Nitrogen regulation of blue light-inducible genes in *Neurospora crassa*

VLADIMIR Y. SOKOLOVSKY,† FRANK-ROMAN LAUTER,‡§ BERND MÜLLER-RÖBER,§ MARTA RICCI,¹ THOMAS J. SCHMIDHAUSER² and VINCENZO E. A. RUSSO¹

¹Max-Planck-Institut für Molekulare Genetik, Abteilung Trautmann, Ihnestr. 73, 1000 Berlin-Dahlem 33, Germany
²Department of Chemistry and Biochemistry, Southern Illinois University Carbondale, Carbondale, Illinois 62901, USA

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Sexual and asexual differentiation of *Neurospora crassa* are influenced by nitrogen availability and blue light. Nitrogen limitation induces the production of protoperithecia on solid medium and conidia in liquid medium. Both developmental processes are stimulated by blue light. We have analysed changes in mRNA levels for a variety of light-inducible genes (*al, bli* and *con*) under conditions of nitrogen limitation. We show that the photoregulated genes *al-1, al-2, bli-4, bli-7, con-5* and *con-10* are also regulated by nitrogen limitation. These genes exhibited nitrogen regulation in the nonphotoresponsive mutant strains *wc-1* and *wc-2*. Therefore, the *wc-1* and *wc-2* gene products, although necessary for photoregulation of these *al, bli* and *con* genes, are not required for nitrogen regulation.

Introduction

During its life cycle, the heterothallic filamentous fungus *Neurospora crassa* undergoes asexual and sexual differentiation. Both developmental processes are influenced by external signals, primarily nutrient deprivation and illumination with blue light (Nelson et al., 1975; Turian, 1977; Degli-Innocenti & Russo, 1984a, b).

The vegetative growth of the fungus is characterized by the formation of mycelia composed of multinucleate, branched hyphae. In liquid shake cultures, nitrogen limitation activates the developmental process, resulting in production of conidia (Guignard et al., 1984; Müller & Russo, 1989). These semi-dormant spores represent the final products of asexual differentiation. This process of conidiation involves a series of discrete morphological stages which have been characterized in detail by scanning electron microscopy (Springer & Yanofsky, 1989). On solid medium, conidiation is induced by desiccation (Nelson et al., 1975) and glucose limitation (Ricci et al., 1991). Once induced, several events in this complex differentiation process are influenced by light, e.g. conidiophores develop towards light (Faull, 1930; Siegel et al., 1968), and illuminated cultures produce conidia faster than dark-grown cultures (Siegel et al., 1968; Turian, 1977).

On solid medium, nitrogen limitation leads to the differentiation of hyphae into the female organs, protoperithecia (Sommer et al., 1987; Ricci et al., 1991). During the sexual process these are fertilized by conidia or vegetative hyphae of the opposite mating type. Once induced by nitrogen deprivation, the formation of protoperithecia is greatly enhanced by blue light (Degli-Innocenti et al., 1983; Sommer et al., 1987). In dark-grown mycelia about 10 protoperithecia form per cm²; this increases to about 300 per cm² after blue light illumination (Sommer et al., 1987).

Recently, two sets of genes have been identified which are of interest with respect to the developmental programs in *N. crassa*. One group, the *con* genes, was isolated on the basis of preferential expression during conidiation (Berlin & Yanofsky, 1985). The functions of these genes are unknown. The second group comprises blue-light-inducible (*bli*) genes; mRNAs corresponding to these genes increase after illumination with blue light (Sommer et al., 1989). The functions of the *bli* genes, and their expression pattern during conidiation are also unknown. Interestingly, some of the *con* genes respond to both light and conidiation (Lauter & Russo, 1991). Two additional genes that are also both light-inducible and expressed during conidiation, albino-1 (*al-l*) (Schmidhauser et al., 1990; T. J. Schmidhauser, M. S. Sachs & C. Yanofsky, unpublished data) and albino-2 (*al-2*) (T. J. Schmidhauser, F.-R. Lauter, V. E. A. Russo & C. Yanofsky, unpublished data), were used in this analysis.
The \textit{al} genes are necessary for carotenoid production (Harding & Turner, 1981). \textit{al-1} encodes the enzyme phytoene dehydrogenase (Schmidhauser et al., 1990) and \textit{al-2}, phytoene synthase (T. J. Schmidhauser, F.-R. Lauter, V. E. A. Russo & C. Yanofsky, unpublished results) of \textit{N. crassa}. Although they are developmentally regulated, expression of these genes is not necessary for asexual and sexual development (Degli-Innocenti & Russo, 1984\textit{a}; Russo, 1986).

