Evaluation of protection against *Chlamydophila abortus* challenge after DNA immunization with the major outer-membrane protein-encoding gene in pregnant and non-pregnant mice

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The protective effect of DNA vaccination with the gene encoding the major outer-membrane protein (MOMP) of *Chlamydophila abortus* has been studied in non-pregnant and pregnant mouse models after chlamydial challenge. OF1 outbred mice were vaccinated intramuscularly three times every 3 weeks, mated and challenged with *C. abortus* 2 weeks after the last injection of DNA. In non-pregnant mice, the MOMP DNA vaccine elicited a specific humoral response with predominantly IgG2a antibodies, suggesting a Th1-type immune response. The induced antibodies showed no *in vitro* neutralizing effect on *C. abortus* infectivity. Moreover, immunization with the *momp* gene showed no reduction in the mean splenic bacterial counts of non-pregnant or pregnant mice or in the mean placental bacterial counts of pregnant mice after the *C. abortus* challenge. Nevertheless, the MOMP DNA immunization induced a non-specific and partial protection in fetuses against challenge.

**INTRODUCTION**

One of the most common causes of considerable loss in breeding is the abortion of goats and sheep induced by the intracellular obligate bacterium *Chlamydia abortus* (*Chlamydiaceae* genus of the *Chlamydiaceae* family) (Papp & Shewen, 1996). Furthermore, these bacteria present a zoonotic risk to pregnant women, since several cases of human chlamydial infections have been reported (Buxton, 1986). A live attenuated vaccine is currently used in small ruminants (Rodolakis, 1983). This vaccine is effective and safe but does not allow the detection of infected animals in vaccinated flocks.

The predominant component and immunodominant antigen of the *Chlamydiaceae* surface membrane, the 40 kDa major outer-membrane protein (MOMP), presents four surface-exposed variable domains (VDI–VDIV) flanked by five conserved regions and exhibits an oligomeric form, probably trimeric, that functions as a porin (De Sa, 1996). In a murine model of *C. abortus* infection, protective immunity can be conferred by antibodies (Buzoni-Gatel et al., 1990) and, more particularly, by the passive transfer of specific antibodies to the 110 kDa oligomeric MOMP (De Sa et al., 1995), so MOMP has been considered as the most likely vaccine candidate against *C. abortus* infections. Native MOMP (Pal et al., 1997; Tan et al., 1990), recombinant MOMP (Tuffrey et al., 1992), synthetic MOMP peptides (Su et al., 1995) and live recombinant vectors (Murdin et al., 1995) expressing *momp* genes from members of the *Chlamydiaceae* have been evaluated in different animal models. Limited immunity and protection were generated in animals when the vaccinal approaches attempted to preserve the conformational structure of MOMP. Nevertheless, most trials were unsuccessful. A recent study showed that protective immunity against a *Chlamydia trachomatis* genital challenge was induced with a vaccine based on the MOMP associated with lipophilic immune response-stimulating complexes (Igietseme & Murdin, 2000).

DNA vaccination represents an exciting means to induce specific immunity to a single antigen and, consequently, fits in the marker vaccine family (van Oirschot et al., 1996). Indeed, the direct injection of a DNA plasmid encoding an immunogenic protein into animals has resulted in both humoral and cell-mediated specific immune responses (Donnelly et al., 1997). Since DNA vaccination preferentially elicits a Th1-type immune response, it appears to be particularly appropriate for preventing intracellular bacterial infection (Strugnell et al., 1997). This novel strategy of vaccination has been assessed in different animal models and seems suitable for veterinary medicine (Babiuk et al., 2000).

DNA vaccination with the *momp* gene has already been tested in *C. trachomatis* (Pal et al., 1999; Zhang et al., 1997), *Chlamyphila pneumoniae* (Penttilä et al., 2000) and...
**METHODS**

*C. abortus* strains. The virulent *C. abortus* strain AB7 isolated from an ovine abortion (Faye et al., 1972) and the vaccinal strain IB obtained by selection of a temperature-sensitive mutant after nitrosoguanidine treatment of *C. abortus* AB7 (Rodolakis, 1983) were used. Bacteria were propagated in yolk sacs of embryonated specific-pathogen-free chicken eggs inoculated at day 7, purified as described previously (Caldwell et al., 1981) and stored at −80 °C.

