Proteus mirabilis strains of diverse type have IgA protease activity

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Summary. A strain of Proteus mirabilis associated with chronic urinary tract infection was found to produce an EDTA-sensitive IgA protease that cleaved the IgA heavy chain into two fragments at sites different from those attacked by other microbial IgA1 proteases. Another 14 P. mirabilis strains of diverse type and from various clinical conditions also produced a similar IgA protease. This enzyme may be a virulence determinant of P. mirabilis.

Introduction

Although infections of the urinary tract with Proteus mirabilis occur less frequently than those with Escherichia coli except in young boys (Bergstrom, 1972; Hallett et al., 1976; Khan et al., 1978) and the elderly (Walkey et al., 1967; Senior, 1979), the consequences may be more serious. The predilection of P. mirabilis is for the upper urinary tract (Fairley et al., 1971; Svandborg Edén et al., 1980) where damage to the renal tubular epithelium (Braude et al., 1960) and stone formation (Griffith et al., 1973) may take place.

The virulence of P. mirabilis for the urinary tract may arise through the interplay of several factors including rapid growth in urine and the formation of potent urease isoenzymes which are important in human and experimental urinary tract infections (MacLaren, 1968 and 1969; Musher et al., 1975; Senior et al., 1980; Senior, 1983a). Motility also appears to be important in establishing ascending pyelonephritis (Pazin and Braude, 1974). P. mirabilis cells, unlike those of E. coli, have the ability to directly invade mammalian cells (Braude and Siemieniski, 1960; Peerbooms et al., 1984) possibly through the activity of a cell-bound haemolysin (Peerbooms et al., 1983). The presence of certain fimbriae (Silverblatt, 1974; Silverblatt and Ofek, 1978), proticines and proticine receptors (Senior, 1979) may also be associated with P. mirabilis virulence but the relative importance of these factors is still unclear (Adegbola et al., 1983; Senior, 1983b).

In this study the ability of P. mirabilis strains associated with urinary tract infections to produce a new type of IgA protease was demonstrated. Production of this enzyme in vivo may enable P. mirabilis to overcome the antibody defence of the urinary tract mucosal surfaces resulting in the establishment of infection.

Materials and methods

Bacteria

Single test strains of Haemophilus influenzae, Neisseria gonorrhoeae and Streptococcus pneumoniae were originally isolated from clinical specimens and had been maintained for several months on cooked blood agar. Fifteen strains of Proteus mirabilis (see table) were selected from a wide variety of clinical sources and places and were of diverse bacteriocine (P/S) and O antigenic types. The strains were identified according to standard methods (Senior and Leslie, 1986) and stored on nutrient-agar slopes at 4°C. P/S typing and O serotyping of strains were performed as previously described (Larsson and Olling, 1977; Senior, 1977; Senior and Larsson, 1983).

Media

Nutrient Agar (Oxoid CM 3) and blood agar (BA) composed of Columbia Agar Base (Oxoid CM 331) with sterile horse blood (Oxoid SR 50) 5% v/v were prepared and sterilised according to the manufacturer's instructions. Cooked blood agar (CBA) was heated BA. Modified New York City Medium (MNYC) was prepared according to the method of Young (1978) but with horse blood (Oxoid SR 50).

Preparation of purified 125I-labelled IgA

IgA was purified from frozen normal human plasma. After thawing, calcium chloride was added to a concentration of 20 mM and fibrin clots were removed by

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filtration through glass wool. Ammonium sulphate was added to the resulting serum to 50% saturation. After standing overnight at 4°C, the precipitate was collected by centrifugation, dissolved in distilled water, applied to a G 200 Sephadex column and eluted with 50 mM Tris-saline pH 7.4. The resultant pure IgA was labelled with 125I by the chloramine-T method of Hunter and Green-wood (1962).