Two regulatory mutants of \textit{N. crassa}, \textit{wc-1} and \textit{wc-2}, are impaired in light regulation. The products of \textit{wc-1} and \textit{wc-2} are required for all photoeffects tested so far (Harding & Turner, 1981; Harding & Melles, 1984; Degli-Innocenti & Russo, 1984\textit{a}, \textit{b}; Russo, 1988; Sommer et al., 1989; Nelson et al., 1989; Schmidhauser et al., 1990; Nawrath & Russo, 1990; Lauter & Russo, 1991), with the exception of photophosphorylation of proteins (Lauter & Russo, 1990). \textit{Wc-1} and \textit{Wc-2} are thought to be regulatory factors responsible for the light transduction pathway.

We report here that the light-regulated genes \textit{al-1}, \textit{al-2}, \textit{bli-4}, \textit{bli-7}, \textit{con-5} and \textit{con-10} are also regulated by nitrogen limitation. Transcript accumulation of those genes is increased upon nitrogen limitation in wild-type as well as \textit{wc-1} and \textit{wc-2} mutant strains.

\section*{Methods}

\textit{Strains and plasmids}. \textit{N. crassa} wild-type (WT) strain (St. Lawrence, STa) was provided by J. R. S. Fincham (Cambridge University, UK). Strains FGSC 4398 (\textit{wc-1}, \textit{a}), FGSC 4396 (\textit{wc-1}, \textit{a}), FGSC 4408 (\textit{wc-2}, \textit{a}) and R251 (\textit{wc-2}, \textit{a}) were used as sources of \textit{wc} RNA. The \textit{wc} strains were isolated after UV mutagenesis and are isogenic to the WT (STa) (Degli-Innocenti & Russo, 1984\textit{a}). The \textit{con} clones were developed by Charles Yanofsky (Stanford University) (Berlin & Yanofsky, 1985; Roberts et al., 1988). The \textit{al-1} and \textit{bl} genes were described previously (Schmidhauser et al., 1990; Sommer et al., 1989). In case of \textit{al-2}, an 800 bp cDNA fragment cloned in pTJS430 was used as hybridization probe.

\textit{Growth of} \textit{N. crassa} \textit{in liquid culture medium}. \textit{N. crassa} mycelia were grown in liquid culture, with shaking, as described by Chambers et al. (1985). Mycelia were grown in darkness in 75 ml medium in 250 ml flasks at 34 °C for 24 h prior to harvest. Modified Vogel's minimal medium (Russo, 1988) supplemented with 2% (w/v) glucose and 50 mm-NH\textsubscript{4}Cl as carbon and nitrogen sources, respectively, was used for cultivation. In nitrogen limitation experiments, mycelia were grown in the dark for 24 h in a medium supplemented with 2 mm-NH\textsubscript{4}Cl. Mycelia were harvested under red safety light by filtration (Chambers et al., 1985) and immediately frozen in liquid nitrogen.

