**Klebsiella pneumoniae** meningitis induces memory impairment and increases pro-inflammatory host response in the central nervous system of Wistar rats

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**INTRODUCTION**

Previous studies have demonstrated that **Klebsiella pneumoniae** and other Gram-negative bacilli have become a common cause of meningitis, mainly in patients with head trauma, neurosurgical operations and impaired host defences (Lu et al., 1999, 2002; Mancebo et al., 1986; Mangi et al., 1975). However, there are few studies concerning **K. pneumoniae** meningitis in the literature (Wen et al., 2007). Bacterial meningitis is a life-threatening infection with high mortality and neurological sequelae (Sellner et al., 2010). Several studies have reported that cognitive processes, such as learning and memory, are affected after bacterial meningitis (Bedford et al., 2001; Schmand et al., 2010).

Recently, Morichi et al. (2013) reported alterations in brain-derived neurotrophic factor (BDNF) in the serum and cerebrospinal fluid (CSF) of paediatric patients with bacterial meningitis. The crucial role of BDNF in learning...
and memory is well described in the literature (Bekinschtein et al., 2007; Rattiner et al., 2005), suggesting that the cognitive dysfunction observed after bacterial meningitis may be linked to changes in this neurotrophin. Morichi et al. (2013) also demonstrated that alterations in BDNF were correlated with increased cytokine levels in the CSF, blood platelet counts and neurological prognoses in patients with bacterial meningitis.

In addition, recent studies from our laboratory have demonstrated that meningitis caused by Streptococcus pneumoniae and Streptococcus agalactiae resulted in cognitive impairment, decreased BDNF levels and increased cytokine/chemokine levels in the brains of rats (Barichello et al., 2010, 2012a, b, 2013). Bacterial meningitis induces a distinct systemic inflammatory response that has been well described in clinical and experimental subjects (Barichello et al., 2013; Morichi et al., 2013). Clinical and preclinical studies have demonstrated that bacterial invasion of the subarachnoid space triggers potent host responses with elevated levels of pro-inflammatory cytokines/chemokines, such as TNF-α, IL-1β, IL-6 and cytokine-induced neutrophil chemoattractant-1 (CINC-1) (Barichello et al., 2013; Morichi et al., 2013; Scheld et al., 2002).

However, the host immune inflammatory response and behavioural alteration caused by K. pneumoniae meningitis infection remain poorly understood (Wen et al., 2007). The current study examined the effects of acute bacterial meningitis induced by K. pneumoniae on memory, and the levels of pro-inflammatory cytokines, chemokines and BDNF in the frontal cortex, hippocampus and CSF of rats.

**METHODS**

**Infecting organism.** K. pneumoniae was cultured overnight in 10 ml Todd–Hewitt broth, diluted in fresh medium and grown to the exponential phase. The culture was centrifuged for 10 min at 5000 g and resuspended in sterile saline to a concentration of 1 c.f.u. ml⁻¹. The size of the inoculum was confirmed by quantitative culture (Barichello et al., 2010; Irazuzta et al., 2002).

**Animal model of meningitis.** Adult male Wistar rats (250–300 g body weight) from our breeding colony were used for the experiments. All procedures were approved by the Animal Care and Experimentation Committee of UNESC, Brazil, and were performed in accordance with the National Institute of Health Guide for the Care and Use of Laboratory Animals (NIH Publications No. 80-23), as revised in 1996. All surgical procedures and bacterial inoculations were performed under anaesthesia consisting of intraperitoneal administration of ketamine [6.6 mg (kg body weight)⁻¹], xylazine [0.3 mg (kg body weight)⁻¹] and acepromazine [0.16 mg (kg body weight)⁻¹] (Grandirard et al., 2007; Hoogman et al., 2007). Rats underwent a cisterna magna tap with a 23-gauge needle, receiving either 10 μl sterile saline as a placebo or an equivalent volume of K. pneumoniae suspension. At the time of inoculation, the animals received fluid replacement (2 ml saline subcutaneously) and were subsequently returned to their cages. Eighteen hours later, meningitis was documented by a quantitative culture of 5 μl CSF obtained by puncture of the cisterna magna (Barichello et al., 2010). The animals were sacrificed 6, 12, 24, 48 or 96 h after meningitis induction to assay TNF-α, IL-1β, IL-6 CINC-1 and BDNF concentrations.