**Cloning of the momp gene.** To produce the vaccinal vector, the *momp* gene was amplified by PCR and cloned into the vector pcDNA3.1 (Invitrogen) carrying the human cytomegalovirus immediate early promoter and the bovine growth hormone polyadenylation signal.

**Immunization and challenge.** All the studies were carried out with 6-week-old female outbred OF1 Swiss mice (IFFA Credo). Prior to DNA vaccination, each mouse was injected with cardiotoxin (Latoxan) into the tibialis anterior muscles of both hind legs to enhance the uptake of DNA. Immunization, each mouse was injected with cardiotoxin (Latoxan) into the tibialis anterior muscles of both hind legs to enhance the uptake of DNA. Immunization and challenge.

**Cloning of the momp gene.** To produce the vaccinal vector, the *momp* gene was amplified by PCR and cloned into the vector pcDNA3.1 (Invitrogen) carrying the human cytomegalovirus immediate early promoter and the bovine growth hormone polyadenylation signal. The PCR was performed using chlamydial genomic DNA (80 ng) as template with dNTPs (200 μM each), specific primers (1 μM each) and 1 U Platinium polymerase (Promega) in a Perkin Elmer 9600 thermocycler with the following program: 94 °C for 5 min, 1 cycle; 30 cycles of 50 °C for 45 s, 72 °C for 2 min and 45 s and 1 cycle of 50 °C for 45 s and 72 °C for 10 min. Specific primers were determined according to the DNA sequence of *C. abortus* AB7 containing an EcoRI site (underlined) and a Kozak sequence (bold) in the forward primer (MOMP-FW: 5’-CTGAAATCCAGCTGAAACATACTTGAAACGGC-3’) and an XhoI site (underlined) in the reverse primer (MOMP-RV: 5’-CTCTGAGCTAGAATCTGAATTGAGCATTC-3’). The resulting fragment was inserted into the vector pcDNA3.1 after linearization by EcoRI/XhoI double digestion to generate pcDNA3.1::MOMP. The plasmid pcDNA3.1::MOMP and the pcDNA3.1 control plasmid were purified after an overnight ligation using a Luria–Bertani culture of recombinant *Escherichia coli* DH5x using the EndoFree Plasmid Mega kit (Qiagen). Plasmid DNA was dissolved at 1 μg μl−1 in endotoxin-free PBS (Sigma). The sequence of the vaccinal vector pcDNA3.1::MOMP was verified by DNA sequencing.

**Immunization and challenge.** All the studies were carried out with 6-week-old female outbred OFI Swiss mice (IFAA Credo). Prior to DNA immunization, each mouse was injected with cardiotoxin (Latoxan) into the tibialis anterior muscles of both hind legs to enhance the uptake of plasmid DNA (Davis et al., 1993). Five days later, mice were anaesthetized by an intraperitoneal injection of ketamine and xylazine (respectively 80 and 8 mg kg−1 body weight) and immunized with pcDNA3.1::MOMP plasmid by intramuscular injections (50 μg in each tibialis anterior). Mice were boosted in the same way at days 21 and 42. The negative-control mice were immunized intramuscularly with endotoxin-free PBS (virelence control) or pcDNA3.1 plasmid. Positive-control mice were immunized with one subcutaneous injection of 4 × 10^6 p.f.u. of the live attenuated 1B vaccine at day 1. Groups of 10 and 20 mice were similarly immunized for evaluation of the protective effect of the DNA vaccine and respectively constituted the non-pregnant and pregnant groups. The five groups of pregnant mice were mated at day 44. Non-pregnant and pregnant mice were challenged at day 58 by an intraperitoneal injection of 4 × 10^6 p.f.u. *C. abortus* AB7. One group of pregnant mice neither immunized nor challenged was kept as a control for the pregnancy (gestation control).

