Wall mannoproteins in cells from colonial phenotypic variants of *Candida albicans*

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*Candida albicans* ATCC 26555 switched at high frequency ($10^{-1}$ to $10^{-3}$) between several phenotypes identified by colony morphology on a defined mineral amino-acid-containing agar medium supplemented with arginine and zinc (LAZ medium). When cells taken from colonies exhibiting distinct morphologies were plated directly onto LAZ agar, spontaneous conversion to all the variant phenotypes occurred at combined frequencies of $2.1 \times 10^{-1}$ to $9.5 \times 10^{-3}$. However, when cells taken from the different colonial phenotypes were plated directly onto an undefined medium (yeast extract/peptone/dextrose; YPD medium), or first incubated in liquid YPD medium and then cloned on YPD agar, all colonies observed exhibited the same phenotype (smooth-white). When cells from the smooth-white colonies were plated as clones on LAZ agar, the original switch phenotype reappeared. These results suggest that environmental conditions such as the growth medium (and possibly the temperature) influence switching by suppressing phenotype expression, but have no effect on genotype. The variant colony morphologies also appeared to be associated with differences in the relative proportions of yeast and mycelial cells. Zymolyase digests of wall preparations obtained from cells belonging to different colonial phenotypes were analysed by SDS-PAGE. After blotting to nitrocellulose paper, the mannoproteins were stained with Concanavalin A, with a polyclonal antiserum enriched in antibodies against mycelium-specific wall components, and with a monoclonal antibody raised against a high-molecular-mass mannoprotein band (260 kDa) specific to the walls of mycelial cells. The results suggest that phenotypic switching might be associated with changes in the degree of glycosylation in high-molecular-mass mannoproteins, or in the way these mannoproteins are bound to other cell wall components.

**Introduction**

The pathogenic character of the dimorphic fungus *Candida albicans* is thought to be associated with its capacity to grow by production of filamentous forms, as adhesion and penetration of the host cells seem to correlate with this morphology. However, this property alone seems to be insufficient to explain the ability of *Candida* to (i) infect different animal tissues, (ii) invade distinct immunocompromised hosts, (iii) resist the action of antifungal drugs, and (iv) evade the host immune system (Bodey & Fainstein, 1985; Chandler, 1985; Odds, 1988; Rogers & Balish, 1980; Soll *et al.*, 1987; Tapper-Jones *et al.*, 1981).

It has been reported that most strains of *C. albicans*, and some strains of *C. tropicalis*, switch at a high frequency, both *in vivo* and *in vitro*, between different phenotypes, distinguishable by colony morphology on agar (Pomès *et al.*, 1987; Slutsky *et al.*, 1985, 1987; Soll *et al.*, 1987). At least three distinct switching systems (the 3153A strain switching system, the WO-1 strain transition system, and the smooth-white–heavy-myceliated transition), which are strain-specific and give rise to a limited number of colony phenotypes, have been described in *C. albicans* (Slutsky *et al.*, 1985, 1987; Soll *et al.*, 1987). It has been suggested that switching may modulate the pathogenic character of *C. albicans* by conferring on the fungal cells the plasticity to avoid the host immune system and to develop resistance to

**Abbreviations:** ConA, concanavalin A; W, wrinkled; SW, smooth-white; ST, star; P, smooth-white-petite; HM, heavy-myceliated; R, ring; F, fuzzy; FR, flat-ring; SR, stippled-ring; HW, hyper-wrinkled; ISWW, intermediate smooth-white-wrinkled.
antifungal drugs (Slutsky et al., 1985). Thus, not only can more than one strain colonize the same individual in some cases (Odds, 1987), but also different phenotypes of a single strain may be found at the same site of infection (Odds, 1987; Soll et al., 1987). In addition, it has recently been reported that a single C. albicans strain, which was responsible for successive episodes of recurrent vaginitis, switched colony phenotypes between episodes (Soll et al., 1989). Significant differences have been reported in the adhesion characteristics as well as cell surface properties (i.e. ultrastructure and antigenicity) between the so-called 'white' and 'opaque' cell phenotypes in the white-adhesion characteristics as well as cell surface properties (i.e. ultrastructure and antigenicity) between the so-called 'white' and 'opaque' cell phenotypes in the white-systems (the WO-1 strain) described in C. albicans (Slutsky et al., 1987). These observations are of great interest, since adherence of fungal cells to the host tissues seems to play a transient but essential role in colonization.

