BACTERIAL PATHOGENICITY

Heat-shock protein 60 homologue of Helicobacter pylori is associated with adhesion of H. pylori to human gastric epithelial cells

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A previous study reported a relationship between the expression of heat-shock protein 60 (HSP60) by Helicobacter pylori and its adhesion to human gastric carcinoma (MKN45) cells. To examine whether the HSP60 homologue of H. pylori is associated with the adhesion of H. pylori to human gastric epithelial cells, an inhibition assay of adhesion of H. pylori to MKN45 cells was performed by flow cytometric analysis with monoclonal antibody (MAb) designated as H20 recognising HSP60 of H. pylori. The rate of adhesion of H. pylori pretreated with MAbH20 to MKN45 cells was lower than that of untreated H. pylori. Primary human gastric epithelial cells from a patient with gastric cancer were also prepared for comparison in the inhibition assay with MAbH20. H. pylori adhered to the primary human gastric epithelial cells, and this adhesion was significantly inhibited by MAbH20. These results suggest that the H. pylori HSP60 homologue recognised by MAbH20 might be associated with the adhesion of H. pylori to primary human gastric epithelial cells as well as to cultured gastric cancer cells.

Introduction

Helicobacter pylori is associated with chronic gastritis, gastroduodenal ulcer and gastric adenocarcinoma [1–3]. Although virulence factors such as flagella [4, 5], adhesin [6], urease [7, 8], vacuolating toxin [9, 10] and the toxin that inhibits secretion of gastric acid in vitro [11, 12] have been identified, the relationship between these H. pylori antigens and the mechanism by which H. pylori persists in the stomach is not clear.

A study of bacteria-epithelial cell adhesion, the first step in H. pylori infection, is important for any understanding of the mechanism of induction of tissue inflammation in the stomach by H. pylori to emerge. Several ligand-receptor interactions between H. pylori and gastric epithelial cells have been reported: N-acetylneuraminylactose-binding fibrillar haemagglutinin [13], haemagglutination activities with strain specificity [13, 14], binding to GM3 ganglioside and sulphatides [15, 16], and a laminin-binding protein [14, 17] have been demonstrated. These indicate that the adhesion of H. pylori to human gastric epithelial cells might be multifactorial.

A recent study demonstrated that there is a correlation between the adhesion of H. pylori to human gastric carcinoma cells and the expression of heat-shock protein 60 (HSP60) on the cell surface [18]. Huesca et al. [19] reported that HSP60 of H. pylori localised on the bacterial surface, which mediates sulphatide recognition, might be related to the adhesion of H. pylori to gastric epithelial cells. HSP60, conserved not only in prokaryotic cells but also in eukaryotic cells, is thought to facilitate folding, unfolding and translocation of polypeptides as chaperonins [20–25]. Accordingly, the interaction between HSP60 of H. pylori and the adherence characteristics of H. pylori is important for an understanding of the mechanism of persistent infection with H. pylori.

In the present study, in order to clarify whether H. pylori HSP60 is associated with adhesion to human gastric epithelial cells, an inhibition assay of adhesion to human gastric carcinoma cells (MKN45) was performed. These experiments were repeated with primary human gastric epithelial cells from a patient with gastric cancer.
Materials and methods

Bacterial strains and culture conditions

_H. pylori_ strain TK1029 used in this study was isolated from gastric biopsy material of a patient as described previously [26]. The bacteria were cultured on Brain Heart Infusion (BHI) Agar (Difco) with defibrinated horse blood 5% in an atmosphere of O₂ 5%, CO₂ 10%, N₂ 85% for 4 days at 37°C. Cells of _H. pylori_ TK1029 were centrifuged at 3000 g for 15 min and resuspended in Hank's Balanced Salts Solution (Gibco) containing gelatin 0.1% (Sigma) (HGS) for flow cytometric analysis.

Cell line

Human gastric carcinoma (MKN45) cells were obtained from the Japanese Cancer Research Resources Bank (JCRB). They were grown at 37°C in RPMI 1640 (Gibco) containing fetal calf serum (FCS; Wako Pure Chemical Ltd, Osaka, Japan) 10% in an atmosphere of air with CO₂ 5%. The cells were resuspended in HGS for flow cytometric analysis.