**Antibody response.** Blood samples were collected from the retro-orbital sinus of non-pregnant mice for the detection of anti-MOMP-specific antibodies before each DNA injection (days 0, 19, 41), just before the challenge infection (day 56) and at day 63. Blood samples were stored overnight at room temperature and then centrifuged (600 g, 15 min, 4 °C) and sera were collected and stored at −20 °C. The specificity of anti-MOMP antibodies was checked by ELISA as described previously (Héchard et al., 2002). The recombinant MOMP used as antigen was kindly provided by Dr R. Ashley (University of Edinburgh, UK) (Wyllie et al., 1999). Reconstituted MOMP (10 μl), denatured by heating (100 °C for 10 min) and diluted in Mg2+−free, Ca2+−free PBS (pH 7.4) at a concentration of 1 μg ml−1, was used for coating microtiter plates (Nunc-Immu No Plate Maxisorp Surface) for 16 h at 4 °C. Sera used for ELISA were serially diluted in Mg2+−free, Ca2+−free PBS (pH 7.4), 0.05 % Tween 20; 1:200 for the less reactive serum or starting from 1:200 to 1:800 for the more reactive serum.

**Determination of antibody isotypes.** Determination of antibody isotypes was carried out using the ELISA as described above. Briefly, after incubation with dilutions (1:50) of the pooled sera of pcDNA3.1::MOMP-vaccinated mice collected at day 56 or 63, the isotypes were revealed by adding 100 μl of a 1:1000 dilution of peroxidase-conjugated anti-mouse IgG1 or IgG2a (Nordic Immunological Laboratories). The A405 was measured on a microplate reader (Multiskan RC, Thermo Labsystems) after 1 h of incubation with the substrate (ABTS) at room temperature.

**In vitro neutralization assay in the presence of the complement.** The neutralization assay was done by the plaque-reduction assay using McCoy cells (Schachter et al., 1974). Pooled sera of PBS−, pcDNA3.1− and pcDNA3.1::MOMP-vaccinated mice were diluted in Mg2+−free, Ca2+−free PBS (pH 7.4) containing 10 % fetal bovine serum and 2-5 % guinea pig serum. Two hundred microlitres of these dilutions (10−1, 10−2 and 10−3) were added to an equal volume (200 μl) of an AB7 chlamydial suspension (10^6 p.f.u.), incubated for 2 h at 37 °C and then titrated on McCoy cells by the plaque assay method (Rodolakis & Chancerelle, 1977). The results are expressed for each serum as the percentage p.f.u. of the PBS control.

**Evaluation of protection.** The protective effect of the DNA vaccine was assessed by the evaluation of the number of living offspring per litter at birth or 8 days after birth and the clearance of bacteria in spleens, placentas and fetuses. After challenge, the outcome of pregnant mice was monitored daily. Mice were considered as protected when the number of living offspring per litter 8 days after birth was significantly different (P < 0.05) from the number for the virulence control as described previously (Rodolakis, 1983). All the non-pregnant mice and five mice of each group of pregnant mice were euthanized at day 63. The spleens from non-pregnant mice and the uteruses and spleens from pregnant mice were removed aseptically. All the placenta from the same uterine horn were dissected from the fetuses. Placentas from the same uterine horn were pooled as well as the fetuses. Pooled placentas, pooled fluids and spleens were weighed and frozen at −80 °C. Organs were homogenized in PBS with a glass grinder, diluted in PBS-DEAE dextran (0.01 %) and titrated by plaque assay on McCoy cells (Rodolakis & Chancerelle, 1977). The course of infection was evaluated by counting...
the number of p.f.u. and expressed as log_{10} p.f.u. per organ for spleens and log_{10} p.f.u. per uterine horn for placentas and fetuses.

Statistical tests. Analysis of the results was performed using the software InStat 2.03 for Macintosh. For all protective results, the mean was calculated using a one-way analysis of variance and a comparison of the means was then carried out through a Student–Newman–Keuls multiple comparison test. The minimal statistical significance was judged at $P < 0.05$.