Wall mannoproteins are the main antigenic components of the C. albicans surface (Casanova et al., 1989; Chaffin et al., 1988; Poulain et al., 1985; Sundstrom et al., 1988), and are also implicated in the attachment of C. albicans to epithelial cells and to inert surfaces (Douglas, 1987). Consequently, in the present study we have (i) examined the switching capabilities of C. albicans ATCC 26555, a common laboratory strain, under different growth conditions, and (ii) monitored the mannoprotein antigens present in the cell wall of the switch phenotypes. The results of this study suggest that phenotypic switching may be accompanied by changes in the pattern of cell-wall-bound antigenic mannoproteins.

Methods

Organism and maintenance of stock cultures. Stock cultures of C. albicans ATCC 26555 were maintained by subculturing every 2-3 weeks on 1-5% (w/v) Bacto-Agar slopes of Sabouraud dextrose medium.

Colonies exhibiting specific morphologies were resuspended into 125 ml Erlenmeyer flasks containing either 25 ml LAZ medium or 25 ml YPD medium, and incubated with shaking (200 r.p.m.) at 28 °C for 10-12 h. Cells from these cultures were suspended in double-distilled water, counted, and plated (approx. 200 cells per plate) on LAZ and/or YPD agar. These plates were then incubated for 5-7 d at 28 °C or 37 °C before examination of colony morphology. The experimental protocols followed in this case are summarized in Tables 2 and 3.

Cell wall preparation. Colonies exhibiting specific morphologies were removed from the surface of the plates, and resuspended independently into chilled 1 mM-phenylmethylsulphonyl fluoride (PMSF) in 0-01 M-Tris/HCl buffer, pH 7-4 (buffer A). Cells were washed three times with buffer A by centrifugation (3000 g for 10-15 min), and broken by vortexing with glass beads (0-45-0.5 mm in diameter; 5 mg beads per mg wet weight of cells). Cell walls were sedimented (1200 g, 10 min) from the cell-free homogenates, and purified by several washes with buffer A as previously described (Casanova et al., 1989). Isolated walls were freeze-dried, and stored at -20 °C until further use.

Solubilization of wall antigens by enzymic digestion. Mannoproteins were solubilized from purified walls by treatment with Zymolyase 20T (a β-glucanase complex; 10 μg of enzyme complex per 100-150 μg of purified freeze-dried cell walls, in 1 mM-sodium azide and 1 mM-PMSF in 0-01 M-Tris/HCl buffer, pH 7-4, at 28 °C for 2 h) (Casanova et al., 1989). After treatment, the wall residue was removed by centrifugation (1200 g, 15 min), and the solubilized material was concentrated by freeze-drying. The total sugar and protein content in the lyophilized material was determined by the method of Dubois et al. (1956) and by the Lowry method, respectively.

PAGE and Western blot techniques. Mannoproteins solubilized after enzymic digestion of the walls were separated by SDS-PAGE, performed basically as described by Laemmli (1970) with the modifications as previously reported (Casanova et al., 1989). Electrophoretic transfer (Western blot) to nitrocellulose paper (Bio-Rad) was performed as described by Burnette (1981) except that the transfer buffer used contained 0-025 M-Tris/0-192 M-glycine (pH 8-3), and 20% (v/v) methanol, and the electrophoretic transfer was carried out at 6-8 V cm⁻¹ for 15-17 h at 4 °C in Trans-Blot cell (Bio-Rad) using an LKB 2197 power supply. Immunochemical detection of proteins on blots was done by the method described in the Bio-Rad Immun-Blot (GAR-HRP) assay kit, based on the procedure of Burnette (1981) and Towbin et al. (1979). A polyclonal rabbit serum enriched in antibodies against the overall mannoprotein composition of purified mycelial walls (mPAb), and a mouse monoclonal antibody (mAb 4C12) raised against a mannoprotein band with an average apparent molecular mass of 260 kDa, specific for the cell wall of the mycelial morphology of C. albicans ATCC 26555, were prepared as previously reported (Casanova et al., 1989) and used as antibody probes. Antibodies were used at a final concentration of 1:500 (for mPAb) or 1:3000 (for mAb) in 0-01 M-Tris/HCl buffer, pH 7-4, containing 3% (w/v) bovine serum albumin as blocking agent, 0-9% NaCl and 0-05% Tween 20. Diluted (1:2000) peroxidase-labelled goat anti-rabbit, or goat anti-mouse IgG (Bio-Rad) were used as indicator antibodies with 4-chloro-1-naphthol as the chromogenic reagent. Concanavalin A (ConA) staining of nitrocellulose blots was done according to Hawkes (1982) with the modifications of Millette & Scott (1994).

