wright & Ulijasz, 2014). The human pathogen *Staphylococcus aureus* possesses two Hanks-type STKs, Stk1 and Stk2, both implicated in regulating virulence and antibiotic resistance (Ohlsen & Donat, 2010; Tamber et al., 2010). Recently, it was established that the *S. aureus* Stk1 phosphorylates and regulates the activity of the global TR MgrA. MgrA belongs to the SarA/MgrA family of TRs that comprise a dimerization domain and a winged helix-turn-helix (HTH) motif (Cheung et al., 2008) (Fig. 1a). MgrA controls different virulence factors, such as the x-toxin, coagulase, protein A, autolysins and the synthesis of capsular polysaccharides. In addition, MgrA also controls the synthesis of several efflux pumps implicated in antimicrobial resistance in *S. aureus* (Truong-Bolduc et al., 2008). Stk1 phosphorylates purified MgrA at two adjacent serines, Ser110 and Ser113, located in the dimerization domain interface (Truong-Bolduc & Hooper, 2010) (Fig. 1a). It was suggested that this phosphorylation antagonizes MgrA dimerization and DNA binding, leading to de-repression of its target genes and therefore activation of the efflux pump under antibiotic-imposed stresses (Truong-Bolduc & Hooper, 2010). Another *S. aureus* global TR controlled by serine/threonine phosphorylation is SarA (Chien et al., 1999; Didier et al., 2010). SarA also controls virulence factors: the x-toxin, the immune evasion molecule SpA, and the haemolysins and proteolytic exotoxins (Cheung et al., 2008). In addition, SarA controls host colonization by regulation of the ica operon responsible for biofilm formation (Valle et al., 2003), as well as capsule synthesis and adherence (Cheung et al., 2008) (Fig. 1a). Didier et al. (2010) have shown that SarA is phosphorylated by both Stk1 and Stk2. Stk1 controls SarA by threonine phosphorylation, which enhances its binding to promoters of key target genes (Didier et al., 2010). In contrast, Stk2 phosphorylates SarA only on serine residue(s) and this provokes a decrease in DNA binding to known SarA-regulated promoters (Didier et al., 2010). Further work is needed on the SarA regulatory system to define the effects of phosphorylation with respect to infection and virulence. Another example of TRs regulated by STKs in *S. aureus* is CcpA (catabolite control protein A). CcpA belongs to a conserved family of LacI/GalR TRs and is a global gene regulator of the central carbon metabolism in many bacterial species. In *Firmicutes*, the HPp protein phosphorylated at a specific
Serine residue interacts with CcpA and facilitates its binding to operator sites (Mijakovic et al., 2002; Fujita, 2009). CcpA is also involved in controlling virulence factors in *S. aureus* and many other bacterial pathogens (Seidl et al., 2006). Recently, it was shown that *S. aureus* CcpA is phosphorylated by Stk1, *in vivo* and *in vitro*, on two threonine residues (Thr18 and Thr33) (Fig. 1a). These residues are situated in the CcpA DNA-binding domain and are presumed to interact with the target DNA by forming hydrogen bonds. CcpA phosphorylation by Stk1 abrogates the protein–DNA interaction, and leads to activation of CcpA-repressed promoters implicated in sugar metabolism and biofilm formation (Leiba et al., 2012) (Fig. 1a). Interestingly, CcpA Thr18 and Thr33 are highly conserved in *Firmicutes* (Fig. 1b). Whilst it has been shown that CcpA orthologues from several other species could not be phosphorylated by their cognate STKs, cross-species phosphorylation was shown to be possible (Leiba et al., 2012).

