Mitochondrial nucleoids from the yeast *Candida parapsilosis*: expansion of the repertoire of proteins associated with mitochondrial DNA

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Molecules of mitochondrial DNA (mtDNA) are packed into nucleic acid–protein complexes termed mitochondrial nucleoids (mt-nucleoids). In this study, we analysed mt-nucleoids of the yeast *Candida parapsilosis*, which harbour a linear form of the mitochondrial genome. To identify conserved as well as specific features of mt-nucleoids in this species, we employed two strategies for analysis of their components. First, we investigated the protein composition of mt-nucleoids isolated from *C. parapsilosis* mitochondria, determined N-terminal amino acid sequences of 14 proteins associated with the mt-nucleoids and identified corresponding genes. Next, we complemented the list of mt-nucleoid components with additional candidates identified in the complete genome sequence of *C. parapsilosis* as homologues of *Saccharomyces cerevisiae* mt-nucleoid proteins. Our approach revealed several known mt-nucleoid proteins as well as additional components that expand the repertoire of proteins associated with these cytological structures. In particular, we identified and purified the protein Gcf1, which is abundant in the mt-nucleoids and exhibits structural features in common with the mtDNA packaging protein Abf2 from *S. cerevisiae*. We demonstrate that Gcf1p co-localizes with mtDNA, has DNA-binding activity in vitro, and is able to stabilize mtDNA in the *S. cerevisiae Δabf2* mutant, all of which points to a role in the maintenance of the *C. parapsilosis* mitochondrial genome. Importantly, in contrast to Abf2p, *in silico* analysis of Gcf1p predicted the presence of a coiled-coil domain and a single high-mobility group (HMG) box, suggesting that it represents a novel type of mitochondrial HMG protein.

INTRODUCTION

Mitochondrial nucleoids (mt-nucleoids) are cytological structures that represent fundamental segregating units of the mitochondrial genome (Kuroiwa, 1982; Sakai et al., 2004). These nucleic acid–protein complexes undergo dynamic remodelling in response to changes in cellular metabolism (Kucej et al., 2008). In baker's yeast, a number of proteins have been identified as specific components of isolated mt-nucleoids (Miyakawa et al., 1995; Miyakawa & Sato, 2001; Sato et al., 2002; Sato & Miyakawa, 2004) and/or cross-linked mitochondrial DNA (mtDNA)–protein complexes (Kaufman et al., 2000; Chen et al., 2005). The mt-nucleoids contain proteins known to interact with mtDNA such as the non-histone protein Abf2, which is considered to be a major mtDNA packaging factor, and a set of enzymes involved in mtDNA replication, transcription and repair. In addition, a number of metabolic enzymes have been shown to be bona fide components of the mt-nucleoids, thus linking mitochondrial inheritance with cell metabolism and physiology (reviewed by Chen & Butow, 2005; Kucej & Butow, 2007). Investigations of human mt-nucleoids indicate that these complexes are linked to the mitochondrial inner membrane and have a layered structure in which mtDNA replication and transcription occur in the central core, whereas RNA processing, translation and assembly of the respiratory chain complexes may occur in the periphery (Wang & Bogenhagen, 2006; Bogenhagen et al., 2008).

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Abbreviations: DAPI, 4',6-diamidino-2-phenylindole; EMSA, electrophoretic mobility shift assay; GST, glutathione S-transferase; HMG, high-mobility group; mtHMG, mitochondrial HMG protein; mtDNA, mitochondrial DNA; mt-nucleoid, mitochondrial nucleoid; mtTBP, mitochondrial telomere-binding protein; NP-40, Nonidet P40; yEGFP3, yeast enhanced version 3 of GFP.
The mtDNA packaging protein Abf2 from Saccharomyces cerevisiae is the best-characterized mt-nucleoid component. It is a 20 kDa DNA-binding protein that contains two high-mobility group (HMG) box domains and, in the presence of topoisomerase I, it introduces negative supercoils into a relaxed circular DNA plasmid (Caron et al., 1979; Diffley & Stillman, 1991). In Abf2p-deficient yeast cells, mtDNA is rapidly lost during growth on glucose media, suggesting that Abf2p plays a key role in the maintenance of the mitochondrial genome (Diffley & Stillman, 1991, 1992). This protein is specifically associated with the isolated mt-nucleoids and has the highest affinity for dsDNA among the mt-nucleoid proteins (Miyakawa et al., 1995) with a preference for GC-rich gene sequences in mtDNA (Kucej et al., 2008). It is also involved in the regulation of the copy number and the recombination of mtDNA (Zelenaya-Troitskaya et al., 1998).