\textit{Culture of} \textit{N. crassa} \textit{on solid medium}. \textit{N. crassa} was grown in 8 cm diameter Petri dishes containing 20 ml agar medium. The agar medium was modified Vogel's minimal medium (Russo, 1988) supplemented with 1.5\% (w/v) Bacto-Agar (Difco), with 1\% (w/v) L-sorbosine, and 0.1\% glucose as carbon sources, with 4 mm-NH\textsubscript{4}Cl as the sole nitrogen source. Solid medium was covered with a circular dialysis membrane (Visking, Serva). For inoculation, 10\textsuperscript{4} conidia per plate were spread onto the dialysis membrane. Mycelia were grown in darkness at 23 °C. After 3 d, membranes overgrown with mycelia were transferred onto new plates that contained zero or 50 mm-NH\textsubscript{4}Cl as sole nitrogen source. After 24 h additional growth in darkness at 23 °C, cells were harvested by scraping them from the upper surface of the dialysis membranes. All manipulations were performed under a red safety light. Collected mycelia were immediately frozen in liquid nitrogen.

\textit{RNA dot blot analysis}. Extraction of total RNA was performed as described previously (Sokolovsky et al., 1990), an extraction method developed for RNAase-rich cultures. RNA dot-blot were performed as described by Boll et al. (1986). Radioactively labelled DNA probes were prepared using random primer labelling technique (Feinberg & Vogelstein, 1983). Quantification of mRNA was done as described previously (Sommer et al., 1989).

\section*{Results}

\textit{Nitrogen regulation of light-inducible genes in mycelia grown in liquid and on solid medium}

We wished to determine the expression patterns of light-regulated genes in response to nitrogen availability, a stimulus known to influence development. Nitrogen limitation induces conidiation in liquid media (Guignard et al., 1984, Müller & Russo, 1989), and both inhibits conidiation (Ricci et al., 1991) and induces protoperithecia formation (Sommer et al., 1987, Ricci et al., 1991) on solid media. Since different developmental processes may be induced by the same stimulus, depending on growth conditions, we have compared the effects of nitrogen limitation on the transcription patterns of photoregulated genes in mycelia grown in liquid culture and on solid agar. Cultures were grown in darkness under conditions of low or high nitrogen supply, and the amount of mRNA expressed from different genes was measured by dot-blot analysis (Boll et al., 1986). Our results, based on RNA extracted from three to five different mycelial cultures grown in liquid medium and two different mycelial cultures grown on solid medium, are compared in Table 1.

Under conditions of nitrogen limitation, mRNA levels for all the photoregulated genes except \textit{bli-3} increased by more than a factor of 3 in wild-type cultures grown in liquid medium. The slight increase in transcript levels of \textit{bli-3} was not considered significant. A greater than 10-fold increase in accumulation was observed for the \textit{al-1} and \textit{bli-7} RNAs. The increase in mRNA level for \textit{bli-7}, a gene which is weakly photoregulated (Sommer et al., 1989), was appreciable; a greater than 50-fold accumulation was observed. In the induced state, the \textit{bli-7} mRNA represented about 0.25\% of the total RNA (Table 1). Under conditions of nitrogen limitation, the variation in \textit{bli-7} transcript levels among different experiments was fairly low (less than 10\%). However, fluctuation of \textit{bli-7} transcript levels has been observed in mycelia grown in complete medium (Tables 1 and 2;
was especially significant in the case of limitation in mycelia grown on solid medium. This effect is considered significant. Interestingly, transcription of and liquid medium, transcript levels of slightly increased in cultures shifted to a nitrogen-free medium; the expression of these genes was also analyzed in mycelia grown on solid agar. As in under nitrogen control in liquid medium (Table 1); an induction factor of more than 10 was observed.

In two independent experiments, nitrogen regulation was also analyzed in mycelia grown on solid agar. As in liquid medium, transcript levels of and increased under conditions of nitrogen limitation (Table 1). The induction factors were comparably high under both growth conditions. In case of and con-10, the variation between the two experiments was too high to infer whether or not the mRNA levels of these genes increased under conditions of nitrogen limitation. As in liquid medium, transcript levels of tub-2 increased slightly in cultures shifted to a nitrogen-free medium; the slight increase in mRNA levels of was not considered significant. Interestingly, transcription of and , although strongly nitrogen-regulated in mycelia grown in liquid cultures, was uninducible by nitrogen limitation in mycelia grown on solid medium. This effect was especially significant in the case of . Under conditions of nitrogen limitation, mRNA, measured in cultures grown on solid agar medium, represented only 1–2 % of the amount measured in cultures grown in liquid medium.

