Case Report

A rare case of breakthrough fungal pericarditis due to fluconazole-resistant *Candida auris* in a patient with chronic liver disease

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**Introduction:** *Candida* pericarditis is a rare clinical entity with a high fatality, primarily attributed to difficulty in diagnosis. Unfortunately, the diagnosis is made post-mortem in more than 50% of cases, and thus a high index of clinical suspicion is crucial.

**Case presentation:** We report a rare case of fungal pericardial effusion caused by the recently recognized multidrug-resistant *Candida auris*, which was cultured from pericardial fluid, blood, bronchoalveolar lavage and urine of a chronic liver disease patient while on empiric fluconazole therapy. The yeast was misidentified as *Candida haemulonii* by the VITEK2 commercial identification system, and was confirmed as *C. auris* by internal transcribed spacer and large ribosomal subunit sequencing. In addition, the VITEK2 AST card erroneously revealed a high amphotericin B MIC (16 μg ml^-1^) and low caspofungin MIC (0.25 μg ml^-1^) that did not correlate with results from the reference Clinical and Laboratory Standards Institute (CLSI) microbroth dilution method. Based on VITEK2 MIC data, the patient was administered caspofungin. However, *in vitro* antifungal susceptibility data for *C. auris* by the CLSI method exhibited high MICs to fluconazole (64 μg ml^-1^) and caspofungin MIC (1 μg ml^-1^) but low MICs to amphotericin B (MIC range, 0.125–0.5 μg ml^-1^). The patient’s repeat pericardial fluid culture, despite caspofungin therapy for 12 days, grew *C. auris* and he died on day 13 of therapy.

**Conclusion:** *C. auris* is a recently reported agent of fungaemia and deep-seated infections and is notable for its antifungal resistance. Although early species identification and rapid antifungal susceptibility testing are needed in cases of critical infections, the reporting of rare yeast isolates exhibiting high MICs to antifungals by automated systems needs a cautionary approach.

**Keywords:** *Candida auris*; *Candida pericarditis*; caspofungin; fluconazole resistance; India.
parameters (blood urea nitrogen 133 mg dl\(^{-1}\) and serum creatinine 3.6 mg dl\(^{-1}\)). Bacterial cultures of blood, sputum and ascitic fluid were negative. The patient was anuric with compensated metabolic acidosis. He was treated empirically with meropenem, colistin, haemodialysis IV albumin and fluconazole (FLU) 150 mg once a day for 2 weeks. Subsequently, 2 weeks after admission, he reported shortness of breath with cough and expectoration. On examination, both lower lung fields revealed bilateral coarse crepitations and a raised jugular venous pressure with tachycardia without hypotension. An electrocardiogram showed low voltage complexes and his chest X-ray showed bilateral consolidation, interstitial oedema and gross cardiomegaly with obliteration of cardiophrenic angle suggestive of pericardial effusion. The findings were confirmed on echocardiography and a significant amount of pericardial effusion with tamponade was observed. The patient was put on ventilator support, and pericardiectomy was done. About 250 ml haemorrhagic fluid was drained and a sheath was placed in situ for further drainage. The pericardial fluid analysis showed a leukocyte count of 3600 \(\mu\)l\(^{-1}\), with predominant polymorphs, and grew yeasts, which were identified as *Candida haemulonii*, by VITEK2 (BioMérieux). The antifungal susceptibility was performed using a VITEK2 AST Yeast card (BioMérieux), which revealed that the isolate had high MICs to FLU (32 \(\mu\)g ml\(^{-1}\)) and amphotericin B (AMB) (16 \(\mu\)g ml\(^{-1}\)) but low MICs to caspofungin (CAS) (0.25 \(\mu\)g ml\(^{-1}\)). *C. haemulonii* was also isolated from urine, blood and bronchoalveolar lavage. All the isolates were sent to a reference Medical Mycology centre for molecular identification. The patient was administered CAS, with a loading dose of 70 mg, followed by 50 mg daily. Although his bilirubin level decreased to 10 mg dl\(^{-1}\) within 1 week, the derangement of renal parameters continued. His repeat pericardial fluid culture and urine grew *C. haemulonii* despite CAS treatment for 12 days. On day 13, his condition deteriorated and he finally succumbed to his illness due to septic shock. All four isolates from pericardial fluid, blood, bronchoalveolar lavage and urine developed a pink colour on CHROMagar *Candida* medium (Difco; Becton Dickinson) and showed ovoid to elongated budding yeast cells. Repeat identification by VITEK2 Compact (BioMérieux) and API20C (BioMérieux) misidentified them as *C. haemulonii* (96% identity) and *Candida sake* (87% identity), respectively. For molecular identification, genomic DNA was extracted as described by Xu et al. (2000). The internal transcribed spacer (ITS) region of the rRNA was amplified using established ITS-1/ITS-4 primers and the D1/D2 region of the large ribosomal subunit (LSU) using NL-1/NL-4 primers (White et al., 1990; Kurtzman & Robnett, 1997). The amplified DNA was sequenced and GenBank BLAST searches were done for species identification. The ITS (GenBank accession nos KF689023.1, KJ126758.1 and KJ126759.1; LSU accession nos KC692056.1, KJ126763.1 and KJ126762.1). Also, LSU sequences of our *C. auris* isolates showed 99% similarity with a Korean *C. auris* isolate (GenBank accession no. JX459779.1).