Chemicals. Gel electrophoresis and blotting reagents were from Bio-Rad. Zymolyase 20T was from Miles Laboratories. SDS molecular mass markers were from Sigma and Pharmacia. Culture media compounds were purchased from Difco. All other chemicals used were purchased from Sigma.
Results

Frequency of colony phenotypes after plating of cells of the original strain

When cells of a stock culture of a standard strain of *C. albicans* (ATCC 26555; this strain had been maintained in our laboratory for over five years by repeated subculturing on slopes of Sabouraud-dextrose medium) were plated as clones on plates of LAZ medium, variant colony morphologies appeared spontaneously at frequencies from $4.6 \times 10^{-1}$ to $8.4 \times 10^{-3}$ (Tables 1 and 2). Since the colony morphologies were (Fig. 1), in most cases, closely similar to those of the colonial phenotypes described by Slutsky *et al.* (1985) in *C. albicans* 3153A, we have used, where possible, the terminology proposed by these authors to describe our colony variants. Of the total number of primary colonies examined after the initial plating (Table 1), 2139 (64.6%) were smooth-white (SW; Fig 1b), 1126 (34.04%) were wrinkled (W; Fig. 1a), 28 (0.84%) were ring (R; Fig. 1e), and 14 (0.42%) were star (ST; Fig. 1c). Different figures were obtained in another plating on LAZ medium, when the variety of switching phenotypes detected was greater (Table 2). Thus, from the total number of colonies scored, 1147 (40.6%) were smooth-white, 387 (13.7%) were stippled-ring (SR; Fig. 1h), 352 (12.3%) were star, 286 (10.1%) were wrinkled, 256 (9.0%) were flat-ring (FR; Fig. 1g) [this colonial variant was somewhat different from the colony morphologies mentioned by Slutsky *et al.* (1985), so we coined the

<table>
<thead>
<tr>
<th>Original source of sample*</th>
<th>No. of primary colonial phenotypes obtained</th>
<th>No. of primary colonies cloned and their phenotype†</th>
<th>No. of secondary colonial phenotypes appearing on LAZ agar</th>
<th>No. of secondary colonies cloned and their phenotype†</th>
<th>No. of tertiary colonial phenotypes appearing and predominant phenotype obtained after plating of cells from the secondary W or SW colonies on LAZ or YPD agar</th>
<th>No. of quaternary colonial phenotypes appearing and predominant phenotype obtained after plating of cells from the tertiary W or distinct SW colonies on LAZ or YPD agar</th>
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<tbody>
<tr>
<td></td>
<td>2139 smooth wt (SW)</td>
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<td>1992 W</td>
<td>1 W</td>
<td>1647 W</td>
<td>1647 W</td>
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<tr>
<td></td>
<td>1126 wrinkled (W)</td>
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<td>1658 SW</td>
<td>1 SW</td>
<td>1914 SW</td>
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<td></td>
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<td>1457 SW</td>
<td>1457 SW</td>
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</tbody>
</table>

* Cells from a 24–48 h culture of *C. albicans* ATCC 26555 on Sabouraud-dextrose slope were suspended in double glass-distilled water and plated (500–1000 c.f.u. per plate) on the defined medium of Lee *et al.* (1975), containing zinc and arginine (LAZ medium; see Methods). Plates were incubated at 28 °C for 5–7 d prior to assessment of colony phenotype.

† Cells from the respective primary, secondary and tertiary colonial morphologies were resuspended in sterile glass-distilled water, counted and plated (approx. 200 c.f.u. per plate) on the indicated growth media. Plates were incubated at 28 °C for 5–7 d.
Table 2. Colony phenotypes obtained after primary plating of cells from the original sample followed by secondary cloning of cells removed from the individual primary colonial phenotypes, and effect of preincubation and subsequent plating of the cells in/on different growth media

Original source of sample: *C. albicans* ATCC 26555 (see footnote in Table 1). See Abbreviations for phenotypes.