In previous studies (Miyakawa et al., 2000; Umezaki & Miyakawa, 2002) we have employed an SDS-DNA PAGE approach to detect the Abf2-like mtDNA-binding proteins in several yeast species, and found only one such protein per analysed species, with molecular masses ranging from 17 to 26 kDa, suggesting that phylogenetically distant yeast species tend to differ in the mode of mtDNA packaging. Abf2-like proteins have been purified from the mt-nucleoids of four species, S. cerevisiae, Kluyveromyces lactis, Pichia jadinii and Williopsis mrakii (Miyakawa et al., 1995, 2000, 2003; Miyakawa & Yawata, 2007). Comparative analysis of K. lactis mt-nucleoid proteins indicates that Abf2p is the most divergent component of the mt-nucleoid (Nosek et al., 2006).

In contrast to most fungal species, Candida parapsilosis mitochondria contain a linear mitochondrial genome of 30 923 bp, terminating with arrays of telomeric repeats (n x 738 bp, where n=1–12; Nosek et al., 1995, 2004). In addition to the linear mtDNA molecules, mitochondria of this yeast contain a series of circular dsDNA molecules termed telomeric circles (t-circles), which are implicated in the mitochondrial telomere transactions (Tomaska et al., 2000; Nosek et al., 2005). The unique molecular architecture of the C. parapsilosis mitochondrial genome prompted us to initiate analysis of its mt-nucleoids and to identify their conserved as well as specific features that may have contributed to the evolutionary emergence of the linear genome form in yeast mitochondria.

In this report, we experimentally identified several proteins that co-purify with mt-nucleoids from C. parapsilosis. In addition, we complemented the protein set with candidates detected by in silico analysis of the complete genome sequence of C. parapsilosis. Importantly, we analysed in more detail a 29 kDa protein named Gcf1p and provide experimental evidence for its association with mt-nucleoids in this species. C. parapsilosis Gcf1p is a novel member of the mitochondrial high-mobility group protein (mtHMG) family, originally identified in silico as putative mtDNA packaging proteins from Candida albicans and Debaryomyces hansenii (Nosek et al., 2006; see also Visacka et al., 2009), which are distantly related to non-histone proteins from S. cerevisiae (Abf2p), slime mould (Glom) and human (h-mtTF1) mitochondria (Diffley & Stillman, 1991; Fisher & Clayton, 1988; Dequard-Chablat & Alland, 2002).

**METHODS**

**Yeast strains and cultivation.** C. parapsilosis strains CBS604T and SR23 (ade2, lys4) were employed in this study. The type strain CBS604T used for isolation of mt-nucleoids was provided by H. Fukuhara (Institut Curie, Orsay, France). Cells were cultured aerobically at 30 °C to stationary phase in yeast extract-peptone-dextrose (YPD) medium that contained 1% (w/v) yeast extract (Oriental Yeast Company, Japan), 2% (w/v) peptone (Kyokuto, Japan) and 2% (w/v) glucose. S. cerevisiae strain YAM101 (MATa/a, Abf2:: TRP1/ABF2, ade2-1/ade2-1, ura3-1/ura3-1, his3-11,15, trp1-1/trp1-1, leu2-3,112/leu2-3,112, can1-100/can1-100), provided by D. A. Clayton (Howard Hughes Medical Institute, Chevy Chase, MD, USA), is the parental strain of YAM101-1a and YAM101-1b, the haploids prepared by tetrad dissection of YAM101, which retained the wild-type ABF2 allele and Δabf2:: TRP1, respectively.

