The role of two periplasmic copper- and zinc-cofactored superoxide dismutases in the virulence of \textit{Salmonella choleraesuis}

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Periplasmic copper- and zinc-cofactored superoxide dismutases (Cu,Zn-SODs, SodC) of several Gram-negative pathogens can protect against superoxide-radical-mediated host defences, and thus contribute to virulence. This role has been previously defined for one Cu,Zn-SOD in various \textit{Salmonella} serovars. Following the recent discovery of a second periplasmic Cu,Zn-SOD in \textit{Salmonella}, the effect of knockout mutations in one or both of the original sodC-1 and the new sodC-2 on the virulence of the porcine pathogen \textit{Salmonella choleraesuis} is investigated here. In comparison to wild-type, while sodC mutants – whether single or double – showed no impairment in growth, they all showed equally enhanced sensitivity to superoxide and a dramatically increased sensitivity to the combination of superoxide and nitric oxide \textit{in vitro}. This observation had its correlate in experimental infection both \textit{ex vivo} and \textit{in vivo}. Mutation of sodC significantly impaired survival of \textit{S. choleraesuis} in interferon-\gamma-stimulated murine macrophages compared to wild-type organisms, and all \textit{S. choleraesuis} sodC mutants persisted in significantly lower numbers than wild-type in BALB/c (Ity\textsuperscript{+}) and C3H/HeN (Ity\textsuperscript{+}) mice after experimental infection, but in no experimental system were sodC-1 sodC-2 double mutants more attenuated than either single mutant. These data suggest that both Cu,Zn-SODs are needed to protect bacterial periplasmic or membrane components. While SodC plays a role in \textit{S. choleraesuis} virulence, the data presented here suggest that this is through overcoming a threshold effect, probably achieved by acquisition of sodC-1 on a bacteriophage. Loss of either sodC gene confers maximum vulnerability to superoxide on \textit{S. choleraesuis}.

\textbf{Keywords:} Superoxide dismutase, \textit{Salmonella}, periplasm, bacterial Cu,Zn-SOD, bacterial virulence

\textbf{INTRODUCTION}

\textit{Salmonella} serovars responsible for disease in various mammalian hosts can proliferate as facultative intracellular pathogens, able to survive within the phagosome and/or phagolysosome of macrophages (Buchmeier & Heffron, 1989). \textit{Salmonella} virulence in mice has been correlated with the net growth of \textit{Salmonella} within these cells of the reticuloendothelial system (Fields \textit{et al}., 1986). This suggests a capacity to resist host defences including the superoxide radical anion (O\textsuperscript{2-}) generated during the respiratory burst, nitric oxide (NO) produced by inducible NO synthase and a range of reactive molecules formed by reaction between these two species. These include various extremely reactive oxygen and nitrogen intermediates (ROIs, RNIs) such as the hydroxyl radical and peroxynitrite that exert a potent microbicidal effect through the indiscriminate oxidative damage they cause to lipids, proteins and DNA in cell...
membranes, the periplasm and cytosol. Diverse mechanisms are beginning to be defined which contribute to the capacity of Salmonella strains to withstand exogenous ROI and RNI stress. The superoxide radical causes oxidative damage to iron-sulphur clusters, and the product of the yggX gene has been shown to exert a protective effect (Gralnick & Downs, 2001). The microbicidal effects of RNIs derived from phagocytic cell metabolism are in part vitiated by a recently described bacterial alkyl hydroperoxide reductase (AhpC) (Chen et al., 1995), while the very production in the macrophage phagosome of these toxic species is impaired through the operation of a Salmonella pathogenicity island 2-encoded type III secretion system that mediates exclusion of NADPH oxidase assembly from the phagosome membrane (Gallos et al., 2001). Overall, the superoxide radical is a key intermediate in the production of these toxic species, and bacterial superoxide dismutases might accordingly be expected to play an important part in endowing pathogenic organisms with resistance to such host defences. Recent light has been thrown on this process with the description of the role of bacterial periplasmic copper- and zinc-cofactored superoxide dismutases ([Cu,Zn]-SODs) in the virulence of various serovars of Salmonella (Farrant et al., 1997; De Groote et al., 1997).