Role of wc genes on nitrogen induced gene expression

Nitrogen regulation of photoinducible genes was also examined in wc strains. The wc mutants are blind for most of the photoeffects tested so far. Transcripts of the photoregulated al (Schmidhauser et al., 1990; Nelson et al., 1989; F.-R. Lauter, T. J. Schmidhauser, C. Yanofsky & V. E. A. Russo, unpublished results), bli (Sommer et al., 1989) and con genes (Lauter & Russo, 1991) do not increase after light stimulation in wc-1 and wc-2 strains.

In order to study nitrogen regulation in these mutants, the fungus was grown in darkness in liquid medium with low or high nitrogen supply. Two alleles each of wc-1 and wc-2 were used. Results are presented in Table 2. The response of al-1, al-2, bli-3, bli-4, bli-7 and tub-2 to nitrogen limitation was similar to WT in the wc mutant strains. RNA accumulation for con-5, con-10 and was slightly altered in wc strains following nitrogen limitation relative to wild-type, but still nitrogen-regulated. The products of the wc-1 and wc-2 genes do not appear to be required for nitrogen regulation of the genes analyzed in these studies.

**Table 1. Gene regulation in response to nitrogen limitation using liquid and solid growth media**

<table>
<thead>
<tr>
<th>Gene</th>
<th>Liquid medium</th>
<th>Solid medium</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>N+</td>
<td>N-</td>
</tr>
<tr>
<td>al-1</td>
<td>4.5</td>
<td>63.7</td>
</tr>
<tr>
<td>al-2</td>
<td>1.2</td>
<td>8.6</td>
</tr>
<tr>
<td>bli-3</td>
<td>11.4</td>
<td>14.7</td>
</tr>
<tr>
<td>bli-4</td>
<td>1.2</td>
<td>4.1</td>
</tr>
<tr>
<td>bli-7</td>
<td>52.1</td>
<td>269.2</td>
</tr>
<tr>
<td>com-5</td>
<td>11.4</td>
<td>47.2</td>
</tr>
<tr>
<td>con-5</td>
<td>0.6</td>
<td>2.7</td>
</tr>
<tr>
<td>am</td>
<td>11.9</td>
<td>190.0</td>
</tr>
<tr>
<td>tub-2</td>
<td>3.7</td>
<td>8.5</td>
</tr>
</tbody>
</table>

Sommer et al., 1989). Two nonphotoregulated genes; tub-2, which encodes β-tubulin, a subunit of cytoskeleton microtubules (Orbach et al., 1986), and am, which encodes NADP-specific glutamate dehydrogenase (GDH) of N. crassa were included for comparison. GDH is an enzyme of the nitrogen assimilatory pathway and converts ammonia to glutamate (Kinnard et al., 1982). Expression of tub-2 was slightly increased under conditions of nitrogen limitation. Transcription of am was under nitrogen control in liquid medium (Table 1); an induction factor greater than 10 was observed.

In two independent experiments, nitrogen regulation was also analyzed in mycelia grown on solid agar. As in liquid medium, transcript levels of al-2 and bli-4 increased under conditions of nitrogen limitation (Table 1). The induction factors were comparably high under both growth conditions. In case of al-1, con-5 and con-10, the variation between the two experiments was too high to infer whether or not the mRNA levels of those genes increased under conditions of nitrogen limitation. As in liquid medium, transcript levels of tub-2 increased slightly in cultures shifted to a nitrogen-free medium; the slight increase in mRNA levels of bli-3 was not considered significant. Interestingly, transcription of am and bli-7, although strongly nitrogen-regulated in mycelia grown in liquid cultures, was uninducible by nitrogen limitation in mycelia grown on solid medium. This effect was especially significant in the case of bli-7. Under conditions of nitrogen limitation, bli-7 mRNA, measured in cultures grown on solid agar medium, represented only 1–2 % of the amount measured in cultures grown in liquid medium.