Antifungal susceptibility testing of the four isolates was performed in duplicate using the Clinical and Laboratory Standards Institute (CLSI) microbroth dilution method (CLSI, 2008). The MIC end points for azoles, AMB and echinocandins were read visually after 24 h, as validated recently by Pfaffer et al. (2008, 2011). All four isolates had reduced susceptibility to FLU (MIC >64 \(\mu\)g ml\(^{-1}\)) and CAS (MIC 1 \(\mu\)g ml\(^{-1}\)). However, posaconazole (MIC ≤ 0.015 \(\mu\)g ml\(^{-1}\)), itraconazole (MIC range 0.03–0.125 \(\mu\)g ml\(^{-1}\)), voriconazole (MIC range 0.06–0.125 \(\mu\)g ml\(^{-1}\)) and 5-flucytosine (MIC range 0.125–4 \(\mu\)g ml\(^{-1}\)) exhibited potent activity. In addition, micafungin (MIC 0.06 \(\mu\)g ml\(^{-1}\)) and anidulafungin (MIC range 0.125–0.25 \(\mu\)g ml\(^{-1}\)) showed good activity. For all the isolates, the drugs tested revealed reproducible MICs when performed by different persons on two occasions revealing only a onefold difference in dilutions. The AMB and CAS susceptibility testing by VITEK2 was again repeated to check the reproducibility in the reference laboratory and revealed similar results of high (8–16 \(\mu\)g ml\(^{-1}\)) and low (0.25 \(\mu\)g ml\(^{-1}\)) MICs, respectively. The MIC values obtained by VITEK2 were in stark contrast to the data obtained using the CLSI method, which were reproducible on two occasions and revealed a high susceptibility to AMB (MIC range 0.125–0.5 \(\mu\)g ml\(^{-1}\)) and a low susceptibility to CAS (1 \(\mu\)g ml\(^{-1}\)) against all four isolates.

**Discussion**

This pericarditis case reported is unusual as it brings to the fore a rare diagnosis due to *C. auris* in a patient with CLD. *C. auris*, is a recently reported agent of fungaemia and deep-seated infections (Lee et al., 2011; Chowdhary et al., 2013, 2014). It is notable for its antifungal resistance and so far has been reported from Japan, Korea, India and South Africa (Kim et al., 2009; Satoh et al., 2009; Lee et al., 2011; Chowdhary et al., 2013, 2014; Magobo et al., 2014). In the present case, the *C. auris* had elevated MICs to the empirically used antifungal (i.e. FLU) and exhibited a high MIC for CAS, which is a matter of concern, especially in patients in peritransplant settings. Similarly, breakthrough infections due to FLU-resistant *C. albicans* and *C. glabrata* and *C. krusei* in neutropenic and transplant recipient patients have been reported (Wingard et al., 1991; Safdar et al., 2001; Myokon et al., 2003). Furthermore, not only was the yeast misidentified as *C. haemulonii* by a commercial identification system, but the high MIC of AMB and low MIC of CAS were also erroneously reported, therefore limiting the use of antifungals for patient management. The VITEK2 system is one of the most common automated systems used for identification and testing the susceptibility of clinically significant yeasts in routine conditions.
microbiology laboratories. However, the possibility of misidentification remains in this commercial identification system, particularly when rare species are reported. This may be attributed to the fact that the system attempts to generate identification of the rare species, which are not available in the existing database, rather than yielding no identification. Furthermore, isolates with antifungal resistance, particularly againstazole-class agents, amphotericin and echinocandins, necessitate the accurate in vitro susceptibility testing of medically significant yeasts. It is noteworthy that C. haemulonii and C. auris differ in susceptibility to AMB, with C. auris being susceptible and C. haemulonii being remarkably resistant to AMB and azoles. Previously, false-positive resistance results for AMB by the VITEK2 system were reported for C. lusitaniae and C. kefyr (Melhem et al., 2014). In the present study, the high AMB MIC value for C. auris, as determined by the VITEK2 system, did not correlate with the results using the CLSI method, suggesting that future studies evaluating a larger number of C. auris isolates would further confirm our observations. Moreover, the antifungal susceptibility data for this rare species are limited, and only five reports, presented in Table 1, have been published (Kim et al., 2009; Satoh et al., 2009; Lee et al., 2011; Chowdhary et al., 2013, 2014). Although early species identification and rapid susceptibility testing are needed in cases of critical infections, the reporting of rare yeast isolates exhibiting high MICs to antifungals by automated systems needs a cautionary approach.

