Short Communication

Identification of DNA sequences that imply a novel gammaherpesvirus in seals

Rogier Bodewes,1† Guillermo J. Sánchez Contreras,2† Ana Rubio García,2 Rebriarina Hapsari,1 Marco W. G. van de Bildt,1 Thijs Kuiken1 and Albert D. M. E. Osterhaus1,3,4,5

Correspondence
Rogier Bodewes
r.bodewes@erasmusmc.nl

1Department of Viroscience, Erasmus MC, Rotterdam, the Netherlands
2Seal Rehabilitation and Research Centre, Pieterburen, the Netherlands
3Viroclinics Biosciences BV, Rotterdam, the Netherlands
4Centre for Infection Medicine and Zoonoses Research, University of Veterinary Medicine, Hannover, Germany
5Artemis Research Institute for Wildlife Health, Utrecht, the Netherlands

Various herpesviruses have been discovered in marine mammals and are associated with a wide spectrum of disease. In the present study we describe the detection and phylogenetic analysis of a novel gammaherpesvirus, tentatively called phocine herpesvirus 7 (PhHV-7), which was detected in samples collected during an outbreak of ulcerative gingivitis and glossitis from juvenile harbour seals (Phoca vitulina) at the Seal Rehabilitation and Research Centre, the Netherlands. The presence of this novel gammaherpesvirus was confirmed by viral metagenomics, while no other viruses other than four novel anelloviruses were detected. However, PhHV-7 DNA was also detected in harbour and grey seals (Halichoerus grypus) without gingivitis or glossitis. Genetic analysis of the partial polymerase gene of PhHV-7 detected in both species revealed limited sequence variation. Additional studies are needed to elucidate whether the viruses discovered played a role in the disease observed.

MAIN TEXT

Herpesviruses are large, enveloped viruses with a single, linear, double-stranded DNA genome. The order Herpesvirales currently consists of three distinct families, Herpesviridae, with viruses of mammals, birds and reptiles, Alloherpesviridae with viruses of fish and frogs and Malacoherpesviridae with viruses of molluscs. The family Herpesviridae is further subdivided into subfamilies Alphaherpesvirinae, Betaherpesvirinae and Gammaherpesvirinae (ICTV, 2013). In 1985, the first herpesvirus in marine mammals, phocine herpesvirus 1 (PhHV-1), was discovered in harbour seals (Phoca vitulina) with pneumonia and hepatitis (Osterhaus et al., 1985). In addition to PhHV-1, more than five herpesviruses are currently described in phocids (Kuiken & Das Neves, 2012; Maness et al., 2011; Venn-Watson et al., 2012). PhHV-1 is to date the only alphaherpesvirus described, while the other five herpesviruses from phocids are classified as gammaherpesviruses.

PhHV-2 was first isolated from leukocytes of harbour seals in 1994 (Harder et al., 1996; Lebich et al., 1994). PhHV-3 was detected in Hawaiian monk seals (Monachus schauinslandi), PhHV-4 was detected in northern elephant seals (Mirounga angustirostris) with ulcerative lesions of the skin and mucosa, and PhHV-5 and PhHV-6 in harbour seals (Goldstein et al., 2006a, b; Maness et al., 2011).

In the present study, we describe an outbreak of ulcerative gingivitis followed by ulcerative glossitis among harbour seals associated with infection with a novel gammaherpesvirus. During April and May 2014, nine juvenile (between 8 and 10 months of age) harbour seals that were being rehabilitated at the Seal Rehabilitation and Research Centre (SRRC) in Pieterburen, the Netherlands, developed initially diffuse reddening and swelling of the gums in both the upper and lower jaw, mild bleeding, retraction of the gingiva exposing the roots of the teeth, dark staining of the teeth and halitosis (Fig. 1a). In a second stage of the disease, the animals developed lingual ulcerations that were oval, about 10 mm in diameter, 2–4 mm in depth and dark red in colour (Fig. 1b).

Seals that developed gingivitis had initially been admitted to the SRRC during the winter season 2013–2014 due to parasitic pneumonia, but were all successfully treated against
lungworm infection. The seals that developed gingivitis were kept in three different enclosures, although eight animals shared one pool together (Table S1, available in the online Supplementary Material). Seals showing gingivitis and glossitis were empirically treated with anti-inflammatory drugs, prophylactic antibiotic treatment and antiseptic mouthwashes. After 1 week, most of the lesions resolved.