<table>
<thead>
<tr>
<th>No. of primary colonial phenotypes obtained</th>
<th>No. of primary colonies cloned and phenotype*</th>
<th>No. of secondary colonial phenotypes appearing and predominant phenotype obtained</th>
<th>No. of secondary colonies exhibiting phenotypes other than that of the predominant secondary phenotype</th>
<th>Combined frequency of switching</th>
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<tr>
<td></td>
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<td>Preincubation and secondary plating in/on:</td>
<td>Preincubation and secondary plating in/on:</td>
<td>Preincubation and secondary plating in/on:</td>
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<tr>
<td></td>
<td></td>
<td>LAZ</td>
<td>YPD</td>
<td>LAZ</td>
</tr>
<tr>
<td>1147 SW</td>
<td>1 SW</td>
<td>2500 SW</td>
<td>3743 SW</td>
<td>538</td>
</tr>
<tr>
<td>387 SR</td>
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<td>2008 SR</td>
<td>4233 SW</td>
<td>193</td>
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<tr>
<td>352 ST</td>
<td>1 ST</td>
<td>1639 ST</td>
<td>4092 SW</td>
<td>150</td>
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<tr>
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<tr>
<td>237 R</td>
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<tr>
<td>126 P</td>
<td>1 P</td>
<td>2162 P</td>
<td>3516 SW</td>
<td>784</td>
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<tr>
<td>25 F</td>
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<td>4 HM</td>
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* Cells removed from these variant colonial morphologies were propagated in liquid LAZ or YPD medium (10-12 h, at 28 °C, with shaking), then resuspended in sterile glass-distilled water, and plated (approx. 200 c.f.u. per plate) on LAZ or YPD medium respectively. Plates were incubated for 5–7 d at 28 °C.
Wall mannoproteins in C. albicans

Fig. 1. Variant colonial phenotypes of C. albicans ATCC 26555. Each panel contains colonies representative of the different phenotypes. Clonal colonies were grown on LAZ agar at 28 °C for 6 d (see Tables 1 and 2). (a) Wrinkled (W); (b) smooth-white (SW); (c) star (ST); (d) smooth-white-petite (P), and heavy-myceliated (HM); (e) ring (R); (f) fuzzy (F) with a confluent SW colony; (g) flat-ring (FR); (h) stippled-ring (SR). Panel (i) shows the simultaneous appearance of different colonial morphologies similar to those shown in the other panels of the figure.

W colonies (Fig. 1a) revealed an irregularly-shaped thick rim with deep wrinkling throughout their entire surface. SW colonies (Fig. 1b) exhibited an unmottled creamy-white surface. Colonies with the ST morphology (Fig. 1c) possessed a slightly thickened perimeter, which surrounded a star with a variable number of arms projecting peripherally and were equidistant from one another. P colonies (Fig. 1d) were similar to SW colonies in appearance, but significantly smaller in size. HM colonies (Fig. 1d) were small and irregular in shape, with aerial mycelia emanating from the entire colony surface. Colonies with the R appearance (Fig. 1e) exhibited a very thick outer edge, encompassing up to one-half of their radius and their centres were slightly mottled in texture. F colonies (Fig.
1f) exhibited a thickened, irregular rim and a thin interior, with aerial mycelia emanating from the entire colony surface. Colonies with the SR appearance (Fig. 1h) were characterized by a peripheral thick rim around a depressed and mottled central area. Finally, FR colonies (Fig. 1g) were similar to SR colonies but were flatter, with a slightly mottled centre delimited by a thick peripheral rim.