**Isolation of mt-nucleoids from C. parapsilosis cells.** The mt-nucleoids were isolated essentially as described previously (Miyakawa et al., 1987). In brief, cells (about 40 g wet weight) were converted to spheroplasts by treatment with Zymolyase 20T (Seikagaku Kogyo). Mitochondria were prepared from disrupted spheroplasts by differential centrifugation. The mt-nucleoids were isolated from mitochondria after lysis of mitochondrial membranes with Nonidet P-40 (NP-40). Each mitochondrial pellet was suspended in NES2 buffer (0.5 M sucrose, 20 mM Tris/HCl, pH 7.6, 2 mM EDTA, 0.8 mM spermidine, 7 mM 2-mercaptoethanol, 0.4 mM PMSF) at a protein concentration of 0.25 mg ml⁻¹ (Figs 2 and 3a, lane 2), and then NP-40 was added to a final concentration of 0.5% (w/v) with gentle stirring for 10 min. The samples were centrifuged at 15 000 g for 10 min to separate supernatant (Figs 2 and 3a, lane 3) from
NP-40-insoluble material (Figs 2 and 3a, lane 4). The supernatant was then centrifuged at 45 000 g for 1 h to separate the mt-nucleoid fraction (Figs 2 and 3a, lane 6) from NP-40-soluble fraction (Figs 2 and 3a, lane 5). The mt-nucleoid fraction was suspended in 1 ml NES2 buffer and stored at −80 °C until use. In this preparation, the purification step using sucrose density-gradient centrifugation was omitted.

**Purification of a 29 kDa protein from the mt-nucleoid fraction.**

The *C. parapsilosis* 29 kDa protein was purified by a combination of NaCl extraction of the mt-nucleoid fraction, HiTrap-SP column chromatography (Sasaki et al., 2003) and DNA–cellulose chromatography. The isolated mt-nucleoids were suspended in NES2 buffer containing 0.4 M NaCl, and incubated for 30 min at 25 °C. After centrifugation at 45 000 g for 30 min, the supernatant was dialysed overnight against HEPES buffer (50 mM HEPES, 150 mM NaCl, 1 mM DTT, pH 7.5). The dialysed fraction was loaded onto the HiTrap-SP column (HiTrap SP HP 1 ml column; Amersham Biosciences) and the column was washed with HEPES buffer. The column was eluted with an NaCl gradient of 150–500 mM in HEPES buffer. The fractions that contained the 29 kDa protein were pooled and threefold with HEPES buffer, and the resulting sample was loaded onto a dsDNA–cellulose column (0.1 ml in a syringe). The column was washed with HEPES buffer and the 29 kDa protein was eluted with HEPES buffer containing 1 M NaCl.

**SDS-PAGE and SDS-DNA PAGE.** For SDS-PAGE, an aliquot of each sample (15 μl) was mixed with 5 μl 4 × SDS sample buffer and loaded on a 7.5–15 % polyacrylamide gel. SDS-DNA PAGE for detecting DNA-binding proteins was performed as described previously (Miyakawa et al., 2000). The stacking and separating gels contained 10 μg ml⁻¹ native calf thymus DNA. After overnight incubation of the gels in 10 mM Tris/HCl, 1 mM EDTA, pH 7.5, 10 mM MgCl₂ and 7 mM 2-mercaptoethanol at 30 °C, the gels were stained with ethidium bromide (1 μg ml⁻¹) for 30 min. Proteins were detected by the silver-staining method (Oakley et al., 1980). A low-molecular-mass calibration kit (Pharmacia) was used for the estimation of molecular mass. For the determination of the N-terminal amino acid sequence, proteins were transferred to a PVDF membrane and the sequences were determined using a Protein Sequencer PPSQ-21 (Shimazu).

**Expression and purification of recombinant Gcf1 protein.** The plasmid pGEX-2T-GCF1, expressing the recombinant version of the Gcf1 protein in fusion with glutathione S-transferase (GST–Gcf1p) was constructed as follows. The GCF1 ORF was amplified by PCR from the genomic DNA of *C. parapsilosis* strain CBS604T using two oligonucleotide primers, 5'-ATGTGAGCTCATGGTAAACT-3' and 5'-TAGCTGTGCTTCTTTAGATTGTG-3'. The PCR product was then inserted into expression vector pGEX-2T (Amersham Biosciences) digested with EcoRI and blunt-ended with Klenow fragment of DNA polymerase I.