The pathogenic bacterium *Mycobacterium tuberculosis* possesses a relatively large arsenal of STKs (11 kinases, named PknA to PknL). This bacterium is known for the emergence of antibiotic resistance phenomena, some of which can be related to regulation dependent on STKs (Molle & Kremer, 2010). Interestingly, many proteins regulated by *M. tuberculosis* STKs are TRs that play crucial roles in virulence. One of these TRs is EthR, a transcriptional repressor (belonging to the TetR family), which regulates the activation process of the antitubercular drug ethionamide (Baulard et al., 2000). Ethionamide is a pro-drug that is activated by a specific M. tuberculosis enzyme to a hydroxamate that is rapidly converted to its active ethionamide form, a pro-drug that is rapidly converted to its active ethionamide form.

![Diagram](https://via.placeholder.com/150)

**Fig. 1.** *Staphylococcus aureus* TRs controlled by STKs. (a) TRs are represented in tan and STKs in red. Phosphorylation reactions are shown as red arrows. Domain architecture and identified phospho-residues are shown for each TR. The dimerization domains (DD) and periplasmic binding domains (PDB) are shown in blue, and the DNA-binding HTH domains are shown in yellow (wHTH, winged HTH; lHTH, LacI family HTH). (b) Alignment of CcpA N termini from different bacteria: *Bacillus selenitireducens* MLS10; *Brevibacillus brevis*; *Bacillus licheniformis*; *Bacillus subtilis* 168; *Lactobacillus brevis* ATCC 367; *Lactobacillus casei* BL23; *Streptococcus pneumoniae* 670-6B; *Streptococcus mutans*; *Streptococcus pyogenes* MGAS2096; *Staphylococcus aureus* N315; *Crotinia butulinum* B; *C. sticklandii*; *C. botulinum* B. Residues phosphorylated by Stk1 in *Staphylococcus aureus* CcpA are indicated with red arrows.
must undergo bioactivation by the mono-oxygenase EthA (DeBarber et al., 2000). Molle and colleagues have shown that EthR is a substrate of the M. tuberculosis STK PknF \textit{in vitro} (Leiba et al., 2014). MS analysis has identified four phosphorylated residues in the unstructured N terminus of the EthR: Thr2, Thr3, Ser4 and Ser7. When these phospho-residues were mutated, \textit{in vitro} binding of EthR to its target ethA promoter decreased (Leiba et al., 2014). Another \textit{M. tuberculosis} TR phosphorylated by STKs is Rv2175c, which is considered to be implicated in cell wall synthesis (Cohen-Gonsaud et al., 2009). This was discovered in a proteomics-based study searching for novel kinase substrates (Canova et al., 2008). It was shown that PknL phosphorylates Rv2175c on a key threonine residue (Thr9), and phosphorylation disrupts the interaction of Rv2175c with the DNA (Cohen-Gonsaud et al., 2009).

In the Firmicutes model organism Bacillus subtilis, there is one global gene regulator, AbrB, known to be phosphorylated by STKs (Kobir et al., 2014). AbrB binds a number of DNA target sequences with the common feature of being structurally flexible and undergoing a conformational change upon TR binding (Bobay et al., 2004). AbrB acts as a global TR (Chumsakul et al., 2011), regulating the expression of stationary-phase functions. Recently, AbrB was reported to be phosphorylated at residue Ser86 in a phosphoproteomics study (Soufi et al., 2010). Kobir et al. (2014) have shown that AbrB can be phosphorylated by three \textit{B. subtilis} STKs: PkC, PkD (YbdM) and YabT. Phosphorylation of AbrB abolishes its binding to target promoters, causing activation of exoprotease production, and antagonizing sporulation and competence development (Fig. 2a). The phosphorylation site of \textit{B. subtilis} AbrB is conserved in other bacteria, mostly Bacillus spp. (Fig. 2b), suggesting that this type of regulation may exist elsewhere.