**Preparation of IgA protease**

Circular disks (approximately 8-5 cm diameter) cut from dialysis tubing were sterilised in water at 121°C for 15 min and placed flat on to the surface of plates of culture media (BA for *P. mirabilis* strains, CBA for *H. influenzae* and *S. pneumoniae*, and MNYC for *N. gonorrhoeae*) by the use of a sterile cotton-wool swab. The surface of the membrane was inoculated with a cotton-wool swab containing one of the bacterial strains and the plates were incubated overnight at 37°C in the presence of CO2 8%. Bacteria were then scraped from the membrane with a microscope slide and transferred to a microcentrifuge tube. The membrane was rinsed with 0.5 ml of 5 mM Tris-HCl buffer pH 7.2 which was also added to the tube. The cells were emulsified in the buffer, centrifuged for 2 min at 11 600 g and the clear supernate containing IgA protease removed.

**Digestion of 125I IgA with IgA protease**

125I IgA (25 μg/ml of phosphate-buffered saline, pH 7.4) was incubated in 2.5-μl amounts with 50 μl of IgA protease preparations in 5 mM Tris-HCl buffer, pH 7-2, for 20 h at 37°C. EDTA at a final concentration of 5 mM or phosphoramidon (Sigma, Dorset, England) at a final concentration of 2 μM was added to some reactions. After incubation, 25 μl of buffer (0.1 M Tris-HCl, pH 8.0, containing 80 mM dithiothreitol, 8% urea, SDS 2% w/v and a trace of bromophenol blue dye) was added to each sample. After standing in boiling water for 2 min the samples were subjected to electrophoresis.

**Polyacrylamide gel electrophoresis and autoradiography**

Test samples and standard protein mol.-wt markers were analysed by SDS discontinuous polyacrylamide gel electrophoresis (PAGE) by the method of Laemmli (1970). The resolving gel was a linear gradient of 5-22.5% w/v acrylamide and electrophoresis was performed at 40 mA until the dye front reached the bottom of the gel. Gels were stained in Coomassie Brilliant Blue R250 0-125% w/v in methanol 50% v/v and glacial acetic acid 20% v/v in water for 1-2 h. They were subsequently destained in methanol 30% v/v and glacial acetic acid 10% v/v in water, dried under vacuum, and exposed to X-Omat AR film at −70°C. After an appropriate period the film was developed.

**Results**

The PAGE analyses of the products of 125I IgA digestion with enzymes produced by *P. mirabilis* strain 64676—a strain associated with chronic urinary tract infections—and those of *N. gonorrhoeae*, *H. influenzae* and *S. pneumoniae* are shown in fig. 1. The *P. mirabilis* enzyme cleaved the IgA heavy chain at positions different from the hinge region cleavage sites of the other bacterial IgA.

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**Table. Characteristics of the *P. mirabilis* strains examined**

<table>
<thead>
<tr>
<th>Strain no.</th>
<th>P/S type</th>
<th>Source and clinical symptoms</th>
<th>O type</th>
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<tr>
<td>1 6476</td>
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<td>O 28</td>
<td>Dundee</td>
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<tr>
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<td>0/7</td>
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<td>O 41</td>
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<tr>
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<td>O 27</td>
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<tr>
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<tr>
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Fig. 1. Polyacrylamide gel analysis of $^{125}$I IgA digested with various microbial IgA proteases. The *P. mirabilis* protease is EDTA sensitive and cleaves the IgA heavy chain at sites different from those attacked by other bacterial proteases. Left; position of mol. wt standards from top to bottom; bovine albumin (66 000), ovalbumin (45 000), glyceraldehyde-3-phosphate dehydrogenase (36 000), carbonic anhydrase (29 000), trypsinogen (24 000), trypsin inhibitor (20 100), $\alpha$ lactalbumin (14 200). Tracks contain $^{125}$I IgA cleaved with enzymes as follows: 1. control untreated IgA—H = heavy chain, L = light chain; 2. *P. mirabilis* strain 64676 protease; 3. *P. mirabilis* strain 64676 protease with 5 mM EDTA; 4. *P. mirabilis* strain 64676 protease with 2 $\mu$m phosphoramidon; 5. *N. gonorrhoeae* protease; 6. *N. gonorrhoeae* protease with 5 mM EDTA; 7. *H. influenzae* protease; 8. *H. influenzae* protease with 5 mM EDTA; 9. *S. pneumoniae* protease; 10. *S. pneumoniae* protease with 5 mM EDTA.

proteases. The mol. wts of the *P. mirabilis*-cleaved fragments were approximately 47 000 and 34 000. The *P. mirabilis* enzyme was inhibited by 5 mM EDTA but not by phosphoramidon, an inhibitor of several metalloproteinases (Umezawa and Aoyagi, 1977). The IgA protease of *S. pneumoniae* was also EDTA sensitive but the IgA proteases of *N. gonorrhoeae* and *H. influenzae* were not inhibited by 5 mM EDTA.