Superoxide dismutases (SODs) are virtually ubiquitous in bacteria, catalysing the conversion of $O_2^−$ into hydrogen peroxide and oxygen (McCord & Fridovich, 1969) in the first of a series of reactions to remove free radicals generated during the reduction of molecular oxygen. Three types of SOD are widely found in bacteria. Two are cofactored by manganese or iron and are located in the cytoplasm where they catalyse dismutation of $O_2^−$ generated in the course of aerobic metabolism. A third SOD is cofactored by copper and zinc ([Cu,Zn]-SOD) and has been identified in the periplasm of a wide range of Gram-negative bacteria [Photobacterium leiognathi (Steinman, 1987); Caulobacter crescentus (Steinman, 1982); Brucella abortus (Stabel et al., 1992); Haemophilus species (Langford et al., 1992; Kroll et al., 1995); Legionella species (St John & Steinman, 1996); Actinobacillus and Pasteurella species (Kroll et al., 1995); Escherichia coli (Benov & Fridovich, 1994; Inlay & Imlay, 1996); and Salmonella typhimurium, Salmonella choleraesuis and Salmonella dublin (Canvin et al., 1996; Farrant et al., 1997)]. Within the periplasm [Cu,Zn]-SOD is inaccessible to cytosolic superoxide but is suitably located to protect the organism from exogenous superoxide and the toxic action of its further reaction products with hydrogen peroxide and nitric oxide. Studies have confirmed such a function both in Salmonella and Neisseria meningitidis (Farrant et al., 1997; De Groote et al., 1997; Wilks et al., 1998).

The degree of dissimilarity (only 54% identity) between the translated Salmonella sodC sequence described by Farrant et al. (1997) and E. coli SodC, its aberrant map position with respect to the Salmonella and E. coli genomes, and its flanking regions encoding proteins resembling products of bacteriophage lambda, suggested that this gene was not the orthologue of the E. coli gene and that Salmonella may have acquired it through phage-mediated transfer from an unknown donor (Farrant et al., 1997). This hypothesis has been supported by the discovery of a new Salmonella gene, sodC-2, encoding a protein 82% identical to the E. coli SodC (Fang et al., 1999). The first sodC gene is now renamed sodC-1. Here we report an investigation of the contribution that each of these genes makes to the virulence of S. choleraesuis. Unexpectedly, we did not find the effect to be additive, but rather, that both genes were necessary for the contribution of [Cu,Zn]-SOD to be discerned.

### METHODS

#### Bacterial strains and culture.

S. choleraesuis strain A50 is a clinical veterinary isolate (Watson et al., 2000). A sodA mutant of S. typhimurium was kindly provided by Dr Anjam Khan (University of Newcastle). E. coli S17-1 ϕpir was used as the host for conjugative transfer of plasmid DNA to Salmonella (Simon et al., 1983). Salmonella strains were made resistant to nalidixic acid by standard procedures. Bacteria were grown inuria–Berti (LB) medium at 37 °C with shaking (130–180 r.p.m.). Where needed, antibiotics were added at: kanamycin, 50 mg l$^{-1}$; nalidixic acid, 30 mg l$^{-1}$; penicillin, 250 mg l$^{-1}$. Bacterial strains were stored in 10% (v/v) glycerol at −70 °C and fresh aliquots were used for each infection experiment.

#### Construction of sodC-2 mutants.

A 344 bp DNA fragment amplified by PCR from within the sodC-2 ORF (nt 508–851, GenBank accession AF056931) and ligated into the suicide vector pRR10(ΔTrfA) (Fang et al., 1992) was generously provided by Ferric Fang (University of Colorado). This plasmid was mobilized from E. coli S17-1 into S. choleraesuis A50 wild-type and sodC-1 mutant (Farrant et al., 1997). Transconjugants were selected on LB agar containing penicillin for the sodC-2 mutants or kanamycin and penicillin for the sodC-1 sodC-2 mutants. Homologous recombination of the plasmid into the Salmonella chromosome created an interruption of sodC-2, confirmed by Southern hybridization.