**Tyrosine phosphorylation of bacterial TRs**

Recently, Derouiche et al. (2013, 2015) discovered two bacterial TRs regulated by tyrosine phosphorylation in \textit{B. subtilis}. The first TR is FatR, a member of the TetR family of TRs, which regulates the metabolism of polyunsaturated fatty acids. In \textit{B. subtilis}, FatR represses the operon fatR–cytA3 and can be displaced from the target sequence in the presence of fatty acids (Lee et al., 2001). \textit{Bacillus subtilis} CytA3 is a cytochrome P450 fatty acid mono-oxygenase which hydroxylates unsaturated long-chain and branched-chain fatty acids in subterminal positions (Gustafsson et al., 2004; Lentz et al., 2004). FatR is phosphorylated by a cognate bacterial tyrosine kinase (BY kinase), PtK (Derouiche et al., 2013). Phosphorylation of FatR occurs at residue Tyr43 in the HTH domain (Fig. 2a). Tyr45 is directly involved in the interaction of the HTH motif with the DNA backbone via a hydrogen bond. Its phosphorylation disrupts the interaction of FatR with the DNA (Derouiche et al., 2013). The \textit{in vivo} consequence of FatR phosphorylation is de-repression of the fatR–cytA3 operon. As Tyr45 is highly conserved (Fig. 2c), \textit{B. subtilis} PtK was able to phosphorylate FatR orthologues from other bacteria (Derouiche et al., 2013). This suggests that tyrosine phosphorylation of the TetR HTH might be a widespread mechanism of transcriptional control of genes implicated in \textit{β}-oxidation. Next, the \textit{B. subtilis} protein SalA was shown to act as a TR and this is regulated by tyrosine phosphorylation (Derouiche et al., 2015). SalA is a member of the ubiquitous family of Mrp ATPases which are present in eukarya and bacteria (Dardel et al., 1991; Vitale et al., 1996). SalA had been previously described as an indirect positive regulator of the production of the exoprotease AprE, by inhibiting the expression scoC which codes for a repressors of aprE (Ogura et al., 2004). Derouiche et al. (2015) have shown that the binding of SalA to its target DNA (scoC promoter) depends on the presence of ATP and is stimulated by phosphorylation of SalA at Tyr327 by the BY kinase PtK (Fig. 2a). This phosphorylation activates SalA ATP binding and hydrolysis, leading to repression of scoC and increased production of the exoprotease AprE (Derouiche et al., 2015).

**Cysteine phosphorylation of bacterial TRs**

The staphylococcal TRs of the SarA/MarR family were recently found to be phosphorylated on cysteine residues (Sun et al., 2012). This type of phosphorylation is considered to be a rare post-translational modification, usually with no known regulatory roles. In \textit{S. aureus}, the STK Stk1 phosphorylates a number of TRs on cysteines: Cys9 of SarA, Cys13 of SarZ and Cys12 of MgrA, within their N-terminal dimerization domain (Fig. 2a) (Sun et al., 2012). \textit{In vivo} and \textit{in vitro} evidence, as well as the structural analysis, demonstrate that Stk1-dependent cysteine phosphorylation regulates several virulence traits and vancomycin resistance (Sun et al., 2012).

**Phosphorylation of TCS HKs and RRs by STKs**

Over recent years an increasing number of studies have described unconventional TCS RRs. In this section we summarize several examples of TCS in which the RRs lack a cognate HK (orphan RRs) and require phosphorylation by a STK. In some cases, STK-dependent phosphorylation also occurs on RRs which have a designated HK. This double phosphorylation, catalysed by HKs and STKs, respectively, diversifies the inputs modulating RR activity. There are also reported cases of cross-phosphorylation between HKs and STKs in bacteria. The \textit{B. subtilis} TCS DegS/DegU is part of a complex regulatory network involving the control of competence, exoprotease production, motility and complex colony and biofilm formation (Msaed et al., 1990; Ogura et al., 2001; Verhamme et al., 2007; Kobayashi, 2007; Mader et al., 2002). DegS is a cytosolic HK, which seems to integrate various signals pertaining to the metabolic condition of the cell. Interestingly, DegS was found to be phosphorylated on...
residue Ser76, situated in its signal-sensing domain (Macek et al., 2007). Jers et al. (2011) demonstrated that DegS Ser76 can be phosphorylated by two *B. subtilis* STKs: PrkD (YbdM) and YabT (Fig. 3a). Phosphorylation of DegS Ser76 stimulates its HK kinase activity and the transfer of phosphate to the RR DegU. As a consequence, the non-phosphorylated degS mutant S76A behaves like a strain with low levels of DegU-P in vivo. These results suggest that STK-dependent phosphorylation of the HK DegS can act as an additional input for activating this TCS.