*Escherichia coli* DH5α cells (Clontech) transformed with the plasmid pGEX-2T-GCF1 were cultivated in LBA medium [1 % (w/v) peptone, 0.5 % (w/v) yeast extract, 0.9 % (w/v) NaCl, 100 μg ampicillin ml⁻¹] until the culture reached OD₆₀₀ 0.6, then IPTG was added to a final concentration of 1 mM and expression was induced for 3 h at 30 °C. Cells were harvested by centrifugation (10 min, 4 °C, 10 000 g), washed once with sterile ice-cold water, resuspended in 5 ml LR buffer (20 mM Tris/HCl, pH 7.4, 100 mM NaCl, 1 mM EDTA, 10 mM 2-mercaptopethanol) containing 1 mg lysozyme ml⁻¹ and 1 x Complete protease inhibitor (Roche Molecular Biochemicals), and incubated for 20 min on ice. The suspension was sonicated for 20 s and then incubated on ice for 20 s. The sonication step was repeated five times. Subsequently, Triton X-100 to a final concentration of 0.1 % (v/v) and 50 U DNase I (Promega) were added, and the suspension was incubated for 15 min on ice and cleared by centrifugation (15 min, 16 000 g, 4 °C). The supernatant was mixed with 300 μl glutathione agarose beads (Sigma-Aldrich) and gently agitation for 1 h at 4 °C. The beads were then washed three times with 700 μl LRT buffer [20 mM Tris/HCl, pH 7.4, 100 mM NaCl, 1 mM EDTA, 10 mM 2-mercaptoethanol, 0.1 % (v/v) Triton X-100]. The recombinant protein was eluted with 500 μl ER1 buffer (50 mM Tris/HCl, pH 7.4, 100 mM NaCl, 1 mM EDTA, 15 mM glutathione). All fractions were analysed by 10 % SDS-PAGE electrophoresis. The recombinant protein was stored at −20 °C.

**Electrophoretic mobility shift assay (EMSA).** Purified recombinant protein GST–Gcf1 (40–100 ng) was mixed with 150 ng *C. parapsilosis* mtDNA digested with BglII and incubated for 1 h at room temperature. The samples were electrophoretically separated in 0.6 % agarose gels in 45 mM Tris-borate, 1 mM EDTA (0.5 x TBE) buffer and DNA was visualized by staining the gels with ethidium bromide...
(0.5 μg ml⁻¹). GST purified using the same procedure as described for GST–Gcf1 was used as a negative control.

**Fluorescence microscopy.** Cells and spheroplasts of *C. parapsilosis* were fixed, respectively, with 5% (v/v) glutaraldehyde in the culture medium and in 0.8 M sorbitol, 20 mM potassium phosphate buffer, pH 7.5, at room temperature for 1 h. Samples were washed twice with NS buffer (0.25 M sucrose, 20 mM Tris/HCl, pH 7.6, 1 mM EDTA, 1 mM MgCl₂, 0.1 mM ZnSO₄, 0.1 mM CaCl₂, 0.8 mM PMSF, 0.05% (v/v) 2-mercaptoethanol; Miyakawa et al., 1987) and mixed with an equal volume of a solution of 4’,6-diamidino-2-phenylindole (DAPI; 2 μg ml⁻¹) in NS buffer on a glass slide. The isolated mitochondria and mt-nucleoids were mixed directly with an equal volume of DAPI (2 μg ml⁻¹) solution. All observations were made with an epi-fluorescence microscope (BHS-RF, Olympus Optical Co.) equipped with appropriate objectives (Dplan Apo 100UVPL and 100UV, Olympus Optical Co.).

Intracellular localization of Gcf1p was done in *C. parapsilosis* SR23 cells transformed with the pBP6-GCF1 plasmid expressing Gcf1p fused with the enhanced version 3 of GFP (yEGFP3). The plasmid was constructed by insertion of the EcoRI fragment was excised from the construct and inserted into the pBP6 vector (Kosa et al., 2007) linearized with Xhol endonuclease. The termini of the vector and the insert molecules were blunt-ended with a Quick Blunting kit (New England Biolabs). The resulting plasmid, pBP6-GCF1, was transformed into *C. parapsilosis* SR23 (Nosek et al., 2002). Transformsants were selected in synthetic dextrose (SD) medium [2% (w/v) glucose, 0.17% (w/v) yeast nitrogen base without amino acids and ammonium sulfate, 0.5% (w/v) ammonium sulfate, 0.02 mg adenine ml⁻¹, 2% (w/v) agar] to visualize the Gcf1–
yEGFP3 fusion. The transformsants were cultivated overnight at 28 °C in liquid SD medium then for 6 h in synthetic galactose (SGal) medium, in which glucose was replaced by 2% (w/v) galactose. The cells were then observed using a BX50 microscope (Olympus Optical Co.).