Effect of growth medium on the switching capabilities of the isolated phenotypes

To test whether the variant colony morphologies isolated after the initial plating were heritable, cells removed from primary SW and W clones were resuspended in sterile distilled water and plated as clones on LAZ medium. As shown in Table 1, the parental SW and W phenotypes reappeared en masse in the secondary progeny along with a low percentage of switch phenotypes, detected at frequencies of \(7.4 \times 10^{-3}\) to \(9.5 \times 10^{-3}\). However, an extremely high level of spontaneous switching was observed when cells from SW, W, ST, SR, and P clones (Table 2) were individually grown in liquid LAZ medium and then plated as clones on LAZ agar. This indicates, in agreement with previous reports (Anderson & Soll, 1987; Slutsksy et al., 1985, 1987; Soll et al., 1987), that the limited zinc content of LAZ medium not only assures but apparently increases expression of the colonial variant phenotypes. Of 2500 secondary mature colonies that appeared after plating of cells from a SW colony, 538 (21.5\%) exhibited morphologies other than that of the original phenotype, which represented a combined frequency of switching of \(2.1 \times 10^{-1}\) (Table 2). The highest combined frequency of switching (\(3.6 \times 10^{-1}\)) was observed in colonies cloned from a P colony; in this case, from 2162 colonies examined, 784 (36.2\%) had a morphological appearance distinct from that of the parental colony phenotype (Table 2). Slightly lower values of combined switching frequencies (from \(2.2 \times 10^{-2}\) to \(9.6 \times 10^{-2}\)) were observed after plating of cells from W, ST or SR clones (Table 2).

In contrast, direct plating on YPD agar of cells removed from secondary SW or W colonies that appeared on LAZ (Table 1) or, alternatively, preincubation of cells removed from primary SW, W, ST, SR and P clonal colonies (Table 2) in YPD broth followed by plating of these cells on the same medium, led to the recovery of SW colonies in all cases (Table 1 and 2), with two exceptions. In the case of the colonies cloned from a primary SW colonial phenotype (3746), all but two SR colonies and one SW colony with a wrinkled sector (combined frequency of switching, \(8.0 \times 10^{-4}\)) exhibited the parental SW phenotype (Table 2). Among the 3517 secondary colonies cloned in YPD medium from a primary P colony all but one P colony were SW, which represented a switching frequency of \(2.8 \times 10^{-4}\) (Table 2). The evident loss of colonial phenotypic switching observed in YPD medium (see Tables 1 and 2) may be possibly caused by the high zinc content in this undefined medium (which contains roughly 500-fold more zinc than LAZ medium as determined by atomic absorption spectrometry), since excess of zinc apparently suppresses the appearance of switch phenotypes in other \(C. albicans\) strains (Anderson & Soll, 1987; Slutsksy et al., 1985, 1987; Soll et al., 1987). However, the parental variant phenotypes (SW, W or ST) reappeared after plating on LAZ medium of cells from SW colonies derived from SW, W or ST colonies, respectively (Tables 1 and 2).

Selective effect of incubation temperature and growth medium composition on wrinkled and star cells

Since the stock cultures of the strain ATCC 26555 used to obtain primary phenotypes had been routinely stored at 4°C for prolonged periods, and because all incubations for initial analysis of switching were carried out at 28°C, we considered it of interest to assess the effects of growth temperature (37°C vs 28°C, taking into account the opportunistic pathogenic character of \(C. albicans\)) in combination with different culture media on the phenotypic switching.

To perform these experiments, cells from stabilized tertiary W and ST single clones grown on solid LAZ medium as described above (see Table 3), were removed and incubated for two consecutive rounds in liquid YPD medium at 28°C (Table 3; Fig. 2), and then plated on LAZ or YPD media. Duplicate sets of plates were incubated at either 28°C or 37°C for 5–7 d. At 28°C on LAZ medium the quaternary colonies examined exhibited the morphological shape corresponding to that of the parental tertiary colonial variant from which the cells were plated as clones (Table 3), that is to say the ST (Fig. 2a) or W (Fig. 2e) phenotypes. However, at 37°C all colonies cloned on LAZ medium from either ST or W tertiary colonies were hyper-wrinkled (HW; Fig. 2b,f; Table 3). When cells were plated on YPD medium and incubated at 28°C, the colony morphology observed was SW regardless of the original tertiary clonal phenotypes (ST or W) from which the cells were plated as clones (Fig. 2c,g). A SW phenotype was also obtained after plating of cells from a single ST colony on YPD medium at 37°C (Fig. 2d). However, when cells from a W colony were plated as clones under the same growth conditions, all quaternary colonies examined exhibited a new morphological appearance, which we designated as intermediate-smooth-white-wrinkled phenotype (ISWW; Fig. 2h; Table 3). As suggested by other authors (Dutton & Penn, 1987).
Table 3. Effect of incubation temperature and growth medium composition on the proportion of tertiary and quaternary colonial phenotypes obtained from secondary smooth-white colonies