**Fig. 2.** *Bacillus subtilis* TRs controlled by serine/threonine/tyrosine phosphorylation. (a) TRs are represented in tan and kinases in red. Phosphorylation reactions are shown as red arrows. Domain architecture and identified phospho-residues are shown for each TR. The dimerization domains are shown in blue, and the DNA-binding HTH domains are shown in yellow (the AbrB DNA-binding domain is also referred to as the loop-hinge helix fold). (b) Alignment of AbrB C-termini from different bacteria: Parvae, *Paenibacillus larvae*; Ppolymyxa, *Paenibacillus polymyxa* E681; Hemodesticaldum, *Heliobacterium modesticaldum* Ice1; Bsutilis, *Bacillus subtilis* 168; Bamyoliquefaciens, *Bacillus amyloliquifaciens* DSM7; Batrophaeus, *Bacillus atrophaeus* 1942; Blicheniformis, *Bacillus licheniformis* DSM13; Bcereus, *Bacillus cereus* G9842; Apasteurianus, *Acetobacter pasteurianus*. The residue phosphorylated by STKs in *B. subtilis* AbrB is indicated with a red arrow. (c) Alignment of N-termini of *Bacillus subtilis* FatR orthologues from different bacteria: Bthuringiensis, *Bacillus thuringiensis* BMB171; Bcereus, *Bacillus cereus*; Bmegaterium, *Bacillus megaterium*; Bsutilis, *Bacillus subtilis* 168; Lcasei, *Lactobacillus casei* BL23; Abaumannii, *Acinetobacter baumannii*; Pseudomonasm, *Pseudomonas mendocina*; Smitis, *Streptococcus mitis* SK597; Kpneumoniae, *Klebsiella pneumoniae* subsp. pneumoniae; Senterica, *Salmonella enterica*. The residue phosphorylated by the *Bacillus subtilis* BY kinase PtkA is indicated with a red arrow.
In *Streptococcus pneumoniae*, RitR (repressor of iron transport regulator) is a TCS-like TR that is required for lung pathogenicity. It controls iron uptake and remediation of iron-catalysed reactive oxygen species (Throup et al., 2000; Ulijasz et al., 2004; Ong et al., 2013). RitR is annotated as a TCS RR, but because it does not possess a cognate HK it is called an orphan RR (Ulijasz et al., 2004). Instead of a conserved aspartate residue in its regulatory domain, RitR has an asparagine residue at the expected phosphate acceptor site. *Streptococcus pneumoniae* phospho-serine/threonine protein phosphatase PhpC, and its cognate STK StkP, were identified as interaction partners of RitR (Ulijasz et al., 2009) (Fig. 3b). StkP was further shown to phosphorylate the RitR C-terminal DNA-binding domain in vitro. Further in vitro studies have suggested that PhpP and StkP compete for interaction with RitR. Both StkP and PhpP seem to affect the expression of RitR target genes in vivo, i.e. the Piu haem transporter. These observations suggest that the orphan RR RitR is in fact regulated by STK-dependent phosphorylation.