**Functional complementation of the Δabf2 mutation in *S. cerevisiae*.** To demonstrate that Gcf1p is able to replace the Abf2p function(s) in *S. cerevisiae* cells, the plasmid pYES2/CT-GCF1 was constructed by insertion of GCF1 ORF lacking the termination codon, amplified from *C. parapsilosis* CBS604² genome DNA using the primers 5’-ATGTGGAGACTCATTGTTAACCCT-3’ and 5’-GATTGTGAATTTGACTCTTGTG-3’, into the pDrive cloning vector (Qiagen). Subsequently, the 771 bp EcoRI fragment was excised from the construct and inserted into the pBP6 vector (Kosa et al., 2007) linearized with Xhol endonuclease. The termini of the vector and the insert molecules were blunt-ended with a Quick Blunting kit (New England Biolabs). The resulting plasmid, pBP6-GCF1, was transformed into *C. parapsilosis* SR23 (Nosek et al., 2002). Transformsants were selected in synthetic dextrose (SD) medium [2% (w/v) glucose, 0.17% (w/v) yeast nitrogen base without amino acids and ammonium sulfate (Difco), 0.5% (w/v) ammonium sulfate, 0.02 mg adenine ml⁻¹, 2% (w/v) agar]. To visualize the Gcf1–yEGFP3 fusion, the transformsants were cultivated overnight at 28 °C in liquid SD medium then for 6 h in synthetic galactose (SGal) medium, in which glucose was replaced by 2% (w/v) galactose. The cells were then observed using a BX50 microscope (Olympus Optical Co.).

**RESULTS**

Isolation of the mt-nucleoids from stationary-phase yeast culture

*C. parapsilosis* cells and spheroplasts contain numerous mt-nucleoids as revealed by DAPI staining (Fig. 1a–c). In the case of *S. cerevisiae* and *K. lactis*, the mt-nucleoids were fractionated from mitochondria by sucrose density-gradient centrifugation, as reported previously (Miyakawa et al., 1987, 1995, 2003). However, the mt-nucleoids of *C. parapsilosis* underwent precipitation even after NP-40 treatment of mitochondria, and the yields of the mt-nucleoids decreased significantly after the sucrose density-gradient centrifugation. Therefore, we routinely fractionated the mt-nucleoids from mitochondrial lysates using differential centrifugation as described in Methods. DAPI staining of the isolated mt-nucleoids showed that they maintained their intact size (Fig. 1d). The patterns of DNA extracted from the mt-nucleoid fraction and digested with restriction enzymes (data not shown) corresponded to mtDNA reported in our previous studies (Nosek et al., 1995, 2004).

The mt-nucleoid fractions isolated from *C. parapsilosis* contained a subset of mitochondrial proteins (Fig. 2, lane 2 versus lane 6). SDS-DNA PAGE analyses of the mt-nucleoid proteins clearly detected the enrichment of a 29 kDa DNA-binding protein in the mt-nucleoids (Fig. 2, lane 11). In this assay we also detected a 37 kDa protein. This protein was abundant in the mitochondrial fraction rather than enriched in the mt-nucleoids. Similar results

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were obtained when the mt-nucleoid fraction from *S. cerevisiae* was analysed and, according to the molecular mass and the requirement for Mg$^{2+}$ ions for its appearance on the gel in the SDS-DNA PAGE assay, this protein was assumed to be the major mitochondrial nuclease Nuc1p (Miyakawa et al., 2000), as SDS-DNA PAGE detects nuclease activity as well as DNA-binding proteins (Rosenthal & Lacks, 1977). However, we did not investigate the nature of the 37 kDa protein from *C. parapsilosis*.

**Identification of proteins from the mt-nucleoid fraction**

We compared the protein composition of the mt-nucleoids with that of the mitochondrial fraction on 7.5, 10, 12.5 and 15 % gels, and 17 proteins present in the mt-nucleoid fraction were subjected to N-terminal sequence analysis as summarized in Fig. 3(a). With the exception of the 29 kDa protein, the majority of visualized proteins were not highly enriched in the mt-nucleoid fraction (see Discussion). We determined N-terminal sequences of 15 proteins, which suggested that the N-terminal amino acid was not blocked in most cases (Table 1), and subsequent TBLASTN searches using the peptide sequences as queries led to identification of 14 coding sequences in the *C. parapsilosis* genome sequence and, subsequently, revealed their homologues from *S. cerevisiae* and *C. albicans* (see Supplementary Tables S1 and S2). In the case of p63, we did not identify the corresponding coding sequence, and proteins p58 and p94 were not amenable to N-terminal sequence analysis. The identified protein set includes aconitase (Aco1p), subunits of pyruvate dehydrogenase (Lat1p, Pda1p and Pdb1p), mitochondrial ribosomal proteins (Mnp1p, Mrpl4p and Mrpl6p), mitochondrial telomere-binding protein (Mtp1p/mtTBP), cytochrome *c* (Cyc1p), cytochrome *c* oxidase subunit 4 (Cox4p), mitochondrial succinate–fumarate transporter (Sfc1p), mitochondrial translation elongation factor (Tuf1p), fatty-acyl coenzyme A oxidase (Pox1p), and an HMG box-containing protein, Gcf1 (see below).