Based on the sudden onset of disease, the observed non-suppurative lesions, the course of the outbreak and absence of apparent problems with water quality and/or nutritional deficiencies, a virus infection was suspected and swab samples were collected from ulcers present on the tongue of affected animals and from the oral mucosa. Swabs were collected in Hank’s balanced salt solution containing 0.5 % lactalbumin, 10 % glycerol, 200 U ml⁻¹ penicillin, 200 µg streptomycin ml⁻¹, 100 U polymyxin B sulfate ml⁻¹, 250 µg gentamicin ml⁻¹ and 50 U nystatin ml⁻¹ (ICN Pharmaceuticals; transport medium) and stored at −70 °C until further processing. Upon defrosting, swab samples were vortexed briefly and nucleic acids were extracted from the supernatant using the High Pure viral nucleic acid kit (Roche). Subsequently, a pan-herpesvirus PCR was performed of which the second PCR was altered by using primers DFA and IYG as described previously (VanDevanter et al., 1996; Venn-Watson et al., 2012). A PCR fragment of the expected size was obtained from swab samples collected from animals with acute clinical signs. This PCR fragment was ligated into the pCR4 Topo vector (Invitrogen) and sequences of at least three clones in two directions were obtained as described previously (van Leeuwen et al., 2010). Analysis of the sequence fragments obtained by BLAST revealed the presence of a novel gammaherpesvirus, tentatively called phocine herpesvirus 7 (PhHV-7).

To exclude the presence of viral co-infections, sequence-independent RNA and DNA virus screening of four tongue swabs from four different animals, and one oral mucosal swab and one serum sample from one of these animals (Table S2), was performed as described previously (van den Brand et al., 2012; van Leeuwen et al., 2010). Reads obtained were analysed by BLAST and BLASTx as described previously (Schürch et al., 2014), and sequences most closely related to various genes of gammaherpesviruses were detected in all swabs but not in the serum sample (Table S2). In addition, sequences that were most closely related to anelloviruses were detected in the serum sample and in two swabs, while no other viral sequences were detected. Based on reads obtained by next-generation sequencing, additional specific primers were designed and the partial polymerase gene (1026 nt; GenBank accession number KM262784) of PhHV-7 was obtained from seal PV13-431 as described previously (van Leeuwen et al., 2010). Alignment and phylogenetic analysis of the partial DNA polymerase gene of PhHV-7 PV13-431 with various other gammaherpesviruses using MAFFT version 7 (Katoh & Standley, 2013) and MEGA6 (Tamura et al., 2013) indicated that this virus was most closely related to PhHV-3, with pairwise identities of 75 % at the nucleotide level and 86 % at the amino acid level (Fig. 2, Table S3). According to the criteria of the International Committee on Taxonomy of Viruses (ICTV) (Pellett et al., 2012), a novel herpesvirus species should have a distinct genome and epidemiological or biological characteristics. Since there was considerable sequence difference between PhHV-7 and the most closely related viruses (PhHV-3 and PhHV-4), and these viruses were detected in different species of seals, PhHV-7 may be classified as a novel species. However, only limited sequence information is available for most herpesviruses detected in marine mammals, and additional sequence information is necessary to further understand the phylogenetic relationships of these viruses.