Cross-talk between Stk1/Stop1 and GraS/GraR signalling pathways was shown in *S. aureus* (Fridman et al., 2013). Stk1/Stop1 is a Hanks-type STK/phosphatase pair and GraS/GraR is a TCS which controls the resistance to cationic antimicrobial peptides. A study by Fridman et al. (2013) reported that Stk1 specifically phosphorylates the RR protein of the GraS/GraR TCS, GraR, at the DNA-binding domain, which increases its DNA-binding activity. Thr128, Thr130 and Thr149, located in the N terminus of its DNA-binding domain, were suggested as phosphorylation sites. This phosphorylation was found to be dependent on the intact tertiary structure of GraR, as denatured GraR did not undergo phosphorylation by Stk1. The specificity of the phosphotransfer between Stk1 and GraR was further investigated in BceR, a homologue of Stk1 in *B. subtilis*, which did not exhibit Stk1-dependent phosphorylation. GraR is involved in regulation of the *dltABCD* operon, which provides the addition of α-alanine to the wall teichoic acid. Wall teichoic acid isolated from *S. aureus* RN6390 AgraR strain showed reduced α-alanine content.

**Fig. 3.** Overview of the cross-talk between bacterial TCSs and STKs. Two-component HKs and RRs are represented in tan, STKs are in red and phosphatases are in green. Domain architecture is indicated for phosphorylated RRs, DNA-binding domains are in yellow, regulatory domains (RD) are in blue and dimerization domains (DD) in light grey. Identified phosphorylated residues are indicated. The depicted TCSs are (a) *Bacillus subtilis* DegS/DegU, (b) *Streptococcus pneumoniae* orphan RR RitR, (c) *Staphylococcus aureus* VraS/VraR, (d) *M. tuberculosis* DosS/DosT/DosR, (e) streptococcal CovS/CovR and (f) *Staphylococcus aureus* GraS/GraR. ROS, reactive oxygen species.
This result suggests that Stk1 an modulate the modification of wall teichoic acid. Depending on the environmental signals, expression of the \( \text{dltABCD} \) operon is controlled by two distinct phosphorylations of GraR, catalysed by either Stk1 or GraS.

In \textit{S. aureus}, the TCS RR VraR is phosphorylated by a cognate HK, VraS (Belcheva & Golemi-Kotra, 2008). In addition, it is also phosphorylated by a cognate STK, Stk1 (Canova et al., 2014) (Fig. 3c). VraR belongs to the \textit{S. aureus} vancomycin resistance-associated sensor and RR system (VraTSR). VraTSR responds to several antibiotics targeting the cell wall. VraR, the RR, modulates the expression of the cell wall stress regulon in response to antibiotics. VraR is regulated by phosphorylation catalysed by VraS and Stk1 (Canova et al., 2014). Stk1-dependent phosphorylation sites on VraR were determined \textit{in vitro}, and confirmed by site-directed mutagenesis. These comprise four threonines: Thr106, Thr119, Thr175 and Thr178. Stk1-mediated phosphorylation sites are of crucial importance for VraR activity. Residues Thr175 and Thr178 are located in the HTH domain of VraR, while the other two sites are in the VraR regulator domain. The structural context of the phosphorylation sites suggests that VraR phosphorylation should inhibit its DNA binding, and this was confirmed experimentally (Canova et al., 2014). VraR regulation is a prominent example of a bacterial TR that is regulated simultaneously by two types of phosphorylation: aspartate and threonine, catalysed by a HK VraS and a STK Stk1, respectively.

A similar case of two phosphorylation systems converging on the same RR was observed in \textit{M. tuberculosis}. Dormancy in \textit{M. tuberculosis} is mediated by the RR DosR (Park et al., 2003). The DosR regulon is transcriptionally activated in response to hypoxia, carbon monoxide and nitric oxide. The activation of the DosR-regulated genes is triggered by two cognate HKs: DosS and DosT (Roberts et al., 2004; Kumar et al., 2007). In addition to DosR- and DosT-dependent phosphorylation, DosR can also be phosphorylated by a STK PknH (Fig. 3d). This phosphorylation occurs on DosR residues Thr198 and Thr205, situated in the key regulatory helix \( \alpha \)-10 (Chao et al., 2010). DosR aspartate and threonine phosphorylations act synergistically; both were shown to enhance DosR DNA binding \textit{in vitro}. Consequently, both types of phosphorylation correlate with transcriptional activation of the DosR regulon \textit{in vivo}. As the effect is synergistic, both types of phosphorylation are required to achieve full transcriptional response.