Preparation of primary human gastric epithelial cells

For the preparation of primary human gastric epithelial cells, a specimen of gastric mucosa (c. 5 cm square, 50 g) was obtained from a patient undergoing surgery for gastric carcinoma. The specimen was negative in tests for both rapid urease and isolation of _H. pylori_. The specimen was stored at 4°C in RPMI 1640 medium containing FCS 10% (RF-10). The specimen was then shaken for 60 min at 37°C with 50 ml of pronase solution (10 mg/ml; Pronase MS, Kaken Seiyaku Co., Ltd, Tokyo, Japan) which is a gastric mucinolytic agent. The cell suspension was centrifuged at 1200 rpm for 10 min, and the cell pellet was suspended in RF-10 with Percoll (Sigma) 10% followed by centrifugation at 1800 rpm for 18 min. Subsequently, the cell pellet was suspended again in RF-10 with Percoll 44% and the prepared suspension was put on to RF-10 with Percoll 70%. After centrifugation at 1800 rpm for 18 min, the whole cells on the Percoll 70% were recovered and washed twice with RF-10. The cells formed the preparation of primary human gastric epithelial cells.

Preparation of monoclonal antibodies

The monoclonal antibody (MAb) was prepared by the methods described previously [27]. BALB/c mice were immunised intraperitoneally (i.p.) with partially purified 60-kDa antigen derived from _H. pylori_ TK1029, which had been extracted from the separating gel of SDS-PAGE. These inclusions in Freund's complete adjuvant (Difco) were made at intervals of 10 days. Ten days after the last injection, the mice were given an intravenous injection of the partially purified antigen and 3 days later the spleens were removed for fusion of spleen cells and mouse myeloma cells (P3-X63-Ag8-U1). The hybridoma producing MAb which reacted with the partially purified antigen, purified HSP60 from _Yersinia enterocolitica_ [28] and the sonicated MKN45 cells were collected. The hybridoma with apparent specific antibody production was cloned by limiting dilution and designated as MAbH20 (class: IgM). The hybridoma cells (10⁶ cells) were inoculated i.p. into a BALB/c mouse pre-treated intraperitoneally with 0.5 ml of pristane (Wako Pure Chemical) 4 days before the inoculation of the cells. About 2 weeks later, ascites fluid was obtained from the mouse. The immunoglobulins in the ascitic fluid were purified with an Immunoglobulin-Easy-Separation kit (Pharmacia Biotech. Co., Tokyo, Japan). The purified MAb was used for flow cytometric analysis.

Microsequencing of protein

To examine whether the molecule of _H. pylori_ recognised with MAbH20 was HSP60, microsequencing of the N-terminal amino acids of the protein was performed. The whole proteins of _H. pylori_ TK1029 were separated by SDS-PAGE and transferred on to an Immobilon membrane (Millipore, Bedford, MA, USA). The band corresponding to the protein recognised by MAbH20 was excised from the membrane and sequenced on a protein sequencer (473A; Applied Biosystems, Foster City, CA, USA).

Flow cytometric analysis

The inhibition assay of _H. pylori_ adhesion to MKN45 and primary human gastric epithelial cells was performed by the method described by Osaki et al. [29]. Lipophilic dye PKH-2 (Zynaxis Cell Sciences, Phoenixville, PA, USA) was used to label _H. pylori_ as described by Taguchi et al. [30]. _H. pylori_ labelled with PKH-2 and MKN45 cells were pre-treated with 200 μg/ml, 40 μg/ml and 8 μg/ml of MAbH20. PKH-2 labelled _H. pylori_ with or without treatment with MAbH20 were incubated with either MKN45 cells or primary gastric epithelial cells at 37°C for 1 h in a 1.5-ml tube with gentle shaking. Non-adherent bacteria were removed by centrifugation with 9 ml of sucrose 15% solution. After the cells were washed four times with HGS, they were resuspended in 300 μl of HGS for flow cytometric adherence assay. A flow cytometer (FACSvantedge Coulter Electronics, Hialeah, USA) was used for the measurement of fluorescence intensity. Fluorescence data were obtained in a logarithmic mode on a 1024 channel scale. The results were shown as peak fluorescence intensity and mean (SD) channel indicating the fluorescence frequency distribution histograms.
Results