**Discussion**

Sexual as well as asexual differentiation of N. crassa is induced by nitrogen limitation (Nelson et al., 1975; Guignard et al., 1984; Sommer et al., 1987; Müller & Russo, 1989; Ricci et al., 1991). Both developmental processes are also influenced by light (Faull, 1930; Sargent et al., 1966; Sargent & Briggs, 1967; Siegel et al., 1968; Turian, 1977; Degli-Inocenti et al., 1983; Sommer et al., 1987). We have analyzed transcript levels of blue-light-inducible genes under conditions of nitrogen limitation. In cultures grown in liquid medium the amounts of al-1, al-2, bli-4, bli-7, con-5 and con-10 mRNAs were also regulated by nitrogen availability in the medium. Therefore, these genes are under dual environmental control, being regulated by either nitrogen or light.

In addition, two nonphotoregulated genes, tub-2 and am, were analyzed. tub-2 RNA levels were independent of the nitrogen supply, but am transcripts increased by a factor of more than 10 when nitrogen was limiting. NADP-specific GDH activity (Dantzig et al., 1979) and am mRNA levels are known to be under nitrogen control (Frederick & Kinsey, 1990).
Table 2. Gene regulation under nitrogen limitation: wild-type vs. wc-1 and wc-2 mutant strains

Total RNA was isolated from N. crassa cultures dark-grown in liquid medium with high or low nitrogen supply (see Methods). Accumulation of specified transcripts was monitored by dot-blot analysis and quantified by densitometry. Values are means (±SEM) from 3–5 independent extractions per growth condition and strain; ratios were calculated for each high/low N pair and mean ratios then calculated. All data were normalized to n-6 (Sommer et al., 1989), a nitrogen-independent gene (not shown). Two alleles of each wc locus were used. R251 is a wc-2 strain isolated in the Russo laboratory after UV mutagenesis, isogenic to the wild-type STa.

<table>
<thead>
<tr>
<th>Gene</th>
<th>WT</th>
<th>wc-1</th>
<th>wc-1</th>
<th>wc-2</th>
<th>wc-2</th>
</tr>
</thead>
<tbody>
<tr>
<td>al-1</td>
<td>20.8 (±5.8)</td>
<td>25.6 (±7.8)</td>
<td>35.1 (±7.4)</td>
<td>48.3 (±15.0)</td>
<td>20.0 (±9.5)</td>
</tr>
<tr>
<td>al-2</td>
<td>7.5 (±2.6)</td>
<td>5.1 (±0.8)</td>
<td>12.1 (±2.7)</td>
<td>9.3 (±3.9)</td>
<td>7.1 (±1.2)</td>
</tr>
<tr>
<td>bli-3</td>
<td>1.9 (±0.7)</td>
<td>1.9 (±0.8)</td>
<td>1.1 (±0.5)</td>
<td>1.8 (±0.7)</td>
<td>1.1 (±0.3)</td>
</tr>
<tr>
<td>bli-4</td>
<td>4.1 (±0.9)</td>
<td>2.5 (±0.5)</td>
<td>5.0 (±1.1)</td>
<td>3.7 (±0.8)</td>
<td>2.3 (±0.6)</td>
</tr>
<tr>
<td>bli-7</td>
<td>176.5 (±87.2)</td>
<td>80.4 (±23.4)</td>
<td>153.0 (±91.3)</td>
<td>85.0 (±19.1)</td>
<td>65.0 (±24.6)</td>
</tr>
<tr>
<td>con-5</td>
<td>4.1 (±0.4)</td>
<td>2.0 (±0.4)</td>
<td>3.0 (±0.2)</td>
<td>2.0 (±0.4)</td>
<td>1.8 (±0.6)</td>
</tr>
<tr>
<td>con-10</td>
<td>4.2 (±1.2)</td>
<td>8.7 (±1.3)</td>
<td>9.2 (±1.1)</td>
<td>7.7 (±1.1)</td>
<td>6.1 (±2.9)</td>
</tr>
<tr>
<td>am</td>
<td>17.8 (±40)</td>
<td>5.1 (±0.8)</td>
<td>20.1 (±7.3)</td>
<td>58 (±1.6)</td>
<td>7.3 (±2.6)</td>
</tr>
<tr>
<td>tub-2</td>
<td>2.1 (±0.6)</td>
<td>1.0 (±0.4)</td>
<td>1.9 (±0.5)</td>
<td>1.3 (±0.3)</td>
<td>1.3 (±0.2)</td>
</tr>
</tbody>
</table>