RESULTS

Analysis of the humoral response

The anti-MOMP IgG antibody response was assessed by ELISA (Fig. 1). The anti-MOMP IgG titre reached a maximum in the non-pregnant mice immunized with pcDNA3.1::MOMP after the third DNA injection (day 56) and decreased after challenge (day 63). This decrease in anti-MOMP IgG antibody titre at day 63 was also observed without challenge (data not shown). The substantial variations of the IgG titre in individual mice on the last two sampling days were probably due to the fact that the murine model used in these experiments was an outbred one. No MOMP-specific IgG antibody was identified in sera from non-immunized mice (data not shown) or from mice immunized with pcDNA3.1 (Fig. 1). The specificity of anti-MOMP antibodies was also examined by immunoblotting. Sera collected from immunized mice before any DNA injection (day 0) or before challenge (day 56) and after challenge (day 63) were tested on crude extracts of *C. abortus* AB7 as well as on boiled recombinant MOMP. Since no MOMP-specific antibodies were detected (data not shown), we could conclude that the anti-MOMP antibody titre was probably too low to detect native or recombinant MOMP by immunoblotting.

The isotypes of anti-MOMP antibodies of sera collected at days 56 and 63 from pcDNA3.1::MOMP-vaccinated mice were tested by ELISA. The predominant isotype was IgG2a (data not shown). No IgG1 was found. The ability of the sera to neutralize the infectivity of the virulent AB7 strain of *C. abortus* was assessed by the plaque-reduction test. The sera of 1B-vaccinated mice had a substantial neutralizing effect (90 % at the 10^{-1} dilution; Fig. 2), while the sera of pcDNA3.1- and pcDNA3.1::MOMP-vaccinated mice respectively had a modest neutralizing effect (60 %) and a non-significant neutralizing effect (20 %).

Efficacy of DNA immunization on pregnancy

As expected, the number of live newborn mice observed in the 1B group was not significantly different from the number of live newborns observed in the gestation control group ($P > 0.05$), but it was significantly higher ($P < 0.01$) than the number monitored in the virulence group (Fig. 3). The numbers of living offspring at birth and 8 days after birth for the gestation control and 1B groups were not significantly different. For the virulence group and the pcDNA3.1- and pcDNA3.1::MOMP-vaccinated mice, the number of living offspring at birth was more than at 8 days after birth, but this difference was not significant. The numbers of live newborn in the virulence, pcDNA3.1, and pcDNA3.1::MOMP groups were not significantly different ($P > 0.05$), but were significantly lower ($P < 0.05$) than in the gestation control and 1B groups. Therefore, pcDNA3.1::MOMP-vaccinated mice were not protected against abortion due to *C. abortus* AB7 challenge.

Protection was also evaluated at a placental and fetal level by titration of *C. abortus* in these organs. Chlamydial titres in placentas were higher than in fetuses ($P < 0.001$). As expected, chlamydial titres in the placentas and fetuses of 1B-vaccinated mice were significantly lower ($P < 0.01$) than in the virulence group (Fig. 4). 1B-vaccinated mice were consequently protected against chlamydial infection. Chlamydial titres in the placentas of pcDNA3.1- and pcDNA3.1::MOMP-vaccinated mice were not significantly lower than in the virulence group (Fig. 4).
different from those of the virulence group ($P < 0.05$). This would indicate that DNA vaccination did not elicit sufficient protection against placental colonization. Nevertheless, the fetuses of pcDNA3.1- and pcDNA3.1::MOMP-vaccinated mice were significantly less infected than those of the virulence group ($P < 0.05$) (Fig. 4). Thus, partial protection was observed in fetuses of 1B-, pcDNA3.1- and pcDNA3.1::MOMP-vaccinated mice.

**Clearance of *C. abortus* AB7 from the spleen**

The induced protection was evaluated by splenic titration of *C. abortus* on pregnant and non-pregnant mice. As expected, chlamydial titres in spleens from non-pregnant or pregnant 1B-vaccinated mice were significantly lower ($P < 0.001$ and $P < 0.01$, respectively) than in the virulence group. Thus, 1B-vaccinated mice were protected against chlamydial infection (Fig. 5). Chlamydial titres in the spleens from pcDNA3.1- and pcDNA3.1::MOMP-vaccinated, non-pregnant or pregnant mice were not significantly different from those from mice of the virulence group ($P > 0.05$). These results suggested that DNA vaccination did not induce splenic protection in pregnant or non-pregnant mice.