Preparation of primary human gastric epithelial cells

The profiles containing side and forward scatter of the prepared cells by flow cytometry are shown in Fig. 1. The group of large cells (arrow) was thought to comprise human gastric epithelial cells. In addition, cell morphology was also ascertained for these large cells (Fig. 2). The viability of these cells was determined with propidium iodide (Wako Pure Chemical), and was found to be 95%. This group of cells was gated, and the gated cells were analysed in the inhibition assay of adhesion of *H. pylori* to primary human gastric epithelial cells.

Sequencing of N-terminal amino acids of the 60-kDa antigen of *H. pylori* recognised with MAbH20

The N-terminal amino acid sequence of the *H. pylori* 60-kDa antigen recognised with MAbH20 was analysed. The sequence, ****IKFSVYAMKLLFEGV (* denotes unsuccessful determination), of the first 19 amino acids was determined. Eleven amino acids (underlined) corresponded to the N-terminal amino acids of *H. pylori* HSP60 previously reported by Macchia et al. [31].

Inhibition of adhesion of *H. pylori* to human gastric carcinoma MKN45 cells by MAbH20

Fig. 3 shows the representative patterns of the inhibition of *H. pylori* adhesion to MKN45 cells by MAbH20, directed to *H. pylori* HSP60. In *H. pylori* pre-treated with MAbH20, the fluorescence intensity of the peaks that indicate the adhesion rate of *H. pylori* to MKN45 were significantly decreased (Fig. 3c). The decrease of the adhesion rate was dependent on the concentrations of MAbH20 used (8, 40 or 200 µg/ml). Each adhesion rate, indicated as mean channel (SD) when treated with MAbH20, was 182.3 (86.8), 185.9 (51.3) and 42.2 (25.4) for the results in Fig. 3a, b and c, respectively. There was a significant difference in adhesion when *H. pylori* was pre-treated with MAb and untreated organisms (Mann-Whitney U test, \( p < 0.0001 \)) (Table 1). However, there was no significant difference in the adhesion rates of *H. pylori* when MKN45 cells were pre-treated with MAbH20 (8–40 µg/ml), in comparison to non-pre-treated control. These results indicate that *H. pylori* HSP60 might mediate the adhesion of *H. pylori* to human gastric carcinoma MKN45 cells.

Inhibition of adhesion to *H. pylori* to primary human gastric epithelial cells

Human gastric epithelial cells from a patient with gastric cancer were prepared for use in the inhibition assay with MAbH20. As shown in Figs. 1 and 2, the prepared cells contained a large population of human gastric epithelial cells. The relative proportion of the population of gastric cells which were epithelial was c. 25%. The mean channel number of the gastric epithelial cells adhered with *H. pylori* TK1029 was 188.1 (SD 51.3), and that of negative control of the epithelial cells without *H. pylori* was 3.1 (1.0) (Fig. 4). The results indicate that *H. pylori* TK1029 could adhere to the primary human gastric epithelial cells. When *H. pylori* was pre-treated with each concentration (125–1000 µg/ml) of MAbH20, the adhesion rates of *H. pylori* to the primary epithelial cells were

Fig. 1. Flow cytometry analysis of primary human gastric epithelial cells from gastric tissue of a patient with gastric cancer. Forward scatter (x axis) indicates cell size and side scatter (y axis) indicates molecular density in the cell. The cell group with greater size and density (arrow) is thought to comprise human gastric epithelial cells.
Fig. 2. Morphological features of the prepared human gastric cells (× 200).

significantly decreased compared with the positive control, depending on the concentration of the MAb (Fig. 4). These results suggest that MAbH20 directed to *H. pylori* HSP60 could inhibit the adhesion of *H. pylori* not only to cultured MKN45 cells but also to primary human gastric epithelial cells.