In the case of DosR, aspartate and threonine phosphorylation elicit a synergistic effect. By contrast, the two phosphorylation systems converging on the streptococcal RR CovR have antagonistic effects. The CovS/CovR TCS regulates the expression of genes involved in the production of capsule, major virulence factors, penetration of blood–tissue barriers and avoidance of the immune system by group A/B streptococci (Federle et al., 1999; Jiang et al., 2005; Whidbey et al., 2013). Rajagopal et al. (2006) showed that deletion of the STK Stk1 leads to inability to produce the key virulence factor \( \beta \)-haemolysin/cytolysin. This effect is based on Stk1-dependent phosphorylation of the residue Thr65 in CovR (Fig. 3e). CovR Thr65 phosphorylation by Stk1 antagonizes CovR phosphorylation at Asp53 by CovS. This inhibitory effect also extend in the opposite direction; CovR phosphorylation at Asp53 decreases Stk1-dependent phosphorylation at Thr65. As a consequence, phosphorylation of CovR at Thr65 (by Stk1) antagonizes the activating effect of CovS-dependent phosphorylation and ultimately decreases CovR affinity for DNA targets.

**Concluding remarks**

Phosphorylation of bacterial TRs by STKs and BY kinases at first glance serves the same purpose as phosphorylation of TCS RRs by their HKs. Phosphorylation affects the affinity of TRs for DNA and serves as an activity switch. However, aspartate phosphorylation of RRs is inherently short-lived, and the signal is rapidly transmitted and quickly extinguished. Conversely, serine/threonine and tyrosine phosphorylation is chemically much more stable in the bacterial cytosol, and dedicated phosphatases are required to remove it from phosphorylated TRs. Another important difference between HKs, on the one hand, and STKs and BY kinases, on the other, is substrate selectivity. HKs discriminate very strictly amongst cognate RRs and typically phosphorylate only one or two targets. STKs and BY kinases are much less specific, and their actions are more pleiotropic (Shi et al., 2014a; Wright & Ulijasz, 2014). Each STK and BY kinase can phosphorylate a number of different cellular substrates (Mijakovic & Deutscher, 2015), and this relaxed substrate specificity can be traced to a lack of co-evolution between the kinase and its substrates (Shi et al., 2014b). These kinases are often capable of extensive cross-talk with other kinases (Shi et al., 2014a). STKs and BY kinases in some cases functionally interact with the TCSs, phosphorylating either HKs or RRs. In this context, we would argue that STKs and BY kinases may act more like signal-integrating than simple signal-transmitting devices. The impact of TCSs on regulation of gene transcription has often been described as a rapid and reversible ‘on/off’ response. With STKs and BY kinases, the dynamics of the response are likely to be different. These kinases are known to phosphorylate substrates less efficiently and are therefore likely to elicit fine-tuning than a classical ‘on/off’ response. Therefore, it would be interesting to see some time-resolved \textit{in vivo} studies measuring the phosphorylation stoichiometry of TRs phosphorylated on serine/threonine/tyrosine residues, and even more interesting to correlate these to the effects on target gene transcription. The thermodynamic stability of phospho-serine/threonine/tyrosine residues probably means that the regulatory effects of phosphorylation may last longer. In several cases described above, serine/threonine/tyrosine phosphorylation of TRs acts as a secondary regulatory mechanism, in addition to some previously known...
ligand or stimulus. This strengthens the notion of fine-tuning or, in some cases, desensitizing the TR to the primary ligand/signal. Evaluating the impact of this transcriptional fine-tuning on the systems level will soon be possible by combining time-resolved transcriptomics and phosphoproteomics studies.

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is regulated by serine phosphorylation in its input domain. PLoS One 6, e14653.


