Specific DNA probes to detect *Escherichia coli* strains producing cytotoxic necrotising factor type 1 or type 2

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**Summary.** Cytotoxic necrotising factors type 1 (CNF1) and type 2 (CNF2) are produced by many *Escherichia coli* strains isolated from man and animals with intestinal or extra-intestinal colibacillosis. In most laboratories, CNF-producing strains are detected by a cell cytotoxicity assay and confirmed with a neutralisation assay or a mouse footpad assay. In this study, we sought to determine whether DNA probes could detect clinical isolates of *E. coli* producing CNF2 or CNF1, or both, without the need for cell cultures or animal assays. Two internal fragments of the gene encoding CNF2 were used as DNA probes: a 875-bp *XhoI-PstI* DNA fragment and an adjacent 335-bp *PstI-ClaI* fragment. A positive response with both DNA probes was associated with CNF2-producing strains, whereas a positive response with only the 335-bp probe was associated with CNF1-producing strains. Results of colony hybridisation experiments with 185 clinical isolates of *E. coli* demonstrated that these DNA probes detected CNF2-producing strains with a sensitivity and specificity of 100% and CNF1-producing strains with a sensitivity and specificity of 99%. These two DNA probes should greatly facilitate epidemiological studies to assess the importance of CNF-producing strains as agents of diarrhoea and septicaemia.

**Introduction**

Although *Escherichia coli* belongs to the normal microflora present in the gastrointestinal tract, certain *E. coli* strains have been associated causally with diarrhoea and extra-intestinal infections in man and animals. During the last 10 years, some of the *E. coli* strains isolated from intestinal or extra-intestinal infections have been reported to produce either cytotoxic necrotising factor type 1 (CNF1) or cytotoxic necrotising factor type 2 (CNF2). CNF1 and CNF2 are two monomeric proteins of 110–115 kDa that induce necrosis in rabbit skin and multinucleation of different eukaryotic cells in culture. CNF1 is chromosomally encoded, whereas CNF2 is encoded by a transferable F-like plasmid called Vir.

Experimental infections of neonatal calves with strains carrying the Vir plasmid and of pigs with CNF-producing strains suggest a role for these toxins in pathogenicity of *E. coli*. Production of CNF1 has been demonstrated in *E. coli* strains from: diarrhoea, urinary tract infections, or bacteraemia in man, intestinal or extra-intestinal infections of cats and dogs, enteritis in piglets, diarrhoea or bacteraemia, or both, in calves. CNF2-producing strains have been isolated from calves and lambs with diarrhoea or bacteraemia, or both.

The results of several epidemiological surveys in Europe indicate that CNF-producing strains could represent 10–50% of the *E. coli* strains from extra-intestinal infections and 5–18% of the *E. coli* strains isolated from diarrhoea. One difficulty encountered in conducting epidemiological investigations of CNF1 or CNF2 production is that the only in-vitro method for detecting CNF is to prepare a bacterial cell lysate of the test *E. coli* strain and assess the cytotoxic effects of that lysate on tissue-culture cells. Furthermore, most laboratories use neutralisation of the cytotoxicity by specific anti-CNFI sera or the CNF2-induced necrosis in mouse footpads to distinguish CNF1 from CNF2. Recently, techniques for CNF detection have been improved by the use of mitomycin C to release the toxin into the culture medium and the use of an immunoenzymatic assay to specifically detect CNF1-producing strains or CNF2-producing strains. Nevertheless, these biological and immunological assays are time-consuming and require reagents not readily available to most laboratories. Thus, a rapid and reliable means of detecting strains producing either type of CNF is needed.
We have recently cloned and sequenced cnf2, the gene encoding CNF2, from the DNA of a Vir plasmid. In this study, we sought to determine whether DNA fragments of cnf2 could be used as DNA probes to detect clinical isolates of E. coli producing CNF2 or CNF1, or both, by colony hybridisation. Because the nucleotide sequence of cnf2 is 85.7% identical to that of the recently sequenced gene encoding CNF1, we anticipated that one part of cnf2 might also react with cnf1-containing strains.