WT, wild-type.

shaking liquid medium, submerged, or on solid agar. In liquid medium, nitrogen and glucose starvation leads to conidiation (Guignard et al., 1984; Müller & Russo, 1989), and no protoperithecia. On solid agar, nitrogen limitation induces protoperithecia formation (Sommer et al., 1987) and suppresses conidiation, while glucose limitation induces conidiation and suppresses protoperithecia formation (Ricci et al., 1991). In our experiments, despite their limited number and the variation between them, it was apparent that the expression of bli-7 and am differed significantly in cultures grown in liquid and on solid medium.

RNA levels for bli-7 are strongly nitrogen-regulated in liquid medium but unregulated on solid agar. This implies that bli-7 is regulated by at least three parameters: light, nitrogen availability, and the method of cultivation (liquid versus solid medium). Gene inactivation experiments should show whether the changes in bli-7 mRNA levels were accidentally parallel to the morphological transformations associated with conidiation or whether the product of this gene is essential for normal asexual differentiation. The amount of bli-7 mRNA in cells grown in liquid medium under conditions of nitrogen limitation was 0.25% of the total RNA. Assuming that bli-7 mRNA is polyadenylated, that poly (A+) mRNA represents 1–2% of the total RNA of N. crassa (Sachs & Yanofsky, 1991) and that the relative amounts of rRNA and tRNA do not change during nitrogen starvation, the amount of bli-7 mRNA can be estimated to be 10–25% of the total poly (A+) mRNA mass.

Why is bli-7 nitrogen-regulated in N. crassa grown in liquid but not on solid medium? One important difference between liquid and solid medium is the amounts of O2 and CO2 that are available to the organism. It is known that CO2 inhibits conidiation in N. crassa (Charles, 1962; Sargent & Kaldenborn, 1972) and differentiation of the fungus Phycomyces (Russo et al., 1981).

How is gene regulation by light and nitrogen achieved in N. crassa? Which elements participate in these regulatory processes? Two genes, wc-1 and wc-2, are likely to play an important role in the blue-light signal transduction chain of N. crassa (Harding & Turner, 1981; Degli-Innocenti & Russo, 1984b) and to influence proper dephosphorylation of proteins (Lauter & Russo, 1990). Their gene products are necessary for photoscarotenogenesis (Harding & Turner, 1981), phototropism of perithelial beaks (Harding & Melles, 1984), photoinduction of protoperithecia (Degli-Innocenti & Russo, 1984a) and photoinduction of circadian rhythms in bd mutants (Russo, 1988). In mycelia of wc-1 and wc-2 mutant strains, transcription of al, bli and con genes (Sommer et al., 1989; Nelson et al., 1989; Schmidhauser et al., 1990; Lauter & Russo, 1991) is not photoinducible.

The results presented here show that nitrogen regulation of al-1, al-2, bli-4, bli-7, con-5 and con-10 was not dramatically altered in wc mutant strains. Therefore, the products of wc-1 and wc-2, while absolutely necessary for light regulation of al, bli and con genes, are not needed for their nitrogen regulation.

RNA accumulation of al-1, al-2, bli-4, bli-7, con-5 and con-10 is regulated by two environmental factors that also influence development of N. crassa: nitrogen availability and blue light. Gene inactivation experiments should show whether some of the bli or con genes...
are needed for sexual or asexual differentiation processes in this fungus. Deletion analyses and study of protein–DNA interactions should lead to the identification of cis and trans acting elements that are involved in dual environmental control of those genes.

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References