#### Extraction of bacterial proteins and detection of SOD activity.

Periplasmic extracts were prepared from 100 ml overnight cultures of S. choleraesuis wild-type, and sodC-1, sodC-2 and sodC-1 sodC-2 mutants by the method of Higgins & Hardie (1983). Proteins were precipitated with ammonium sulphate (650 g l$^{-1}$), concentrated by centrifugation and stored at −20 °C for later analysis. To detect SOD, proteins were separated in pre-cast wide range (pH 3–10) IEF gels (Bio-Rad). Gels were stained to reveal SOD activity using the method of Beauchamp & Fridovich (1971) as modified by Steinman (1985). [Cu,Zn]-SOD activity was identified by specific inhibition with 5 mM diethyldithiocarbamate (DEDCC; Sigma), a copper chelator (Benov & Fridovich, 1994). [Fe]-SOD was identified by inhibition of its activity with hydrogen peroxide (Crapo et al., 1978).

#### Bacterial sensitivity to superoxide and nitric oxide generated in vitro.

Sensitivity to increased cytosolic $O_2^-$ flux induced by paraquat was assessed by the method of De Groote et al. (1995). Sensitivity to exogenous $O_2^-$ and NO was assessed by exposure of suspensions (10$^8$ c.f.u. ml$^{-1}$) of stationary-phase organisms in phosphate-buffered saline (pH 7.4) (PBS) to these radicals, generated in solution respectively from reactions...
between xanthine (X) and xanthine oxidase (XO) (Fridovich, 1970) and on acidification of ice cold alkaline 2,2′-hydroxy-nitrosohydrozazono bis-ethanamine (spermine NONOate, SPER/NO) (De Groote et al., 1997). Bacterial suspensions in these solutions were incubated with shaking at 37 °C and aliquots were removed periodically and assayed for viable counts.

**Preparation of macrophages.** Porcine alveolar macrophages were isolated as previously described (Farrant et al., 1997). Macrophages were suspended in Iscove’s modified Dulbecco’s medium containing 10% (v/v) foetal calf serum (FCS), 100 mg gentamicin l−1 and 100 U ml−1 each of penicillin and streptomycin, and distributed into 1 ml flat-bottomed wells of tissue culture plates to give 5 × 10⁶ cells per well. Wells were incubated overnight at 37 °C in an atmosphere of 5% CO₂ to allow cells to adhere as monolayers. Two hours before infection monolayers were washed and covered with fresh antibiotic-free medium. Murine peritoneal macrophages were isolated from BALB/c mice at 4 days after intraperitoneal (i.p.) injection of 5 mM sodium periodate by peritoneal lavage. Macrophages were suspended in RPMI 1640 medium containing 10% FCS and 20 U murine interferon γ (IFNγ) ml−1, and distributed to 0-1 ml flat-bottomed wells at 5 × 10⁵ cells per well. Cells were left to adhere as monolayers as before, and washed and covered with fresh medium at the start of experiments.

**Infection of monolayers.** Bacteria from overnight cultures were washed and resuspended in tissue culture medium at a concentration of 5 × 10⁶ c.f.u. ml−1. For opsonization, bacteria were incubated with 10% normal porcine or murine serum at 37 °C for 30 min on a rolling platform at 120 r.p.m. Bacteria were added to monolayers to give a verified multiplicity of infection of 5–10 bacteria per macrophage. In experiments with IFNγ-stimulated macrophages, the infected monolayers were gently centrifuged (150 g for 10 min at 4 °C). Experiments were performed in triplicate.

