

Escherichia coli carrying the *bla*_{CTX-M-15} gene of ST648

*bla*_{CTX-M} genes have emerged as the dominant genes encoding extended-spectrum β -lactamases in the world. Among more than 90 *bla*_{CTX-M} genes that have been identified so far, *bla*_{CTX-M-15} is the most widespread variant. Although *Escherichia coli* of O antigen type 25 (O25) and sequence type 131 (ST131) is largely responsible for the intercontinental spread of *bla*_{CTX-M-15}, some other sequence types have also been found associated with this gene. During a local surveillance of *bla*_{CTX-M} genes, an isolate of ST648 carrying *bla*_{CTX-M-15} was identified and is reported here. As an ST648 isolate carrying *bla*_{CTX-M-15} was found in the USA (Sidjabat *et al.*, 2009) recently, the findings here together with the USA report suggest a possible intercontinental spread of this lineage.

E. coli clinical isolate WCE227 was collected from a urine sample and was resistant to cefotaxime (MIC >32 $\mu\text{g ml}^{-1}$) and ceftazidime (MIC >16 $\mu\text{g ml}^{-1}$) as determined by the Phoenix automated microbiology system (BD). The WCE227 was hospital acquired as it was recovered from a patient hospitalized for more than 48 h, although the patient did not receive antimicrobial agents prior to sample collection. The *bla*_{CTX-M} gene was detected and was subsequently identified as *bla*_{CTX-M-15} by PCR and sequencing as described elsewhere (Zong *et al.*, 2008).

WCE227 was identified as being of the O25b subtype by O25b allele PCR. Phylogenetic group typing (Clermont *et al.*, 2000) revealed that WCE227 belonged to group D (subgroup D₁, having *chuA* but lacking *yjaA* and TspE4.C2). WCE227 was of ST648 as determined by multilocus sequence typing (MLST) following an established scheme based on seven housekeeping genes (*adhA*, *fumC*, *gyrB*, *icd*, *mdh*, *purA* and *recA*) available through the University College Cork MLST database (<http://mlst.ucc.ie/mlst/dbs/Ecoli>).

Since *bla*_{CTX-M-15} usually co-exists with a few other resistance determinants, *aac*(3)-II,

*bla*_{TEM}, *bla*_{OXA-1} (also called *bla*_{OXA-30}) and plasmid-encoded quinolone resistance determinants including *qnrA*, *qnrB*, *qnrS* and *qepA*, were screened by PCR, and *aac*(6')-Ib-cr was identified by sequencing. WCE227 had *bla*_{OXA-1} and *aac*(6')-Ib-cr but no others. WCE227 were resistant to trimethoprim (TMP)/sulfamethoxazole and had a class 1 integron with the *dfrA17*–*aadA5* cassette array that was determined by PCR and sequencing. Class 1 integrons contain *sulI*, a sulfonamide-resistance gene, in the 3' conserved segment. *dfrA17* encodes TMP-insensitive dihydrofolate reductases conferring resistance to TMP. As WCE227 was resistant to ciprofloxacin, the *gyrA* allele was partially sequenced. This revealed Ser83Tyr and Asp87Asn substitutions that have been seen in fluoroquinolone-resistant isolates before (Cagnacci *et al.*, 2008).

Mating was carried out in brain heart infusion broth (Oxoid) with *E. coli* DH5 α Rf, a spontaneous rifampicin-resistant mutant of DH5 α (Δ *lacZ*) as the recipient strain. WCE227 was sensitive to rifampicin and transconjugants were selected on 4 μg cefotaxime ml^{-1} plus 250 μg rifampicin ml^{-1} . In WCE227, *bla*_{CTX-M-15} was carried by a conjugative plasmid, for which the incompatibility (Inc) group could not be assigned by PCR-based replicon typing (Carattoli *et al.*, 2005). Surprisingly, *aac*(6')-Ib-cr and *bla*_{OXA-1} were not co-transferred with *bla*_{CTX-M-15} in WCE227, in contrast to the findings of reports from elsewhere. The *dfrA17*–*aadA5* cassette array was also not located on the plasmid carrying *bla*_{CTX-M-15} in WCE227.

ST648 isolates carrying *bla*_{CTX-M} have been seen before, including two isolates carrying *bla*_{CTX-M-15} from the USA (Sidjabat *et al.*, 2009) and two carrying *bla*_{CTX-M-1} or *bla*_{CTX-M-32} from Spain (Blanco *et al.*, 2009). All of those ST648 isolates like WCE227 belonged to phylogenetic group D. Interestingly, ST648 without *bla*_{CTX-M-15} but carrying *aac*(6')-Ib-cr had been found in London, UK, prior to the

epidemic of *bla*_{CTX-M-15} (Jones *et al.*, 2008). The findings from the UK together with the fact that *aac*(6')-Ib-cr was not located on the plasmid carrying *bla*_{CTX-M-15} in WCE227 suggest that the ST648 lineage acquired the two genes independently. This is in contrast to the ST131 lineage, for which *bla*_{CTX-M-15} and *aac*(6')-Ib-cr were usually co-transferred on individual plasmids, most of which were FII-like.

Acknowledgements

This work was partially supported by a grant from the National Natural Science Foundation of China (project no. 30900052).

Zhiyong Zong and Rujia Yu

Center of Infectious Diseases, West China Hospital, Sichuan University, Chengdu, PR China

Correspondence: Zhiyong Zong (zongzhiyong@gmail.com)

Blanco, M., Alonso, M. P., Nicolas-Chanoine, M. H., Dahbi, G., Mora, A., Blanco, J. E., López, C., Cortés, P., Llagostera, M. & other authors (2009). Molecular epidemiology of *Escherichia coli* producing extended-spectrum β -lactamases in Lugo (Spain): dissemination of clone O25b:H4-ST131 producing CTX-M-15. *J Antimicrob Chemother* **63**, 1135–1141.

Cagnacci, S., Gualco, L., Debbia, E., Schito, G. C. & Marchese, A. (2008). European emergence of ciprofloxacin-resistant *Escherichia coli* clonal groups O25:H4-ST 131 and O15:K52:H1 causing community-acquired uncomplicated cystitis. *J Clin Microbiol* **46**, 2605–2612.

Carattoli, A., Bertini, A., Villa, L., Falbo, V., Hopkins, K. L. & Threlfall, E. J. (2005). Identification of plasmids by PCR-based replicon typing. *J Microbiol Methods* **63**, 219–228.

Clermont, O., Bonacorsi, S. & Bingen, E. (2000). Rapid and simple determination of the *Escherichia coli* phylogenetic group. *Appl Environ Microbiol* **66**, 4555–4558.

Jones, G. L., Warren, R. E., Skidmore, S. J., Davies, V. A., Gibreel, T. & Upton, M. (2008). Prevalence and distribution of plasmid-mediated quinolone resistance genes in clinical

isolates of *Escherichia coli* lacking extended-spectrum β -lactamases. *J Antimicrob Chemother* 62, 1245–1251.

Sidjabat, H. E., Paterson, D. L., Adams-Haduch, J. M., Ewan, L., Pasculle, A. W., Muto, C. A., Tian,

G. B. & Doi, Y. (2009). Molecular epidemiology of CTX-M-producing *Escherichia coli* isolates at a tertiary medical center in western Pennsylvania. *Antimicrob Agents Chemother* 53, 4733–4739.

Zong, Z., Partridge, S. R., Thomas, L. & Iredell, J. R. (2008). Dominance of *bla*_{CTX-M} within an Australian extended-spectrum β -lactamase gene pool. *Antimicrob Agents Chemother* 52, 4198–4202.