<table>
<thead>
<tr>
<th>No. of secondary colonies cloned and phenotype*</th>
<th>Parental primary phenotype†</th>
<th>No. of tertiary colonial phenotypes appearing on LAZ medium and predominant phenotype$</th>
<th>No. of tertiary colonies cloned and phenotype‡</th>
<th>No. of quaternary colonial phenotype appearing and predominant phenotype§</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 SW W</td>
<td></td>
<td>4927 (0) W</td>
<td>1 W</td>
<td>2910 (0) 3076 (0) 2743 (0) 2825 (0)</td>
</tr>
<tr>
<td>1 SW ST</td>
<td></td>
<td>4616 (0) ST</td>
<td>1 ST</td>
<td>3112 (0) 2976 (0) 2786 (0) 2851 (0)</td>
</tr>
</tbody>
</table>

Plating medium and incubation temperature:

- LAZ (28 °C)
- LAZ (37 °C)
- YPD (28 °C)
- YPD (37 °C)

* Cells taken from these secondary colonial phenotypes (see Table 2) were preincubated in liquid YPD medium (10-12 h at 28 °C with shaking), resuspended in sterile distilled water and plated (approx. 200 c.f.u. per plate) on LAZ medium. Plates were incubated at 28 °C for 5-7 d. Phenotypes are as in Abbreviations and Fig. legends.

† See Table 2.

‡ Cells removed from each of these two tertiary colonial phenotypes were independently propagated in liquid YPD medium at 28 °C for 10-12 h with shaking for two consecutive rounds (see Fig. 2 for further experimental details), resuspended in sterile glass-distilled water and plated (approx. 200 c.f.u.) on LAZ and YPD medium. Duplicate sets of plates were incubated at either 28 °C or 37 °C for 5-7 d.

§ Numbers in parentheses indicates the switch phenotypes in tertiary and quaternary colonies, respectively.
1989) the colony morphology appears to be determined by the relative proportions of mycelial or yeast forms, as the cells removed from SW colonies were exclusively in the budding form of growth (Fig. 2, panel 2), whereas W and HW colonies contained mostly mycelial cells (Fig. 2, panel 1). ST and ISWW colonies contained roughly equal amounts of budding cells and mycelial filaments.

Characterization of wall mannoproteins isolated from different colony phenotypes

Purified walls obtained from cell samples removed from ST (Fig. 2a), W (Fig. 2e), HW (Fig. 2b, f), SW (Fig. 2c, d, g), and ISWW (Fig. 2h) colonies, were digested with Zymolyase and the solubilized mannoprotein material separated by SDS-PAGE. Mannoproteins electrophoretically transferred to nitrocellulose paper by Western blotting, were detected with ConA, a polyclonal antiserum enriched in antibodies against wall components specific for the mycelial morphology (mPAb), or a monoclonal antiserum (mAb 4C12).

The mannoprotein nature of the molecules solubilized by the β-glucanase complex was revealed by ConA–peroxidase staining of the nitrocellulose blots (Fig. 3a). Different mannoproteins with molecular masses ranging from higher than 600 kDa down to about 30 kDa were visualized. Mannoproteins with apparent molecular masses > 200 kDa were particularly abundant in enzymic digests of wall preparations from cells of SW (Fig. 3a, lanes 3, 4, 7), ST (Fig. 3a, lane 1) and ISWW (Fig. 3a, lane 8) colonies. Polydispersity of the high-molecular-mass mannoproteins was high, especially in those lanes that were loaded with material released by Zymolyase from cells belonging to heavily myceliated colonial phenotypes (Fig. 3a, lane 6). This phenomenon may result from an increase in the glycosylation of some wall mannoprotein species.

A polyclonal antiserum generated against mannoprotein components of purified wall preparations from blastospores bearing germ tubes (mPAb; Casanova et al., 1989) was also tested on the enzymic digests by immunoblotting, and several of the high-molecular-mass mannoprotein species visualized by ConA (Fig. 3a) had antigenic determinants which were recognized by the mPAb preparation (Fig. 3b). Very large antigenic mannoproteins (> 500 kDa) were detected almost exclusively in the digests of wall preparations from cells of SW (Fig. 3b, lanes 3, 4, 7), ST (Fig. 3b, lane 1), and ISWW (Fig. 3b, lane 8) colonies. This result agrees with that for ConA (above).