Materials and methods

E. coli strains

One hundred and eighty-five E. coli strains isolated from calves, pigs, cats, dogs and human patients with diarrhoea or extra-intestinal infections were screened with the cnf probes described below. These strains included control strains producing some of the most common toxins produced by E. coli: classical heat-stable (StaP and Stb) and heat-labile (LT-I and LT-Ia) enterotoxins; Shiga-like toxins (SLT-I and SLT-II) and z-haemolysin; 115 of these isolates were tested previously for CNF. Seventy E. coli strains were new isolates from colibacillosis cases in Belgium. Three E. coli strains transformed with the Vir plasmids that encode CNF2 were also included in this study: strains 711(pVir) and H209(pVir) were obtained from the late H. W. Smith (Houghton Poultry Research Station, Huntingdon). Strain 712(pJL1) was kindly provided by C. L. Gyles (Department of Veterinary Microbiology and Immunology, University of Guelph, Ontario, Canada). Host strain H209 was a non-pathogenic E. coli strain isolated from the faeces of a healthy individual. Host strains 711 and 712 were two E. coli K-12 strains. The Vir plasmids were isolated from the ovine septicaemic strain S511 and the bovine septicaemic strain JL2. Three E. coli strains were systematically used as controls in this study: strain 711 as the CNF-negative control, strain 711(pVir) as the CNF2-negative control, and the previously described wild-type strain BM2-1 as the CNF1-positive control.

Cell culture assays

The E. coli isolates were inoculated into 100-ml flasks containing 10 ml of Trypticase Soy Broth (TSB; bioMérieux, Charbonnières-les-Bains, France). Cultures were incubated at 37°C for 24 h with shaking (160 rpm) and then centrifuged at 10 000 g for 20 min. The pellets were resuspended in 1 ml of phosphate-buffered saline, pH 7.2 (PBS) containing gentamicin (100 mg/L) and subjected to two freeze-thaw cycles from 37°C to −20°C. Extracts were clarified by centrifugation and tested in the HeLa and Vero cell culture assays as described previously. The CNF2 or CNF1 titre was the highest two-fold dilution of toxic material that caused multinucleation of 50% of the cells after incubation for 72 h (cytotoxic dose 50%, CD50). The serum neutralisation titre was defined as the highest dilution of antisera that neutralised at least 90% of the multinucleating effect produced by CNF. Neutralising antisera against CNF2 and CNF1 were produced in New Zealand White rabbits by four immunisations consisting of multiple intradermal injections performed at 3-week intervals.

Preparation of DNA probes

Two internal DNA fragments of cnf2 were cloned into the high copy vector pBluescript II SK(+) (Stratagene, La Jolla, USA). The resulting recombinant plasmids were designated pEOSW1 and pEOSW3. Plasmid pEOSW1 contained a 335-bp PstI-ClaI DNA fragment of cnf2 and pEOSW3 contained an 875-bp XhoI-PstI DNA fragment of cnf2 (fig. 1). The cnf2 DNA inserts in pEOSW1 and pEOSW3 were isolated by restriction endonuclease digestion of the plasmid DNA followed by gel electrophoresis and electro-elution. These DNA fragments were then radiolabelled with [α-32P]-deoxyribonucleotide triphosphates according to the directions supplied in a multiprimer DNA labelling kit (Boehringer, Brussels, Belgium).

Colony hybridisations

DNA colony hybridisations were accomplished as described previously. Whatman 541 paper filters (Belgolabo, Belgium) were placed on to colonies grown overnight on Luria-Bertani agar. After 2 h, the filters were peeled off and placed colony-side up on to Whatman 3MM paper saturated with the following solutions: sodium dodecyl sulphate (SDS) 10% w/v for 3 min, 0.5 M NaOH for 15 min and 1.0 M Tris- HCl (pH 7.6)−1.5 M NaCl for 5 min, twice. The filters were allowed to dry and hybridised overnight with the DNA probes at 65°C in a solution comprising 3 x SSC (0.45 M NaCl-0.045 M sodium citrate), 10 x Denhardt solution, 0.1% w/v and salmon sperm DNA 1%. The filters were washed three times for 30 min at 65°C in 3 x SSC containing SDS 0.1%, and then subjected to autoradiography.

Plasmid hybridisation assay

Plasmid DNA was extracted from lysed bacteria according to the method of Kado and Liu. Plasmids were separated by electrophoresis overnight at 4°C in an agarose 0.5% gel. The DNA within the gels was denatured and the gels were dried. The gels were subsequently hybridised in situ with the DNA probe under the hybridisation conditions described above.