Of interest, gammaherpesviruses have been associated with ulcerative lesions of the skin and mucosa in a number of animal species, among which are several carnivore species including northern sea otters (Enhydra lutris kenyonii), California sea lions (Zalophus californianus), northern elephant seals (Mirounga angustirostris) and a fisher (Martes pennanti) (Gagnon et al., 2011; Goldstein et al., 2006b; Smith et al., 2008; Smolarek Benson et al., 2006; Tseng et al., 2012; Venn-Watson et al., 2012). In humans infected with human immunodeficiency virus, human herpesvirus 4 (Epstein–Barr virus) can cause oral hairy leukoplakia and both human herpesviruses 4 and 8 were detected in oral ulcers (Di Alberti
et al., 1997; Hille et al., 2002; Syrjänen et al., 1999). Although gammaherpesviruses are generally considered lymphotropic and are often associated with lymphoproliferative disease (Ackermann, 2006), these studies suggested that infection with certain gammaherpesviruses, in combination with a stressor or underlying disease, can induce the development of ulcerative lesions. To elucidate the putative role of PhHV-7 in the ulcerative gingivitis and glossitis observed, and to determine its prevalence among seals present in the SRRC, oral mucosal swabs were collected from all seals with gingivitis (n=9) and two additional control groups with no signs of gingivitis (Table S1) and tested by use of a PhHV-7-specific PCR. The first control group consisted of 14 harbour seals in the same age range, and reason for stranding, as the group with gingivitis. Two of these animals were kept in the same enclosure as the animals with gingivitis, while the others were kept in other enclosures. The second control group consisted of eight juvenile grey seals (Halichoerus grypus) that were present at the SRRC at the same time the outbreak of gingivitis occurred. These grey seals were held in other enclosures than the group with gingivitis. Nucleic acids extracted from the swabs were tested for the presence

**Fig. 2.** Phylogenetic analysis of the partial polymerase gene of PhHV-7. The amino acid sequence of the partial polymerase gene of PhHV-7 detected in PV13-431 was aligned with various other herpesviruses. Subsequently, an LG (Le and Gascuel) + G + I tree was built by analysis with PROTEST 3.0 (Darriba et al., 2011) based on an alignment of 450 nt in MEGA6 with 500 bootstrap replicates. Only bootstrap values >70 are shown. GenBank accession numbers: alcephaline herpesvirus 1, AF005370; asinine herpesvirus 5, KC825357; ateline herpesvirus 3, AF083424; atlantic bottlenose dolphin gammaherpesvirus, AY952776; bovine herpesvirus 6, KJ705001; caprine herpesvirus 2, HQ116812; dwarf sperm whale gammaherpesvirus, AY949830; equid herpesvirus 2, NC_001650; equid herpesvirus 5, GQ325597; human herpesvirus 1, P04293; human herpesvirus 4, NC_007605; human herpesvirus 8, U93872; macacine herpesvirus 4, NC_006146; macacine herpesvirus 5, NP_570750; mustelid herpesvirus 1, AF376034; otarid herpesvirus 1, AF236050; otarid herpesvirus 3, JX080682; phocine herpesvirus 1, U92269; phocine herpesvirus 2, GQ429152; phocine herpesvirus 3, DQ093191; phocine herpesvirus 4, DQ183057; phocine herpesvirus 5, GQ429153; phocine herpesvirus 7, KM262784; saimiriine herpesvirus 2, X64346.
of PhHV-7 DNA by PCR using forward primer GGATTTAATCCGATATCTGGAG and reverse primer CTGCAGATGGCTAGGGTTTC to amplify a fragment (138 nt) of the partial polymerase gene of PhHV-7. Nucleic acids extracted from transport medium were used as a negative control in the PCR. A PCR fragment of the expected size was detected in seven of nine swab samples collected from the animals with ulcerative gingivitis, while PhHV-7 DNA was also detected in 12 of 14 swab samples collected from the harbour seal control group and in seven of eight swab samples collected from the grey seal control group. Sequence analysis of a proportion of the PCR products obtained indeed confirmed the presence of PhHV-7 DNA. The persistence of PhHV-7 infection was evaluated by testing swab samples collected from four harbour seals that were still present at the SRRC five weeks after the outbreak with gingivitis (Table S1). In two of the four animals tested, PhHV-7 DNA was detected by PCR, which showed that PhHV-7 remained present in seals at the SRRC for more than 1 month.