**Assays of TNF-α, IL-1β, IL-6, CINC-1 and BDNF concentrations.** Animals were sacrificed by decapitation at 6, 12, 24, 48 or 96 h after meningitis induction. The hippocampus, frontal cortex and CSF were immediately isolated on dry ice and stored at −80 °C to analyse the cytokine, chemokine and BDNF levels. Brain tissue was homogenized in extraction solution (100 mg ml⁻¹) containing 0.4 M NaCl, 0.05 % Tween 20, 0.5 % BSA, 0.1 mM PMSF, 0.1 mM benzethonium chloride, 10 mM EDTA and 20 kallikrein inhibitor units aprotinin, using an Ultra-Turrax homogeniser (Fisher Scientific). Brain homogenates were centrifuged at 3000 g for 10 min at 4 °C, and the supernatants were collected and stored at −20 °C. The concentration of chemokines and cytokines were determined using ELISA and commercially available antibodies according to the procedures provided by the manufacturer (R&D Systems). BDNF levels were measured as described previously (Frey et al., 2006), using commercial sandwich ELISA kits according to the manufacturer’s instructions (Chemicon). The results were expressed as pg (100 μl CSF)⁻¹ and pg (100 mg cerebral tissue)⁻¹.

**Open-field task.** Eighteen hours after meningitis induction, the animals received ceftriaxone [100 mg (kg⁻¹ body weight)⁻¹] given subcutaneously, over the course of 7 days. Ten days after meningitis induction, the animals were free from infection. Blood cultures performed after the course of treatment were negative (data not shown). The animals recovered their weight and grooming habits, and blood counts returned to control levels. Habituation to an open field was performed in a 40 × 60 cm open field surrounded by 50 cm high walls made of brown plywood with a frontal glass wall. The floor of the open field was divided into 12 equal rectangles by black lines. The animals were gently placed on the left rear quadrant and allowed to explore the arena for 5 min (training session). Immediately following training, the animals were taken back to their home cage and were submitted to a similar open-field session (test session) 24 h later. The number of line crossings and rearings were recorded for both sessions. The decrease in the number of crossings and rearings between the two sessions was used to measure retention of habituation (Vianna et al., 2000). After the animals were sacrificed by decapitation, the hippocampus and frontal cortex were immediately isolated on dry ice and stored at −80 °C for BDNF analysis.

**Step-down inhibitory avoidance task.** The apparatus and procedures have been described in previous reports (Quevedo et al., 1997; Roesler et al., 2003). Briefly, the training apparatus was a 50 × 25 × 25 cm acrylic box (Albarsch) with a floor consisting of parallel 1 mm calibre stainless steel bars spaced 1 cm apart. A 7 cm wide, 2.5 cm high platform was placed on the floor of the box against the left wall. In the training session, the animals were placed on the platform, and the latency before they stepped down on the grid with all four paws was measured with an automatic device. Immediately after stepping down on the grid, the animals received a 0.4 mA, 2.0 s foot shock and were returned to their home cage. A retention test trial was performed 24 h after training (long-term memory). The retention test trial was procedurally identical to the training, except that no foot shock was presented. The retention test step-down latency (maximum, 180 s) was used as a measure of inhibitory avoidance retention. Reactivity to the foot shock was evaluated in the same apparatus used for inhibitory avoidance, except that the platform was removed. Each animal was placed on the grid and allowed a 1 min habituation period prior to the start of a series of 0.5 s shocks delivered at 10 s intervals. The shock intensities ranged from 0.1 to 0.5 mA in 0.1 mA increments. The adjustments in shock intensity were made in accordance to each animal’s response. The intensity was raised by one increment when no response occurred and was lowered by one increment when a response was observed. A ‘lurch’ response was defined as the...
withdrawal of one paw from the grid floor, and a 'jump' response was defined as rapid withdrawal of three or four paws. Two measurements of the 'flinch' threshold were made, and then two measurements of the 'jump' threshold were made. For each animal, the means of the two scores for the flinch and the jump thresholds were calculated (Bevilaqua et al., 2003; Izquierdo et al., 1998).