**The 29 kDa protein is a putative mtDNA packaging protein**

The properties of the 29 kDa protein (i.e. molecular mass, highly basic pI, relatively high abundance in mt-nucleoids and DNA-binding activity in SDS-DNA PAGE) support the hypothesis that it represents an mtDNA packaging protein in *C. parapsilosis* and may have similar function(s) to those of Abf2p, and this prompted us to investigate this protein in more detail.

Previously, we have purified an Abf2p homologue from *K. lactis* mitochondria by extraction of the mt-nucleoids by treatment with 0.2 M HCl followed by hydroxyapatite column chromatography and dsDNA–cellulose chromatography.
The N-terminal amino acid sequence of the 29 kDa protein was determined to be STAKTTRSSKDKEEA (Table 1). A TBLASTN search using this peptide as a query against the C. parapsilosis genome database was effective, and the 29 kDa protein was purified to near homogeneity, as indicated by the presence of a single band on SDS-PAGE (Fig. 3b). The DNA-binding activity of the purified 29 kDa protein and its enrichment in the mt-nucleoids were confirmed by SDS-DNA PAGE assays. To confirm that the protein 29 kDa protein revealed that it can bind DNA substrates in DNA mobility, indicating that they are bound by the protein.

Recombinant Gcf1p is an mtDNA-binding protein that localizes to mitochondria

Analyses of the mt-nucleoid fraction as well as the purified 29 kDa protein revealed that it can bind DNA substrates in EMSA. The fusion protein showed a number of foci corresponding to protein dimers, possibly engaged in the formation of protein dimers, possibly compensating for the absence of the second HMG box.

The absence of an apparent preference for any of the conserved amino acid positions (Fig. 4).

Table 1. C. parapsilosis proteins identified in the mt-nucleoid fraction by N-terminal sequencing

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The C. parapsilosis genome sequence database revealed a single ORF, CPAG_03094, with a length of 251 codons. Bioinformatic analyses of deduced amino acid sequences by MitoProt II, PSORT II and MITOPRED indicated high probabilities (0.9673, 65.2 % and 99 %, respectively) of its being targeted to mitochondria (Supplementary Table S1). The import signal predicted by MitoProt II (amino acids 1–28) corresponds with the experimentally determined N-terminal sequence (amino acids 34–48) of the purified protein. Analyses with the COILS and SMART utilities revealed the presence of a coiled-coil domain (amino acids 4–106) and an HMG box (amino acids 166–230), respectively. A BLASTP search against the GenBank database revealed two hits with a relatively low score; EAK92709 (GCF1/orf19.400/orf19.8030; E-value 1e-10) and CAG90643 (DEHA0g13059 g; E-value 2e-05) in the C. albicans and Debaryomyces Hansenii genomes, respectively, which were identified in our previous in silico study (Nosek et al., 2006) as putative mtDNA packaging proteins from the mtHMG protein family. Based on common features with C. albicans Gcf1p (see Visacka et al., 2009), we named the 29 kDa protein (CPAG_03094) Gcf1p (or CpGcf1p) and the corresponding gene GCF1 (or CpGCF1). In contrast to S. cerevisiae Abf2p, which contains two HMG boxes involved in DNA binding, the Candida Gcf1 proteins apparently have only a single HMG box that corresponds to HMG box 2 of Abf2p. In addition, Gcf1 proteins have a coiled-coil domain not present in Abf2p, which could be engaged in the formation of protein dimers, possibly compensating for the absence of the second HMG box.

Although the Gcf1 and Abf2 proteins exhibit structural similarities, their overall homology is weak, with only a few conserved amino acid positions (Fig. 4).
the mtDNA fragments suggests that the protein binds to DNA without sequence specificity. The gradual shift of the DNA bands is probably caused by the fact that a population of DNA fragments of a given size is represented by molecules bound by different numbers of protein molecules.