Finally, a monoclonal antibody (mAb 4C12) raised against a mycelium-specific wall mannoprotein band with an apparent molecular mass of 260 kDa (Casanova et al., 1989), reacted with various mannoprotein bands with apparent molecular masses of > 200 kDa (Fig. 3c). Such species were detected in the wall digests of cells from ST (Fig. 3c, lane 1), W (Fig. 3c, lane 5), HW (Fig. 3c, lanes 2, 6) and ISWW (Fig. 3c, lane 8) colonies, which all contained, as stated above, variable proportions of mycelial growth forms. In contrast, mAb 4C12 did not react with the enzymically solubilized material from the walls of SW phenotype cells (Fig. 3c, lanes 3, 4, 7), which were exclusively in the budding form of growth, although these digests contained a great assortment of mannoprotein species, as revealed by ConA–peroxidase staining (Fig. 3a, lanes 3, 4, 7).

Discussion

The spontaneous high frequency of switching in colony morphology on agar media reported in this study of C. albicans ATCC 26555 (a common laboratory strain) is similar to the switching system initially described by Slutsky et al. (1985) in strain 3153A of C. albicans.

Various genetic mechanisms have been invoked to explain the molecular basis of phenotypic transitions in C. albicans (Morrow et al., 1989; Pomés et al., 1987; Rustenchenko-Bulgac et al., 1990; Scherer & Stevens, 1988; Slutsky et al., 1985; Söll et al., 1987; Suzuki et al., 1989) Our results suggest that some environmental conditions (i.e. the growth medium and/or the incubation temperature) may differentially affect expression of a set of genes (which among other possibilities, may change the activity of enzymes that synthesize wall components) responsible for the manifestation of differ-

Fig. 2. Effect of temperature and growth medium on the expression of variant colony morphologies. From star (ST) or wrinkled (W) tertiary clonal colonies (see Table 3), cells were inoculated into liquid YPD medium and rotated on a gyratory incubator (250 r.p.m.) at 28 °C for 10–12 h. Cells from these cultures were transferred to fresh liquid YPD medium at an initial concentration of 5 × 10⁶ spheres or cells per ml⁻¹, and incubated again as described above for an additional 10–12 h. Cells from these cultures were suspended in water and plated on solid LAZ medium (a, b, e, f), and YPD medium (c, d, g, h). Plates were incubated at 28 °C (a, e, c, g), and at 37 °C (b, f, d, h). Samples removed from the surface of colonies exhibiting the star (a), wrinkled (c), hyper-wrinkled (HW; b, f), or intermediate smooth-white-wrinkled (ISWW; h) morphologies contained variable amounts of budding cells and mycelial filaments (panel 1; see text), whereas cells removed from smooth-white colonies (c, d, g) were exclusively in the budding form of growth (panel 2). Panels 1 and 2 are phase-contrast observations. Bar, 12.5 μm.
Fig. 3. Western blots of 5 to 15% slab gradient gels loaded with Zymolyase-released material (approx. 400 μg per well expressed as total sugar content in the sample subjected to electrophoresis) from purified wall preparations obtained by ballistic disruption of cells taken from the different variant colony morphologies shown in Fig. 2. As indicated at the top of Fig. 2, cells from stabilized tertiary star or wrinkled colonies were plated as clones on LAZ or YPD media, at either 28 °C or 37 °C (see text, and Table 2 for further experimental details). Lane 1, ST colonies (Fig. 2a); lane 2, HW colonies (Fig. 2b); lanes 3 and 4, SW colonies (Fig. 2c, d); lane 5, W colonies (Fig. 2e); lane 6, HW colonies (Fig. 2f); lane 7, SW colonies (Fig. 2g); lane 8, ISWW colonies (Fig. 2h). Blots were stained either with ConA (a), with germ-tube-specific PAbs (b), or with mAb 4C12 (c). Molecular masses of the proteins standards are indicated to the left of each panel.
ent colonial phenotypes. These results agree with previously published reports indicating that certain extracellular conditions can affect switching in the white-opaque transition of C. albicans (Rikkerink et al., 1988; Slutsky et al., 1987).