Statistics. For the cytokine, chemokine and BDNF level analyses, the variables were reported as the means ± SEM of the five to six animals in each group. Differences among groups were evaluated using ANOVA followed by a Student–Newman–Keuls post-hoc test. *P<0.05, **P<0.01, ***P<0.001; statistical significance compared with the control group.

Fig. 1. Kinetics of BDNF, TNF-α, IL-1β, IL-6 and CINC-1 expression in the hippocampus determined at 6, 12, 24, 48 and 96 h after induction of K. pneumoniae meningitis. The concentrations of the cytokines, chemokine and BDNF were assessed by ELISA, and the results are presented as pg (100 mg tissue)⁻¹. The results are the means ± SEM of five to six animals in each group. Differences among groups were evaluated using ANOVA followed by a Student–Newman–Keuls post-hoc test. Data from the habituation to an open-field task were reported as the means ± SEM of the 10 animals in each group, and were analysed by a paired Student’s t-test and ANOVA followed by Tukey’s post-hoc test. Data from the inhibitory avoidance task were reported as the median and interquartile ranges, and comparisons among groups were performed using a Mann–Whitney U test. Within-group differences were analysed by a Wilcoxon test. A value of P<0.05 indicated statistical significance compared with the control group.
RESULTS

Fig. 1 shows the effects of *K. pneumoniae* meningitis on the TNF-α, IL-1β, IL-6, CINC-1 and BDNF levels in the hippocampus at 6, 12, 24, 48 and 96 h after meningitis induction. In the hippocampus, TNF-α was increased at 6 and 12 h (*P*<0.05 and *P*<0.001, respectively); IL-1β and IL-6 were increased at 6, 12 and 24 h (*P*<0.05); CINC-1 was elevated at 6, 12, 24, 48 and 96 h (*P*<0.05) and BDNF levels were increased at 24 h and decreased at 96 h (*P*<0.01 and *P*<0.05, respectively) after *K. pneumoniae* meningitis induction.

Fig. 2 shows the effects of *K. pneumoniae* meningitis on the TNF-α, IL-1β, IL-6, CINC-1 and BDNF levels in the frontal cortex at 6, 12, 24, 48 and 96 h after meningitis induction. In the frontal cortex, IL-1β levels were increased at 6 (*P*<0.001), 12 (*P*<0.001) and 24 h (*P*<0.01); IL-6 levels were increased at 6, 12, 24 and 48 h (*P*<0.01 and *P*<0.05, respectively); CINC-1 was elevated at 6, 12, 24 and 48 h (*P*<0.05) and BDNF levels were increased at 24 h and decreased at 96 h (*P*<0.01 and *P*<0.05, respectively) after *K. pneumoniae* meningitis induction.
were elevated at 6 ($P<0.05$), 12 ($P<0.001$) and 24 h ($P<0.05$) and CINC-1 levels were elevated at 6 ($P<0.001$), 12 ($P<0.001$), 24 ($P<0.05$), 48 ($P<0.001$) and 96 h ($P<0.001$) after *K. pneumoniae* meningitis induction. In the same brain structure, the BDNF levels were increased at 6 ($P<0.05$), 12 ($P<0.01$), 24 ($P<0.001$) and 48 h ($P<0.01$) but decreased at 96 h ($P<0.05$) after *K. pneumoniae* meningitis induction.