**C. parapsilosis GCF1** complements the Δabf2 mutation in S. cerevisiae cells

Yeast cells lacking functional Abf2p are able to maintain mtDNA when cultivated in media with non-fermentable carbon sources, as Aco1p substitutes for its DNA-binding activity, but these cells quickly lose their mtDNA in glucose-containing medium when Aco1p is repressed (Diffley & Stillman, 1991, 1992; Chen *et al.* , 2005). Although Gcf1p exhibits very low sequence homology with Abf2p, amino acid alignment revealed several conserved residues, mostly within the HMG box 2. Therefore, we determined whether Gcf1p is able to substitute for Abf2p in mtDNA maintenance by complementation of a Δabf2 mutation in S. cerevisiae cells. We constructed the plasmid pYES2/CT-GCF1 containing the GCF1-coding sequence driven by the galactose-inducible GAL1 promoter, and introduced it into S. cerevisiae Dabf2 cells. The expression of GCF1 clearly increased the proportion of respiratory-competent cells of the Δabf2 mutant, indicating that Gcf1p participates in the maintenance of mtDNA in vivo.

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**C. parapsilosis genome**

To complement the set of mt-nucleoid proteins identified experimentally, we searched for other components by *in silico* analysis of the *C. parapsilosis* genome sequence. As the
bioinformatic analysis cannot itself confirm that they are components of mt-nucleoids in C. parapsilosis, we consider these proteins to be candidate components for participating in the structure and/or function of these nucleic acid–protein complexes. Using amino acid sequences of 30 bona fide mt-nucleoid proteins from S. cerevisiae (reviewed by Chen & Butow, 2005; Kucej & Butow, 2007) as queries in TBLASTN searches we identified almost all counterparts present in assembled contigs covering 9.2 times the C. parapsilosis genome (Supplementary Table S1). Our efforts did not lead to unambiguous identification of a homologue of Abf2 protein, as hits with the highest score were nuclear non-histone HMG proteins, such as Nhp6a and Nhp6b. This is in line with our hypothesis that Gcf1p represents the major mtDNA packaging protein in C. parapsilosis.

**DISCUSSION**

**Proteins associated with mt-nucleoids in C. parapsilosis**

In this study we experimentally identified 14 proteins present in the mt-nucleoid fraction prepared from C. parapsilosis mitochondria (Table 1, Fig. 3a). Among them, five proteins (Aco1p, Pda1p, Pdb1p, Mnp1p and Rim1p) were known to be bona fide mt-nucleoid components in S. cerevisiae, suggesting that at least some features of the mt-nucleoids are conserved between phylogenetically distant yeast species. The homologue of Rim1p has been previously identified in C. parapsilosis mt-nucleoids in vivo as mtTBP, which plays the dual role of single-stranded DNA-binding protein and mitochondrial telomere-binding factor (Tomaska et al., 1997, 2001; Nosek et al., 1999). Importantly, we detected a putative mtDNA packaging protein, Gcf1p, which is distantly related to Abf2p of S. cerevisiae, and an additional eight proteins (Cox4p, Cyc1p, Lat1p, Mrpl4p, Mrpl6p, Pox1p, Sfc1p and Tuf1p), whose counterparts have not been detected in S. cerevisiae mt-nucleoids. Moreover, we found a 37 kDa protein as a minor component of the mt-nucleoids, although its identity was not examined (Fig. 2). It remains to be

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**Fig. 6.** Gcf1p binds mtDNA in EMSA. The assay was performed using recombinant GST–Gcf1 protein and C. parapsilosis mtDNA digested with BglII (lanes 1–5). Purified GST was used in the control experiment (lanes 6 and 7). Digested mtDNA samples (150 ng) were incubated for 1 h at room temperature without (lane 1) or with 40 ng (lane 2), 60 ng (lane 3), 80 ng (lane 4) or 100 ng (lane 5) recombinant GST–Gcf1 protein, or 80 ng (lane 6) and 100 ng (lane 7) GST. Samples were separated by electrophoresis in a 0.6 % agarose gel. iDNA digested with PstI was used as the molecular mass marker (M).