To elucidate whether sequence variation exists between PhHV-7 detected in grey seals and harbour seals, the nucleotide sequences of the partial polymerase gene (816 nt) of PhHV-7 detected in six harbour seals (PV13-564, PV13-592, PV13-565, PV14-099, PV14-165, PV14-169) and three grey seals (HG13-545, HG13-635, HG14-126) were obtained by PCR using forward primer CTGCTACTCCACCATGATTTCAG and reverse primer GTGAGGTCTGATATTTCCACGTG. Alignment and phylogenetic analysis of the PCR fragments obtained indeed revealed variation in the nucleotide and deduced amino acid sequence of PhHV-7, not only in harbour and grey seals but also within PhHV-7 detected in harbour seals (Figs 3 and S1). Pairwise identities on the nucleotide level between PhHV-7 in grey and harbour seals ranged between 97.3 and 98.6 %, while pairwise identities at the amino acid level ranged between 98 and 98.9 %. Of interest, sequence variation between other DNA viruses (phocine herpesvirus 3 and 4, and a HKV + G (Hasegawa–Kishino–Yano) tree was built as determined by JModelTest 2.1.3 (Darriba et al., 2012) based on an alignment of 409 nt in mEGA6 with 500 bootstrap replicates. Only bootstrap values >70 are shown. PhHV-7 detected in animals with ulcerative gingivitis and glossitis is indicated with a G. GenBank accession number: phocine herpesvirus 3, DQ093191; phocine herpesvirus 4, DQ183057; phocine herpesvirus 7 PV13-431, KM262784; phocine herpesvirus 7 PV13-564, KP163991; phocine herpesvirus 7 PV13-565, KP163992; phocine herpesvirus 7 PV13-592, KP163993; phocine herpesvirus 7 PV14-099, KP163994; phocine herpesvirus 7 PV14-165, KP163995; phocine herpesvirus 7 PV14-169, KP163996; phocine herpesvirus 7 HG13-545, KP163997; phocine herpesvirus 7 HG13-635, KP163998; phocine herpesvirus 7 HG14-126, KP163999.

In addition to this novel herpesvirus, the near complete genomes of two novel anelloviruses, tentatively called seal anelloviruses (SeAv) 4 and 5 (SeAv4, 2223 nt, GenBank accession KM262783; SeAv5, 2056 nt, GenBank accession KM262782), and the complete genomes of two other novel anelloviruses, tentatively called SeAv6 and SeAv7 (SeAv6, 2143 nt, GenBank accession KM262785; SeAv7, 2165 nt, GenBank accession KM262781), were obtained by assembly of reads from the deep sequencing data for serum of PV13-431. The genome organizations of the two partially sequenced anelloviruses are similar to a torque tenovirus.
detected in a pine marten (MmTTV1) (van den Brand et al., 2012), while those of the two completely sequenced anelloviruses were similar to anelloviruses previously detected in seals, with three major ORFs and a TATA-box (Bodewes et al., 2013a; Ng et al., 2011) (data not shown). Alignment and phylogenetic analysis of the amino acid sequences of ORF1 of SeAv4-7 with various other anelloviruses, using CLUSTAL OMEGA (Sievers et al., 2011) and MEGA6 (Tamura et al., 2013), revealed that SeAv4 and SeAv5 were most closely related to each other (65% identity at the nucleotide level and 64% at the amino acid level) and to MmTTV1 (46% identity at the nucleotide level and 32–34% at the amino acid level). SeAv6 was most closely related to SeAv3 and SeAv7 (67–68% identity at the nucleotide level and 60% at the amino acid level), while SeAv7 was also very closely related to SeAv3 (94% identity at the nucleotide level and 97% at the amino acid level) (Fig. S3, Table S4). Genera within the Anelloviridae are currently defined as having less than 44% identity at the nucleotide level and to MmTTV1 (46% identity at the nucleotide level and 64% at the amino acid level), SeAv5 were most closely related to each other (65% identity at the nucleotide level and 60% at the amino acid level), while SeAv7 was also very closely related to SeAv3 (94% identity at the nucleotide level and 97% at the amino acid level) (ICTV, 2013). This indicates that SeAv 4 and 5 belong to a novel species within a putative novel genus that currently consists solely of MmTTV1. SeAv6 and SeAv7 belong to the same species as SeAv3. Anelloviruses have not been associated with ulcerative lesions in humans and animal species (Hino & Miyata, 2007; Ng et al., 2011; Toyoda et al., 2000), and additional studies are necessary to elucidate the role of these viruses in the lesions observed in these harbour seals.

In conclusion, a novel herpesvirus and four novel anelloviruses were identified during an outbreak of ulcerative gingivitis and glossitis among harbour seals. Additional studies are necessary to understand the pathogenesis and epidemiology of infections with PhHV-7 and other gammaherpesviruses in various species of seal.

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**References**