Fig. 3 shows the effects of *K. pneumoniae* meningitis on the TNF-$\alpha$ and IL-1$\beta$ levels in the CSF at 6, 12, 24, 48 and 96 h after meningitis induction. In the CSF, the TNF-$\alpha$ and IL-1$\beta$ levels were elevated at 6 ($P<0.01$), 12 ($P<0.001$) and 24 h ($P<0.01$) after *K. pneumoniae* meningitis induction.

Fig. 4 shows the effects of *K. pneumoniae* meningitis on the inflammatory response, BDNF levels and cognition of Wistar rats. The extent of the inflammatory response was monitored by measuring the production levels of the inflammatory mediators TNF-$\alpha$,
IL-1β, IL-6 and CINC-1. Meningitis induced by K. pneumoniae led to elevated levels of TNF-α, IL-1β and IL-6 in the hippocampus, elevated IL-1β and IL-6 in the frontal cortex and increased concentrations of TNF-α and IL-1β in the CSF, primarily at 6, 12, 24 and 48 h after K. pneumoniae inoculation. In addition, acute bacterial meningitis increased CINC-1 at all the times that it was assayed and in both brain structures evaluated. However, it is evident that the increase in CINC-1 levels regressed in a time-dependent manner. CINC-1 (the rat homologue of human IL-8) is one of the most important chemokines, and is induced by pro-inflammatory cytokines, including TNF-α and IL-1β. CINC-1 is involved in leukocyte transmigration into tissues, an activity not only of leukocytic cells (monocytes and neutrophils) but also non-leukocytic cells (endothelial cells, fibroblasts and epithelial cells) (Kuwahara et al., 2006; Mukaida, 2003).

Our data indicated that TNF-α, IL-1β, IL-6 and CINC-1 participated in inflammatory processes induced by K. pneumoniae meningitis, primarily 6, 12 and 24 h after meningitis induction. Herein, we also demonstrated that K. pneumoniae meningitis induced an increase in BDNF levels in the hippocampus at 24 h, but this was followed by a decrease at 96 h. In the frontal cortex, the BDNF levels increased at 6, 12, 24 and 48 h but decreased 96 h after K. pneumoniae meningitis induction. In agreement with our study, Morichi et al. (2013) demonstrated that serum BDNF levels were elevated in patients with central nervous system infections, and the elevation was particularly notable in those with bacterial meningitis. In the same study, BDNF expression in the CSF was correlated with CSF interleukin IL-6 levels, blood platelet counts and neurological prognoses in patients with bacterial meningitis. Previous studies have demonstrated that cytokines such as IL-1β, IL-6 and TNF-α may provoke the dysregulation of several growth factors, including BDNF, fibroblast growth factor-2, macrophage migration inhibitory factor and erythropoietin (Anisman & Hayley, 2012; Jurgens & Johnson, 2012). Moreover, bioactive BDNF is also expressed in immune cells, lesions of multiple sclerosis and animal models of autoimmune encephalomyelitis (Lee et al., 2012). Immune cell-derived BDNF could be relevant to the observed protective effects. This neurotrophin is expressed in T cells, B cells, activated monocytes and macrophages (Kerschensteiner et al., 1999). Furthermore, neurons are the primary origin of BDNF in the CNS (Lewin & Barde, 1996). In this study, we found that K. pneumoniae meningitis induced an initial increase in the BDNF levels, but over time, a reduction of this neurotrophin occurred compared with the control animals.

In addition, we demonstrated that K. pneumoniae meningitis induced learning and memory impairment 10 days after meningitis induction. A potential contributing factor to the cognitive decline during K. pneumoniae infection could be inflammation of the hippocampus and frontal cortex. Previous studies have demonstrated that elevated inflammatory mediators can have damaging effects on cognitive function (McAfoose & Baune, 2009; Yirmiya & Goshen, 2011).

In conclusion, cognitive performance was impaired after K. pneumoniae-induced bacterial meningitis in Wistar rats and could be associated with alterations in the levels of cytokines, chemokines and BDNF in the central nervous system.

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