Previously reported predisposing factors for Candida pericarditis include thoracic surgery, malignancy, chronic steroids, diabetes, transplants, burns and catheter injury to the myocardium (Schrank & Dooley, 1995; McNamee et al., 1998; Neugebauer et al., 2002; Puius & Scully, 2007; Parikh et al., 2009; Tang et al., 2009). The majority of these patients, owing to underlying conditions, received broad-spectrum antibiotics leading to colonization by Candida, which can gain access to the mediastinum or bloodstream following invasive procedures. Our patient was a chronic alcoholic with decompensated liver disease, and had been on antibiotics for 2 weeks. Table 2 updates the cases of Candida pericarditis reported so far globally. C. albicans has been implicated in 47.5% of cases reported globally, followed by C. tropicalis (10%) and C. glabrata (5%); notably, in one-third of cases, species identification was not done. Overall, Candida pericarditis is often recognized late because of non-specific clinical symptoms and often-negative blood cultures for Candida. In the majority of the cases reviewed in Table 2, AMB remains the treatment of choice. Also, early pericardial drainage is essential for a successful outcome. In the present case, the initial empirical treatment with FLU for 2 weeks was followed by CAS, for which the isolate had a MIC of 1 μg ml⁻¹. Recently, the epidemiological cut-off values (ECVs) for the echinocandins were reported to be ≤0.25 μg ml⁻¹ for seven species. C. glabrata, C. parapsilosis and

### Table 1. Antifungal susceptibility data of previously published C. auris isolates

<table>
<thead>
<tr>
<th>Reference</th>
<th>Method of susceptibility tested</th>
<th>MIC range (μg ml⁻¹)</th>
<th>AMB</th>
<th>FLU</th>
<th>VRC</th>
<th>ISA</th>
<th>POS</th>
<th>CAS</th>
<th>MFG</th>
<th>AFG</th>
<th>FC</th>
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<tbody>
<tr>
<td>Satoh et al. (2009)</td>
<td>Not mentioned</td>
<td>0.5–1</td>
<td>0.06–0.25</td>
<td>0.06</td>
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<td>Kim et al. (2009)</td>
<td>CLSI (2008)</td>
<td>0.25–1</td>
<td>0.06–0.25</td>
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<tr>
<td>Lee et al. (2011)</td>
<td>CLSI (2008)</td>
<td>0.25–1</td>
<td>0.06–0.25</td>
<td>0.06</td>
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<td>Chowdhary et al. (2013)</td>
<td>CLSI (2008)</td>
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<td>0.06–0.25</td>
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<tr>
<td>Present study</td>
<td>CLSI (2008)</td>
<td>0.25–1</td>
<td>0.06–0.25</td>
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AMB, amphotericin; FLU, fluconazole; VRC, voriconazole; ISA, isavuconazole; POS, posaconazole; CAS, caspofungin; MFG, micafungin; AFG, anidulafungin.
C. guilliermondii ECVs for echinocandins are ≤2 μg ml⁻¹; however, no breakpoints or ECVs for rare Candida spp. such as C. auris have yet been published (Pfaller & Diekema, 2012). Moreover, Espinel-Ingroff et al. (2013) recently reported inter-laboratory MIC variations of CAS for Candida spp. using the CLSI method, which are similar to the European Committee on Antimicrobial Susceptibility Testing (EUCAST) MIC variations observed for the four common Candida spp. from seven laboratories. Furthermore, use of anidulafungin and micafungin MICs as markers for the echinocandins instead of CAS is recommended for making clinical therapeutic decisions (Arendrup et al., 2011; Pfaller et al., 2014). In our case, we could not demonstrate clinical success with CAS, although the MICs with both the CLSI and VITEK2 methods were ≤1 μg ml⁻¹. In addition, the failure of CAS administration in the eradication of C. auris is in consonance with previous reports demonstrating that this yeast is difficult to eradicate and causes persistent fungaemia despite therapy (Lee et al., 2011; Chowdhary et al., 2013, 2014). High mortality rates and therapeutic failure in 66% of patients with C. auris infections have been reported (Chowdhary et al., 2014). Finally, the importance of correct identification and standardized antifungal susceptibility testing for optimal management strategies of patients with invasive infections due to rarely emerging resistant Candida spp. can hardly be overemphasized.
References