Evaluation criteria

The standard cytotoxic assay in combination with seroneutralisation was the reference assay for the detection of CNF1 and CNF2 phenotype. The sen-
Table I. Distribution of hybridisation results with the DNA probes derived from pEOSW3 and pEOSW1 according to the CNF phenotype of the 185 E. coli strains tested

<table>
<thead>
<tr>
<th>E. coli phenotype</th>
<th>Number tested</th>
<th>pEOSW1</th>
<th>pEOSW3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Non-CNF producer</td>
<td>64</td>
<td>1 (+)</td>
<td>0</td>
</tr>
<tr>
<td>CNF1-producer</td>
<td>67</td>
<td>66 (+)</td>
<td>0</td>
</tr>
<tr>
<td>CNF2-producer</td>
<td>54</td>
<td>54 (+++)</td>
<td>51 (+++)</td>
</tr>
</tbody>
</table>

Table II. Sensitivity and specificity of the detection of CNF-producing E. coli by colony hybridisation with the DNA probes derived from pEOSW3 and pEOSW1

<table>
<thead>
<tr>
<th>Parameters selected</th>
<th>Phenotype expected</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive with probes from pEOSW3 and pEOSW1</td>
<td>CNF2-producer</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>Positive with probe from pEOSW1 and negative with probe from pEOSW3</td>
<td>CNF1-producer</td>
<td>99</td>
<td>99</td>
</tr>
</tbody>
</table>

Results

Two internal DNA fragments of cnf2 were constructed for use as probes for CNF-producing strains: a 875-bp XhoI-PstI fragment isolated from pEOSW3 and an adjacent 335-bp PstI-ClaI fragment isolated from pEOSW1 (fig. 1). Three pairs of Vir transconjugant strains and a collection of 185 E. coli strains isolated from calves, pigs, cats, dogs and human patients with diarrhoea or extra-intestinal infections were used to test the sensitivity and specificity of the detection of CNF-producing strains by colony hybridisation. The HeLa cell cytotoxic assay in combination with seroneutralisation was the reference assay for the detection of CNF1 and CNF2. This standard assay was used to divide the collection of E. coli strains into non-CNF, CNF1, and CNF2 producers.
DETECTION OF CNF1 AND CNF2 OF *E. coli* 431

Fig. 2. Autoradiographs of two identical blots of 48 *E. coli* strains isolated from calves with diarrhoea or cases of septicaemia. C1, CNF2-positive control; C2, CNF-negative control; C3, CNF1-positive control. Panel A, colony hybridisation with the *PstI-ClaI* fragment of pEOSW1; B, colony hybridisation with the *XhoI-PsrI* fragment of pEOSW3. The two isolates that gave black spots with both probes were identified as CNF2-producers in the seroneutralisation assay. The 26 isolates that gave grey spots with the probe derived from pEOSW1 were identified as CNF1-producers in the seroneutralisation assay. The 22 other isolates were CNF-negative in the cytotoxic assays.

Fig. 3. Hybridisation of different Vir plasmids of *E. coli* with the probe derived from pEOSW1. A, agarose gel electrophoresis of plasmid extracts of eight *E. coli* strains; B, autoradiograph of the same gel hybridised with the probe derived from pEOSW1. The plasmid DNA preparations were from the following *E. coli* strains: lane 1, S5, a CNF2-producer; 2, 712, a K-12 strain; 3, H209(pVir), a CNF2-producer; 4, H209, a human normal flora isolate; 5, 712(pL1), a CNF2-producer; 6, JL21vir, CNF negative; 7, JL21vir*, a CNF2-producer; 8, strain 712.

*coli* clinical isolates into 64 non-CNF producers, 67 CNF1-producers, and 54 CNF2-producers. Results of colony hybridisation experiments with these 185 *E. coli* strains indicated that the 875-bp *XhoI-PsrI* fragment specifically hybridised with the CNF2-producing strains, whereas the adjacent 335-bp *PsrI-ClaI* fragment hybridised with both CNF1-producing and CNF2-producing strains (table I). By selecting one or
the other of these hybridisation responses as the positive response criteria, CNF2-producing strains were detected with a sensitivity and specificity of 100%, and CNF1-producing strains with a sensitivity and specificity of 99% (table II). Only one CNF1-producing strain (115KH89) was not detected by any probe and one non-CNFP producer (43KH89) hybridised with the 335-bp PstI-ClaI fragment.

An example of the results observed in colony hybridisation experiments is shown in fig. 2 where two identical blots of 48 E. coli strains isolated from calves with diarrhoea or cases of septicaemia were hybridised with each CNF probe. The autoradiograph in fig. 2 (panel A) depicts colony hybridisation results with the 335-bp PstI-ClaI fragment. The CNF2-positive control (C1) gave a black spot, the CNF-negative control (C2) gave no spot and the CNF1-positive control (C3) gave a grey spot. Two CNF2-positive isolates gave black spots, 26 CNF1-positive isolates gave grey spots, and 22 negative isolates gave no spot at all. The autoradiograph in fig. 2 (panel B) is the result of colony hybridisation with the 875-bp XhoI-PstI fragment. The CNF2-positive control (C1) gave a black spot. The CNF-negative control (C2) and the CNF1-positive control (C3) gave no spots. The two CNF2-positive isolates gave black spots with the 875-bp XhoI-PstI fragment, the 26 CNF1-positive isolates gave very faint or no spots, and the 22 negative isolates gave no spot at all.