The appearance of two new colonial variants, the hyper-wrinkled (HW) and intermediate-smooth-white-wrinkled (ISWW) phenotypes, when cells were plated respectively on LAZ and YPD agar and incubated at 37°C, was interesting. HW colonies had mostly mycelial filaments, and ISWW colonies contained roughly equal amounts of budding cells and hyphae, although the latter were observed on YPD medium (Table 3), which suppressed phenotype expression and produced only SW colonies that contained only yeast cells. As 37°C is the natural temperature of the human body, and mycelial filaments are thought to be more invasive than budding cells, the appearance of these seemingly temperature-dependent switch phenotypes (HW and ISWW) may relate to the in vivo role of switching in pathogenesis.

On the other hand it has been reported that in the white-opaque high-frequency transition system (Slutsky et al., 1987), the switch between white and opaque phenotypes is accompanied by changes in the adhesion characteristics and surface properties of cells (Kennedy et al., 1988), and by changes in the antigenic composition of the cell surface (Anderson & Soll, 1987; Anderson et al., 1989, 1990). Attempts to determine the surface components of C. albicans involved in adherence and antigenicity have revealed that cell wall mannoproteins fulfil this role (Casanova et al., 1989; Chaffin et al., 1988; Douglas, 1987; Sundstrom et al., 1988). A novel feature of this work is the characterization of antigenic mannoproteins released by Zymolyase from the cell walls of different phenotypes.

We have detected a complex mannoprotein composition in the walls of cells belonging to the ST, W, HW, SW and ISWW quaternary clones (Fig. 2a–h). Some of these mannoproteins exhibited reactivity towards the mPAb preparation, and differences in the pattern of antigenic mannoproteins were observed depending on the phenotype considered. However, it has to be stressed that the mPAb, which was prepared by repeated absorption of an antiserum raised against mycelial cell walls with blastoconidia, still contained antibodies against yeast-specific wall components (Casanova et al., 1989). Hence, the differences in the pattern of antigens revealed by the mPAb could be due to the expression of yeast- and mycelium-specific cell-wall mannoproteins, whose relative proportions in the Zymolyase digests may vary depending on the percentage of yeasts and mycelial filaments present in each colonial phenotype (see above). Further evidence reinforcing this possibility came from experiments performed with the monoclonal antibody mAb 4C12. This antibody was initially raised against a mannoprotein band with an apparent molecular mass of 260 kDa, which was specific for the mycelial-phase walls (Casanova et al., 1989). However, further experiments in our laboratory have revealed that mAb 4C12 also recognizes a 180 kDa ϒ-glycosylated mannoprotein (p180) which is secreted by regenerating protoplasts obtained from mycelial cells, and which appears to be the precursor of the 260 kDa mycelium-specific band (Elorza et al., 1989). As expected, the monoclonal antibody reacted only with mannoprotein antigens in the wall digests from cells of the ST, W, HW, or ISWW clones, which all contained cells in the hyphal growth form.

However, it is also possible that changes in the molecular structure of the wall mannoproteins may affect, directly or indirectly (Goochee & Monica, 1990), the way these mannoproteins are bound to other wall components, which may occur as a consequence of the expression of a particular phenotype. This possibility arises from the observation that mAb 4C12 reacted with mannoprotein bands whose relative proportions in the quaternary ST, W, HW, or ISWW colony digests (equal amounts of the different samples expressed as total sugar content were subjected to electrophoresis), as well as their degree of polydispersity, appear to vary depending on the colonial switch phenotype considered. Given that mAb 4C12 is apparently directed against an epitope in the peptide moiety of p180, the different mannoprotein bands detected by the monoclonal antibody may reflect variations in the glycosylation levels of p180, or, more likely, in the way this molecule interacts with other wall components, which would result in the solubilization by Zymolyase of supramacromolecular complexes with different mobilities. There is evidence for the formation of covalent bonds between chitin and glucans (Surarit et al., 1989) or chitin and glucans (Surarit et al., 1989) or chitin (unpublished observations). If these changes occur concomitantly with the expression of specific colonial phenotypes, they may confer new surface properties upon the cells expressing such phenotypes, which could in turn modulate the opportunistically pathogenic character of C. albicans.

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