The kinetics of IgA degradation by the enzymes from *P. mirabilis* strain 64676 are shown in fig. 2. The IgA heavy chain (mol. wt 68 000) appeared to be cleaved to a fragment of mol. wt 47 000 in a single step, because larger fragments of intermediate size were not seen before the formation of the 47 000-mol. wt fragment. The kinetics of formation of the fragment of 34 000 mol. wt appeared to be synchronous with that of the fragment of mol. wt 47 000. This suggested that the smaller fragment did not arise by secondary proteolysis of the larger fragment. Cleavage products of the IgA heavy chain smaller than mol. wt 34 000 were not observed.

Fifteen strains of *P. mirabilis* (see table), with 14 distinct P/S types and at least seven different O antigens, isolated from patients with various clinical symptoms and from healthy carriers in several geographical locations, all produced an IgA protease (fig. 3). The products of IgA cleavage by the enzyme from each strain were of the same size as those generated by the IgA protease from *P. mirabilis* strain 64676.

Discussion

IgA proteases are a group of enzymes that specifically cleave the heavy chain of the IgA subclass of human immunoglobulin A (IgA) at sites
within a 16-amino-acid proline-rich polypeptide in the hinge region. This polypeptide sequence is absent from IgA2 which, as a consequence, is resistant to the protease. IgA1 proteases are stably produced in vitro and in vivo (Sorensen and Kilian, 1984) by several human pathogens (Kilian et al., 1983a), including S. pneumoniae (Kilian et al., 1979; Male, 1979; Mulks et al., 1980a), N. gonorrhoeae (Plaut et al., 1975; Blake and Swanson, 1978), N. meningitidis (Plaut et al., 1975; Mulks et al., 1980b) and H. influenzae (Kilian et al., 1979 and 1983b; Male, 1979; Mulks et al., 1980a). Non-pathogenic members of the same genera do not produce IgA proteases (Mulks and Plaut, 1978; Kilian et al., 1979; Male 1979) which suggests that this enzyme may be involved in virulence.

While investigating virulence mechanisms of P. mirabilis for the urinary tract, we found that the uropathogenic strain 64676 produced an enzyme able to cleave the IgA heavy chain to a fragment of mol. wt 47 000. Removal of carbohydrate from the heavy chain by glycosidase activity would not result in such a reduction in mol. wt, whereas deglycosylation of the several carbohydrate chains would result in the formation of cleaved products intermediate in size between intact heavy chain and the observed final cleavage product. Such intermediates were not observed, which suggests that the enzyme was a protease rather than a glycosidase. There are also several possible explanations for the formation of two fragments from the heavy chain, but further studies are required before any other conclusions are made on the nature of this enzyme.

Since the size of the fragments cleaved by the enzymes from the other 14 strains of P. mirabilis of diverse type were indistinguishable from those cleaved by the P. mirabilis strain 64676 IgA protease, it appears that most strains of P. mirabilis produce a similar type of IgA protease. IgA protease production by P. mirabilis may, therefore, be an important virulence factor, not only in urinary tract infections but in infections at other sites.
We confirmed previous findings (Kilian et al., 1983a) that the IgAl proteases of *N. gonorrhoeae* and *H. influenzae* are relatively resistant to EDTA whereas that of *S. pneumoniae* is EDTA sensitive. The *P. mirabilis* IgA protease was metal chelator-sensitive but differed from other IgAl proteases by cleaving IgA outside the hinge region. It may, therefore, also have activity on IgA2 which would be advantageous to a pathogen.

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