**Bacterial uptake and killing by macrophages.** Bacterial uptake and killing were assessed by a modification of the gentamicin-protection assay of Buchmeier & Heffron (1989). Infected monolayers of IFNγ-stimulated macrophages were incubated for 15 min, followed by a wash and further incubation with Iscove’s modified Dulbecco’s Medium or RPMI 1640 containing 10% v/v FCS and 100 mg gentamicin l−1. After 1–24 h incubation, monolayers were washed twice with PBS to remove any remaining superficially adherent viable bacteria and lysed with 0.1% (w/v) sodium deoxycholate in PBS. To assess viable counts of phagocytosed bacteria, lysates were plated on to MacConkey agar containing 30 mg nalidixic acid l−1. For experiments with unstimulated macrophages, the initial incubation was extended to 1 h.

**Experimental infection of mice.** Mice were purchased from Charles River Laboratories. In a first experiment, groups of five 8–10 week-old BALB/c female mice were infected i.p. with 10⁶ c.f.u., and five C3H/HeN mice with 5 × 10⁸ c.f.u. A group of sham-inoculated mice received saline only. At 4 and 5 days post-infection, the mice began to exhibit symptoms of systemic disease and were killed by CO₂ asphyxiation. Spleens and livers were removed and number of viable organisms in the organs was determined as above.

Groups of three 8–10 week-old BALB/c female mice were infected by oral administration of 10⁶ c.f.u. Salmonella suspended in 0.1 ml 0.9% saline. Sham-infected mice received saline only. At 5 days post-infection, as mice began to exhibit symptoms of systemic disease, they were killed by CO₂ asphyxiation. Spleens and livers were removed and the number of viable organisms in the organs was determined as above.

**RESULTS**

**Characterization of mutants and growth characteristics**

The presence of sodC-1 and sodC-2 was established in _S. choleraesuis_ by Southern blotting (Farrant et al., 1997 and further data not shown) and the set of sodC-1 and sodC-2 mutants of strain A50 were constructed as described in Methods. Strains were examined for SOD activity. Two bands of SOD activity, sensitive to DEDC inhibition and so characteristic of [Cu,Zn]-SOD, were present in the wild-type but one or other band, as expected, was absent in the single mutant strains (Fig. 1). SodC-2 is more acidic (pI 5–6) than SodC-1 (pI ~ 8) and runs very close to a band of [Fe]-SOD activity, sensitive to hydrogen peroxide (H₂O₂ sensitivity data not shown). _S. choleraesuis_ A50 wild-type and mutant strains were grown aerobically in broth culture with shaking for 78 h, without replenishment of the LB medium. There was no significant difference in gen-

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**Fig. 1.** Paired IEF gels stained for SOD activity (a) without and (b) with selective inhibition of [Cu,Zn]-SODs by 5 mM DEDC. The pH range of the ampholytes was 10 (top) to 3 (bottom). Lanes 1–4 contain proteins extracted from (1) _S. choleraesuis_ A50 wild-type; (2) _S. choleraesuis_ sodC-1 mutant; (3) _S. choleraesuis_ sodC-2 mutant; and (4) _S. choleraesuis_ sodC-1 sodC-2 double mutant. Arrowheads mark the positions of SodC-1 (top), SodC-2 (middle) and iron-cofacted SOD (bottom).
result of the experiment (carried out in triplicate) is shown.

\[ \text{S} \] and PBS strains in PBS.

\[ \text{XO} \] and nitrogen species.

Effect of mutations on sensitivity to reactive oxygen and nitrogen species \textit{in vitro}

As expected, wild-type and sodC mutant strains were equally resistant to paraquat, a redox cycling reagent that penetrates the cytosol, greatly increasing superoxide production there. \textit{S. typhimurium} AsodA (lacking cytosolic Mn-cofactored SOD) was used as the control and was shown to be more sensitive (data not shown). \textit{S. choleraesuis} mutant strains were significantly \((P < 0.01)\) more susceptible than the wild-type to killing by extracellular superoxide generated by X/XO (Fig. 2). A dramatically increased sensitivity of the mutants was observed to the synergistic killing by X/XO and the NO donor (Fig. 2). SPER/NO by itself had no antimicrobial activity against either \textit{S. choleraesuis} wild-type or mutant strains. No additional difference in sensitivity was shown by the double mutant in any of these assays.