**Discussion**

A recent report showed that there is a significant correlation between the expression of *H. pylori* HSP60 on the cell surface and the adhesion of *H. pylori* to human gastric carcinoma MKN45 cells [18]. Phadnis *et al.* [32] have shown that *H. pylori* HSP60 exists on the cell surface and adheres to extracellular antigen. Huesca *et al.* [19] have also reported that *H. pylori* HSP60 localising on the bacterial surface, which mediates sulphatide recognition, might be related to the adhesion of *H. pylori* to gastric epithelial cells. These findings suggest that *H. pylori* HSP60 might be an adhesin and could be directly associated with the adhesion of *H. pylori* to gastric epithelial cells.

The present study established that MAbH20 is directed against *H. pylori* HSP60 and also demonstrated that the MAb inhibits the adhesion of *H. pylori* to MKN45 cells. The results indicate that *H. pylori* HSP60 could
be related to the adhesion of *H. pylori* to human gastric carcinoma MKN45 cells.

As MKN45 cells are not thought to be natural cells derived from human gastric tissue, primary human gastric epithelial cells were prepared from a patient with gastric cancer. As shown in Fig. 4, *H. pylori* adhered to the primary human gastric epithelial cells. Osaki et al. [29] previously reported that the adhesion of *H. pylori* to three human gastric carcinoma cell lines (MKN45, KATOIII and MKN28) differed, showing that the cell line MKN45 was the most...
Table 1. Inhibitory effect of MAbH20 on adhesion of *H. pylori* to MKN45 cells

<table>
<thead>
<tr>
<th>Concentration of MAbH20 (μg/ml)</th>
<th>Pre-treatment of <em>H. pylori</em></th>
<th>Pre-treatment of MKN45</th>
<th>No pre-treatment of MKN45</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>NT</td>
<td>NT 186.6 (92.0)</td>
<td>NT 185.7 (52.1)</td>
</tr>
<tr>
<td>8</td>
<td>42.2 (25.4)</td>
<td>185.9 (51.3)</td>
<td>182.3 (86.8)</td>
</tr>
<tr>
<td>40</td>
<td>27.8 (16.5)</td>
<td>175.9 (81.3)</td>
<td>170.6 (83.8)</td>
</tr>
<tr>
<td>200</td>
<td>24.6 (15.1)</td>
<td>161.4 (73.1)</td>
<td>156.5 (54.9)</td>
</tr>
</tbody>
</table>

NT, not tested.

*Mean fluorescence intensity of negative control of MKN45 cells was 2.7 (SD 0.9).†Mean ± SD of the mean.

‡Statistically significant (p < 0.0001) in comparison to control without MAbH20.

§Statistically significant (p < 0.001) in comparison to control without MAbH20.

sensitive to *H. pylori* adhesion. The results indicate that unknown host factors are associated with the adhesion of *H. pylori* to gastric epithelial cells.

The adhesion of *H. pylori* to the primary human gastric epithelial cells was inhibited by MAbH20 directed against *H. pylori* HSPG60. These findings indicate that *H. pylori* HSPG60 might be associated with adhesion of *H. pylori* not only to MKN45 but also to human primary gastric epithelial cells.

However, the inhibition of adhesion of *H. pylori* to MKN45 and the prepared human gastric epithelial cells by MAbH20 was not complete. The reason for these phenomena might be explained by the fact that the association between the adhesin(s) of *H. pylori* and host receptors is multifactorial [13–17] (Fig. 5). In addition, it is speculated that the intensity of the expression of the receptor for *H. pylori* HSPG60 on the surface is different in prepared human gastric epithelial cells and in established human gastric cancer MKN45 cells.

![Fig. 4. The inhibitory effect of MAbH20 on the adhesion of *H. pylori* to primary human gastric epithelial cells.](image)

![Fig. 5. Hypothetical scheme to explain the adhesion of *H. pylori* to human gastric epithelial cells and human gastric cancer MKN45 cells.](image)
cancer MKN45 cells (Fig. 5). Further studies are in progress to determine whether *H. pylori* HSP60 is directly associated with the adhesion of *H. pylori* to human gastric epithelial cells.

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### References