**DISCUSSION**

In veterinary medicine, vaccines need to overcome a large number of hurdles, including the cost of production, the ease of delivery, safety and the complete protection of animals against disease. DNA vaccination has recently been tested against many pathogens and is promising in veterinary medicine (Babiuk *et al.*, 2000). The aim of this study was to investigate the protective effect of DNA vaccination with the MOMP-encoding gene against *C. abortus* challenge in a murine model. The mouse model usually employed for the evaluation of *C. abortus* virulence (Rodolakis *et al.*, 1989) as well as vaccine efficiency (Rodolakis, 1983) was the OF1 outbred mouse model of abortion. This model presented various advantages. It was easy to perform, since it consisted of evaluation of the living offspring per mouse. Moreover, the protective effects of vaccines obtained in OF1 outbred mice were correlated with those observed in the ovine model. Finally, this mouse model has been used successfully in the past to evaluate the efficiency of 1B vaccine and is now taken as a reference (Rodolakis, 1983). Nevertheless, this OF1 mouse model presented two major disadvantages. Firstly, as it involves outbred mice, this model is not suitable for...
evaluation of the cellular immune response. Secondly, it can be weakly discriminatory, since a small difference in the titre of the C. abortus challenge (nearness abortive dose 50 %) can modify the number of living offspring very significantly. Together with the high variability between the results obtained with outbred mice, this could have made the protection observed non-significant. Thus, the comparable mean numbers of living offspring obtained from pcDNA3.1- and pcDNA3.1::MOMP-vaccinated mice drove us to investigate more precisely the protection in other models of infection evaluation, including the splenic, placental and fetus bacterial burden inquiries.

Although DNA immunization with the momp gene elicited production of MOMP in vivo, since it induced specific anti-MOMP antibodies, it failed to reduce the abortive effect of C. abortus challenge. No protection was observed after MOMP DNA immunization in either the placental colonization or splenic colonization model. However, partial reduction of infection was observed in fetuses of pcDNA3.1- and pcDNA3.1::MOMP-vaccinated mice. As placentas were infected, fetuses were probably protected by a barrier effect, but this protective effect was not sufficient to allow the survival of living offspring. This non-specific and partial protection could be explained by the immunostimulating properties of the CpG sequences present in the vaccinal plasmid backbone (Krieg, 2000). These unmethylated CpG motifs are recognized as a danger signal by the vertebrate immune system and trigger a Th1-biased immune response.

In summary, DNA vaccination with a plasmid vector encoding the momp gene elicited a weak humoral response induced by the DNA vaccination, we failed to detect epitope-specific antibodies. Probably due to the weak humoral response induced by the DNA vaccination, we failed to detect either conformational or linear MOMP after immunoblotting, a less sensitive technique compared with ELISA.

Different methods have been evaluated in an attempt to increase the immune response generated by DNA vaccines (Gurunathan et al., 2000). More particularly, the vaccination protocol could be improved, as has already been done in a C. trachomatis model (Dong-Ji et al., 2000), using a DNA prime and a protein boost strategy known to enhance both humoral immune response and protection (Song et al., 2000). In addition, it might be possible to amplify the protective immune response by co-administration of an immunoregulatory cytokine-encoding plasmid with the vaccinal plasmid (Gurunathan et al., 2000). A protein boost and the co-administration of cytokine-encoding plasmids can also be combined in order to achieve protection (Scheeinkin et al., 2001).

In summary, DNA vaccination with a plasmid vector encoding the momp gene of C. abortus elicited a weak humoral response, with the IgG2a isotype predominant. Despite the antigenic properties of MOMP, DNA vaccination failed to protect pregnant mice from abortion against C. abortus challenge. In contrast, fetuses from DNA-vaccinated mice were non-specifically and partially protected. Therefore, it would be of interest to optimize the vaccination protocol and to use genetic adjuvants including cytokines or co-stimulating molecule genes in order to enhance the immune response.

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