\[ \text{Cu,Zn} \]-SODs and interaction of \textit{Salmonella} with phagocytic cells \textit{in vitro}

\textit{S. choleraesuis} sodC mutant strains showed a significant increased susceptibility to killing by activated macrophages compared to the wild-type strain (Fig. 3). There was no difference in the susceptibility of the single mutants, or between either single and the double mutant.

Macrophage activation was necessary to elicit these differences. Using non-activated murine and porcine macrophages, prepared as described by Farrant et al. (1997), no differences were found between wild-type and sodC mutant strains in either bacterial uptake or subsequent bacterial killing, whether or not bacteria were opsonized (results not shown).

\[ \text{Cu,Zn} \]-SODs and pathogenesis of systemic disease

After i.p. infection, at the humane end point when mice began to show signs of systemic infection, the load of organisms in spleen and liver was measured as described. Whether recovered from \textit{Ity} or from \textit{Ity} mice, the wild-type strain was recovered in highly significantly greater numbers than the sodC mutant strains (Fig. 4). However, the double mutation did not attenuate any more than either single mutation. In all cases, the strains retained the expected antibiotic resistance, showing that no reversion of the sodC genotype had occurred. A similar result was obtained using lower i.p. inocula (100 c.f.u.) of bacteria (data not shown) and with groups of mice infected orally (Fig. 5). Once again the double mutation did not have any additional effect. We have previously demonstrated that complementation of one sodC mutation (sodC-1) with a plasmid-borne copy of the gene restored \textit{S. choleraesuis} to wild-type virulence in this model (Farrant et al., 1997).
Role of two [Cu,Zn]-SODs in *Salmonella* virulence

### Recovery of organisms [log c.f.u. (g organ)–1]

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Fig. 4. Effect of *sodC* mutations on the virulence of *S. choleraesuis* after i.p. challenge of (a) BALB/c (4 days after challenge) and (b) C3H/HeN mice (5 days after challenge). Bars show the recoveries of wild-type (wt, black bars), *sodC-1* (C1, vertical hatched bars), *sodC-2* (C2, horizontal hatched bars) and double (C1+C2, cross-hatched bars) mutant strains from spleen and liver. Results are expressed as c.f.u. recovered (mean ± SEM) from 5 mice per bacterial strain.

Fig. 5. Effect of *sodC* mutations on the virulence of *S. choleraesuis* in BALB/c mice 5 days after oral challenge. Bars show the recoveries of wild-type (wt, black bars), *sodC-1* (C1, vertical hatched bars), *sodC-2* (C2, horizontal hatched bars) and double (C1+C2, cross-hatched bars) mutant strains, and virulence plasmid-cured strain (P−, white bars) from spleen and liver. Results are expressed as c.f.u. recovered (mean ± SEM) from 3 mice per strain.

Fig. 6. Graphical representation of relationship between sequences of (a) *Salmonella* SodC-2 and other bacterial SodCs; (b) as (a), but including *Salmonella* SodC-1; (c) RecF from the same bacterial species.
Phylogenetic comparison of *Salmonella* SodC-1 and SodC-2 with SodC from other bacteria