To further characterise the CNF probes as genetic tools, both were tested by in-situ hybridisation against plasmid DNA from eight E. coli strains that did or did not contain the CNF2-encoding Vir plasmids pJL1 or pVir. Results obtained with the 335-bp fragment are presented in fig. 3. A strong signal was specifically associated with the presence of pJL1 or pVir. Similar results were obtained with the 875-bp pEOSW3 fragment as a probe (data not shown). The mol. wt of pVir and pJL1 was estimated to be c. 85 MDa. However, a slower electrophoretic mobility of pJL1 was observed when the plasmid was extracted from the K12 strain 712.

Discussion

The results of colony hybridisation with cnf2 DNA fragments derived from pEOSW1 and pEOSW3 indicated that these fragments were both sensitive and specific as probes for detecting CNF1- and CNF2-producing strains of E. coli. A positive response with both probes was associated with CNF2-producers, whereas a positive response with the probe derived from pEOSW1 alone was associated with CNF1-producers. Because the seroneutralisation assay confirmed the phenotype of the strains detected with the two DNA probes derived from pEOSW1 and pEOSW3, we believe that these cnf probes can be used in place of animal assays and cell-culture assays for the detection of CNF-producing strains.

For some E. coli strains, colony hybridisation results may be more accurate than the cell cytotoxicity assay. For example, one isolate that produces a Shiga-like toxin (verotoxin) reacted strongly with both probes. However, no multineucleation effect was detectable on the cell cytotoxicity assay because the cell monolayer was destroyed by the Shiga-like toxin. By neutralising the toxic effect of SLT with a specific antiserum, we were able to confirm that this strain also produced a fully active CNF2 cytotoxin. This strain is the only published example of a strain producing both of these cytotoxins. The significance of this observation in relation to pathogenesis is unclear. Characterisation of CNF2-producing strains has shown previously that CNF2-producing strains do not constitute a homogeneous group of E. coli, a finding in contrast to those E. coli isolates that produce CNF1. This difference in CNF1 and CNF2 distribution amongst E. coli strain types may be related to the fact that CNF2 is encoded by a transferable F-like plasmid whereas CNF1 is chromosomally encoded. However, even with the potential mobility of cnf2, the production of both CNF2 and SLT by the same strain remains a rare event.

Both probes were also used successfully to detect the large Vir plasmids coding for CNF2. The estimated mol. wt of pVir and pJL1 (85 MDa) is similar to the 92 MDa reported previously. Curiously, we noticed a slight variation in the electrophoretic mobility of pJL1 when the plasmid was extracted from the E. coli K12 strain 712. This variation may reflect a difference in pJL1 supercoiling when the plasmid is resident in recipient E. coli K12 strain 712 compared to wild-type E. coli strain JL21. Alternatively, the plasmid may have undergone some spontaneous deletion in the wild-type background. To test this possibility, we are in the process of analysing the restriction maps of both forms of pJL1.

The sequences of the CNF toxin genes are 85.7% identical. The results of this study confirmed the close relationship at the DNA level between CNF1 and CNF2. Indeed, a 335-bp internal fragment of the gene coding for CNF2 identified CNF1-producing strains. However, CNF1-producing strains did not hybridise with the second probe, an adjacent 875-bp internal fragment of cnf2. Computer analysis showed less mismatching between the smaller DNA probe and cnf1: the 335-bp and the 875-bp DNA probes are, respectively, 88.2% and 86.8% identical to similar fragments of cnf1. Recently, Falbo et al. reported that they were unable to detect three CNF2-producing strains with four large fragments of cnf1. Little information about the hybridisation conditions used by these investigators was given, but two of their DNA probes contained part of the upstream and downstream regions of the CNF1 toxin gene. These regions diverge between cnf1 and cnf2. Since cnf1 and cnf2 have a low GC content of 35%, we think that both the size of the probes and the hybridisation conditions (absence of formamide in our hybridisation solution)
are critical parameters for the detection of CNF-producing strains by colony hybridisation.

We are now using the two DNA probes routinely for the detection of CNF-producing strains. As colony blots can be duplicated easily and used with other DNA probes, we are testing a large collection of E. coli isolates with DNA probes for CNFs and other potential virulence factors. Our preliminary results confirm that CNF1 and α-haemolysin production are closely linked\(^7^{14}\) and that a majority of CNF2-producing strains also produce an F17-like adhesin.\(^8^{5}\) Large scale use of these probes will greatly facilitate estimation of the incidence of CNF-producing strains among clinical isolates of E. coli.

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References