**Fig. 7.** C. parapsilosis GCF1 complements Δabf2 mutation in S. cerevisiae. The plasmids pYES2/CT and pYES2/CT-GCF1 were transferred into the strains YAM101-1a (ΔABF2) and YAM101-1b (Δabf2). The transformants were cultivated for 24 h in SD or SGal liquid media and then inoculated in serial dilutions onto YPDG plates (labelled as SD→YPDG or SGal→YPDG).
determined which of these proteins represent species-specific features of *C. parapsilosis* mt-nucleoids. Alternatively, proteins not found in *S. cerevisiae* mt-nucleoids may be loosely associated and therefore have escaped detection in previous studies. The latter hypothesis is supported by the presence of protein subunits from the pyruvate dehydrogenase complex (Lat1p) and mitochondrial ribosome (Mrp1p and Mrp6p), as other proteins from these complexes (i.e. Pda1p, Pdb1p and Mnp1p) associate with mt-nucleoids in both yeast species. The occurrence of three mitochondrial ribosomal protein subunits (Mnp1p, Mrp14p and Mrp16p) and a translation elongation factor Tu (Tuflp) in the mt-nucleoid fraction is particularly interesting, as this supports a hypothesis that mitochondrial proteosynthetic machinery is closely linked with the structure and/or dynamics of mt-nucleoids, which is further strengthened, for example, by the coupling of transcription and translation in yeast mitochondria as well as the involvement of bona fide mt-nucleoid components such as Rpo41p, Sl1p and Mnp1p in these processes (Kaufman et al., 2000; Bryan et al., 2002; Rodeheffer & Shadel, 2003; Sato & Miyakawa, 2004). This is in line with the observation that mt-nucleoids purified from human mitochondria contain a number of mitochondrial ribosomal proteins, tRNA synthetases and several translation factors, including the elongation factor Tu, indicating that translation occurs in the peripheral region of the mt-nucleoids (Bogenhagen et al., 2008).

In general, it might be expected that the degree of enrichment in the mt-nucleoid fraction in the case of the proteins performing dual functions will be dependent on their distribution between the mt-nucleoid and the corresponding protein complex involved in a particular biochemical process (e.g. the Krebs cycle). For example, in *S. cerevisiae*, only a fraction of the mitochondrial enzyme aconitase (Aco1p) is associated with mt-nucleoids and participates in mtDNA maintenance (Chen et al., 2005). Naturally, proteins involved solely in mt-nucleoid functions, such as Gcf1p, should be enriched by the purification. Indeed, Gcf1p is highly enriched in the mt-nucleoid fraction and represents one of its major constituents. On the other hand, we must acknowledge that our mt-nucleoid fractions were prepared without density-gradient centrifugation and therefore may contain contaminants in addition to bona fide mt-nucleoid proteins. For example, cytochrome c (Cyc1p) is not likely to be an mt-nucleoid protein, and the conditions employed for solubilization of the inner membrane may induce artefactual association of cytochrome c with the mt-nucleoids due to its basic pl. Similarly, fatty-acyl coenzyme A oxidase (Pox1p) is not a mitochondrial protein and may represent contamination from peroxisomes.

Using *in silico* analysis of the *C. parapsilosis* complete genome sequence we identified additional candidate components of the mt-nucleoids. Our results indicate that the *C. parapsilosis* genome encodes a similar set of proteins to those known to be associated with mt-nucleoids of *S. cerevisiae* (Supplementary Table S1). In most cases, the homology between *C. parapsilosis* and *S. cerevisiae* proteins was relatively weak. Therefore, we also confirmed their identities by comparison with *C. albicans* counterparts and predicted the probability of their import into mitochondria (Supplementary Table S1). To illustrate overall conservation of the mt-nucleoid components from *C. parapsilosis*, we compared proteins identified experimentally as well as candidates detected by *in silico* analysis with their homologues from *S. cerevisiae* and *C. albicans* (Fig. 8, Supplementary Table S2). The analysis revealed that several proteins known to interact directly with mtDNA (i.e. Abf2p/Gcf1p, Mip1p, Pif1p, Rim1p/mtTBP and Rpo41p) are conserved relatively weakly. It is possible that the faster accumulation of changes in their sequences reflects the evolution of mtDNA sequences. However, this explanation may be acceptable only in certain cases, when proteins recognize specific motifs in mtDNA. For example, Rpo41p has been shown to bind the terminal sequences of the linear mtDNA molecules from *C. parapsilosis* (Tomaska et al., 1997, 2001; Nosek et al., 1999). Similarly, Rpo41p recognizes sequence motifs of the mitochondrial promoter (Matsunaga & Jaehning, 2004), which varies among yeast species. However, other proteins, such as Gcf1p, do not


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