The multiple sequence comparison programme UnrootedTree (at http://cbgr.inf.ethz.ch/Server/MultAlign.html) was used to analyse graphically the relationship between the published sequences of *Salmonella* SodCs and SodC from the disparate species *E. coli*, *Haemophilus influenzae*, *Actinobacillus pleuropneumoniae*, *Mycobacterium tuberculosis* and *C. crescentus* (Fig. 6). A similar analysis can be carried out for any set of homologous proteins, and for comparison is shown for the central metabolic (DNA repair) enzyme RecF from the same bacterial species (Fig. 6c). These phylogenetic trees are drawn on the principle that the length of the path joining loci representing protein sequences is in proportion to their degree of dissimilarity. Short and long distances thus signify, respectively, similar or divergent sequences. The striking degree of similarity between the RecF and SodC trees before *Salmonella* SodC-1 is taken into account (Fig. 6a compared to Fig. 6c) suggests that sodC and recF are not subject to different pressure of mutation in the different species analysed. *Salmonella* and *E. coli* protein sequences are generally very similar and the short distance separating the RecF sequence loci corresponds to that separating *Salmonella* SodC-2 and *E. coli* SodC. However, when the tree is reconstructed to include *Salmonella* SodC-1 (Fig. 6b), this branch diverges widely from both *Salmonella* SodC-2 and *E. coli* SodC.

**DISCUSSION**

Here we have shown that extracytoplasmic [Cu,Zn]-SODs in *Salmonella* protect organisms against exogenously generated O$_2^-$$. In addition we have demonstrated the contribution of SodC-1 and SodC-2 to the virulence of *S. choleraesuis* in an experimental infection model, establishing [Cu,Zn]-SOD as a determinant of *Salmonella* virulence, reducing the effectiveness of ROI- and RNI-mediated host defence mechanisms. While the two SodCs together are associated with full virulence of *S. choleraesuis*, our data show that their contributions are not additive. Inactivation of both sodC genes leads to no further attenuation in the resistance of *S. choleraesuis* to O$_2^-$ and NO generated in vitro, or in virulence, than is seen with either single mutant. This suggests that the contribution of [Cu,Zn]-SOD in these respects is conditional on there being a threshold level of enzyme activity. With two functional sodC genes, *S. choleraesuis* achieves enhanced survival under the conditions of oxidative stress encountered during experimental murine infection by the oral or parenteral route, reproduced ex vivo in experimental infection of activated macrophages. This observation finds a correlate in the observations of Battistoni et al. (2000). An *E. coli* strain engineered to increase [Cu,Zn]-SOD production was found to be much more resistant to intracellular killing than the wild-type strain, while there was very little difference in survival between the wild-type and a sodC-null mutant.

While a [Cu,Zn]-SOD gene has by now been found in many bacterial species, the discovery of two different, functional, versions of the gene – and very recently a third, sodC-3 (Figueroa-Bossi et al., 2001) – in the one pathogen is without precedent. The gene sodC-2 in *Salmonella* is the true orthologue of sodC in *E. coli*, established by extended sequencing of the chromosomal locus in each organism. In *E. coli* a 7 kb domain contains consecutive ORFs: *slyA*-B1643-B1644-B1645-sodC-B1647-B1648-B1649-nemaA. A search with the translation of each of these genes/putative genes against the *S. typhimurium* genome database (at http://genome.wustl.edu/gsc/Projects/S.typhimurium/) demonstrated in each case high identity (> 80 %) for corresponding ORFs flanking sodC-2. In contrast, sodC-1 and sodC-3 appear to have been acquired on lysogenic bacteriophages (Gifsy-2 and Fels-1 respectively) (Figueroa-Bossi & Bossi, 1999; Figueroa-Bossi et al., 2001). A likely origin of sodC-1 from outside *Salmonella* is revealed by all-against-all protein sequence comparison of a set of SodC sequences. SodC-1 is as divergent in sequence from *Salmonella* SodC-2 and *E. coli* SodC as the examples of the sodC from such upper respiratory tract pathogens as *H. influenzae*. While an anomalously high mutation rate could theoretically be the explanation for the SodC-1 sequence divergence, the association with phage genes strongly supports the hypothesis that this gene has an exogenous origin: sodC-1 joins a growing list of virulence genes likely to have been acquired through horizontal transfer. Carriage of sodC-1 on lysogenized Gifsy-2 is a characteristic of the more virulent *Salmonella* serotypes, such as *S. typhimurium*, *S. choleraesuis*, *S. dublin* and *Salmonella enteritidis*. The less virulent serotypes have only sodC-2 (Fang et al., 1999). Taken with the data presented here for *S. choleraesuis*, this suggests that selection pressure may have favoured acquisition of one or even two extra [Cu,Zn]-SOD genes by *Salmonella* serovars adapted to survive within macrophages. A mechanism to promote sodC-1 acquisition may be proposed. Gifsy-2 elements are released upon exposure to H$_2$O$_2$, raising the intriguing possibility that infection of macrophages and exposure to reactive oxygen species generated in the respiratory burst may, through induction of a lytic cycle for Gifsy-2, potentiate transmission of the genes it carries, among them protective sodC-1, to other salmonellae (Figueroa-Bossi et al., 1999).

The proposition that horizontally acquired [Cu,Zn]-SOD genes may make a special contribution to bacterial virulence, not conferred by endogenous sodC, gains support from observations in the genus *Neisseria*. Commensal neisserial species, common colonists of the human upper respiratory and genital mucosa, do not in general have sodC genes (our unpublished results) but *N. meningitidis*, the only neisserial species regularly causing invasive, life threatening, infection, is the exception and in this pathogen [Cu,Zn]-SOD unequivocally contributes to virulence (Wilks et al., 1998). As seen for sodC-1 in *Salmonella*, meningococcal sodC appears to have been acquired by horizontal transfer, in
this case probably from co-commensal \textit{Haemophilus} species (Kroll \textit{et al.}, 1998). In contrast, studies of other pathogens with only a single \textit{sodC} gene and in which there is no evidence, for example from sequence anomaly at the locus, of horizontal acquisition of the gene, have generally yielded equivocal or negative evidence for the involvement of [Cu,Zn]-SOD in virulence [\textit{B. abortus} (Tatum \textit{et al.}, 1992; Latimer \textit{et al.}, 1992), \textit{A. pleuropneumoniae} (Sheehan \textit{et al.}, 2000) and \textit{M. tuberculosis} (Dussurget et al., 2001; Piddington \textit{et al.}, 2001)].

Fang \textit{et al.} (1999) have studied the contribution that \textit{sodC}-1 and \textit{sodC}-2 make to the virulence of \textit{S. typhimurium} and reached a different conclusion to ours. It is not, however, possible directly to compare our findings as there are significant differences in experimental design. In a different experimental model – much more prolonged murine infection to a fatal end point – they observed that only the \textit{sodC}-1 \textit{sodC}-2 strain was attenuated in lethality for \textit{Ity} \textsuperscript{a} mice (a different strain to that used here). In an \textit{Ity} \textsuperscript{c} strain (the same as that used here), all \textit{sodC} mutants, single and double, were slower to kill mice than the wild-type. Although the result reported for the double mutant suggested greater attenuation, the reproducibility of this single observation cannot be assessed with the data available. It may be that the contrast reflects differences in the pathogenesis of \textit{S. typhimurium} and \textit{S. choleraesuis} in mice. Despite being highly virulent for pigs (Watson \textit{et al.}, 2000), \textit{S. choleraesuis} A50 is significantly less virulent for mice than \textit{S. typhimurium} and \textit{S. dublin} (Farrant \textit{et al.}, 1997).

In conclusion, in \textit{S. choleraesuis} it seems clear that the acquisition of an additional \textit{sodC} gene on a lysogenic bacteriophage has effectively led to a gain of function (plausibly through increased total activity) for [Cu,Zn]-SOD, contributing to virulence. Until now the contribution that \textit{sodC} makes to bacterial virulence has seemed wholly capricious – clearly contributing in some organisms, but not at all in others. From this work we would tentatively suggest that it is in particular in those organisms in which there is evidence of horizontal acquisition of \textit{sodC} – by bacteriophage or plasmid transfer, or through transformation with chromosomal DNA – that evidence may be found of a virulence-associated gain of function, the capacity to neutralize host defences that depend on the generation of superoxide and its further reaction products.

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