JC virus granule cell neuronopathy is associated with VP1 C terminus mutants

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The polyomavirus JC (JCV) infects glial cells and causes progressive multifocal leukoencephalopathy (PML). We described a novel JCV-variant with a 10 bp deletion in the C terminus of the VP1 capsid protein, JCV_{GCN1}. This mutant was associated with lytic infection of cerebellar granule cell neurons and cerebellar atrophy in an human immunodeficiency virus/PML patient. This condition, also observed independently from PML, was named JCV granule cell neuronopathy (JCV GCN). We characterized JCV mutations in cerebrospinal fluid (CSF) of four other JCV GCN patients, and reviewed the literature on 10 reported cases. The strain from one patient harboured the identical GCN1-deletion, while the other patients had novel mutations in the same area, named JCV_{GCN2–4}, causing variable changes in VP1 structure. One patient also had wild-type JCV in the CSF. To study the mechanisms leading to JCV GCN, we compared viral replication kinetics from JCV_{GCN1} with the prototype JCV_{Mad1}, the PML isolate JCV_{HWM} and the prototype JCV_{Mad1D} engineered with the GCN1-deletion. While all strains replicated at low levels in the medulloblastoma cell line DAOY from a cerebellar neuronal tumour, JCV_{Mad1} replicated better in astroglial SVG cells than JCV_{Mad1D} or JCV_{GCN1} and all strains replicated at higher levels in COS-7 kidney cells, suggesting that the GCN1-deletion confers a disadvantage for viral growth in central nervous system white matter. The GCN1-deletion remained stable after 100 days in culture and VP1 protein was produced in all cell lines, indicating that JCV_{GCN1} is replication-competent in vitro.

These data highlight an important and previously overlooked aspect of JCV-pathogenesis. Detection of GCN-type JCV strains in CSF may help clinicians diagnose JCV GCN.

INTRODUCTION

Infection with the polyomavirus JC (JCV) is widespread and 50–86 % of healthy adults are JCV seropositive (Knowles et al., 2003; Weber et al., 1997). The virus remains quiescent in immunocompetent people, but reactivates in the setting of immunosuppression. It is the aetiological agent of progressive multifocal leukoencephalopathy (PML). We described a novel JCV-variant with a 10 bp deletion in the C terminus of the VP1 capsid protein, JCV_{GCN1}. This mutant was associated with lytic infection of cerebellar granule cell neurons and cerebellar atrophy in an human immunodeficiency virus/PML patient. This condition, also observed independently from PML, was named JCV granule cell neuronopathy (JCV GCN). We characterized JCV mutations in cerebrospinal fluid (CSF) of four other JCV GCN patients, and reviewed the literature on 10 reported cases. The strain from one patient harboured the identical GCN1-deletion, while the other patients had novel mutations in the same area, named JCV_{GCN2–4}, causing variable changes in VP1 structure. One patient also had wild-type JCV in the CSF. To study the mechanisms leading to JCV GCN, we compared viral replication kinetics from JCV_{GCN1} with the prototype JCV_{Mad1}, the PML isolate JCV_{HWM} and the prototype JCV_{Mad1D} engineered with the GCN1-deletion. While all strains replicated at low levels in the medulloblastoma cell line DAOY from a cerebellar neuronal tumour, JCV_{Mad1} replicated better in astroglial SVG cells than JCV_{Mad1D} or JCV_{GCN1} and all strains replicated at higher levels in COS-7 kidney cells, suggesting that the GCN1-deletion confers a disadvantage for viral growth in central nervous system white matter. The GCN1-deletion remained stable after 100 days in culture and VP1 protein was produced in all cell lines, indicating that JCV_{GCN1} is replication-competent in vitro.

These data highlight an important and previously overlooked aspect of JCV-pathogenesis. Detection of GCN-type JCV strains in CSF may help clinicians diagnose JCV GCN.
granule cell neurons in a 39-year-old human immunodeficiency virus (HIV)-seropositive patient with cerebellar atrophy who also had classic lesions of PML in the white matter of both frontal lobes, but no demyelinating lesions in the cerebellum (Du Pasquier et al., 2003; Tyler, 2003). JCV infection of granule cell neurons was confirmed in another HIV-seropositive patient with cerebellar atrophy and no clinical or radiological evidence of PML. This patient had a severe cerebellar syndrome characterized by progressive alteration of coordination, gait instability and ataxia. This novel syndrome, distinct from PML, was named JCV granule cell neuronopathy (JCV GCN) (Koralnik et al., 2005). JCV GCN was subsequently reported as well by others in HIV-positive and -negative patients throughout the world, with or without PML (Table 1) (Du Pasquier et al., 2003; Bustamante et al., 2009; Granot et al., 2009; Hecht et al., 2007; Koralnik et al., 2005; Otis & Moral, 2005; Shin et al., 2008; Tan & Brew, 2009).

Whereas the JCV-coding region is extremely conserved the hypervariable non-coding-regulatory region (RR) has been associated with neurotropism and neurovirulence. Full-length molecular characterization of the strain, named JCV<sub>GCN1</sub>, infecting granule cell neurons in our initial patient, showed a 10 bp out of frame mutation in the C terminus of the VP1 major capsid protein, leading to an alteration of the last 13 aa sequence (Dang & Koralnik, 2006). To investigate further the pathogenic mechanism leading to JCV infection of granule cell neurons, we characterized JCV VP1 sequence variation of four additional JCV GCN patients from the USA and Brazil, and studied the phenotype of JCV<sub>GCN1</sub> in vitro. Each patient had either identical or similar mutations in the JCV VP1 C terminus, and the JCV<sub>GCN1</sub> molecular clone was replication-competent in several cell lines in vitro, but displayed different characteristics than undeleted JCV strains.

RESULTS

JCV strains with mutated VP1 C terminus gene (GCN-type JCV) can be found in every JCV GCN case

The clinical characteristics of the JCV GCN cases we have investigated are listed in Table 1 in chronological order of discovery. The clinical presentation of cases 1–3 has been reported in detail (Dang & Koralnik, 2006; Du Pasquier et al., 2003; Hecht et al., 2007; Koralnik et al., 2005; Tyler, 2003), but the authors of each publication were contacted to provide an update on the clinical outcome of their patients.

Case report patient #4 (Table 1)

A 25-year-old HIV-infected Brazilian woman with multidrug resistance to antiretrovirals presented with 2 months history of right-sided clumsiness and unsteady gait. Neurological examination showed a right cerebellar syndrome. Her absolute CD4<sup>+</sup> T-cell count was 3 mm<sup>−3</sup> and the plasma HIV viral load was 478 130 copies ml<sup>−1</sup> (5.7 log<sub>10</sub>). A computed tomography (CT) scan of the brain demonstrated right cerebellar atrophy (Fig. 1a). Magnetic resonance imaging (MRI) confirmed that this patient exclusively had cerebellar atrophy without PML lesions in cerebellum (data not shown). Cerebrospinal fluid (CSF) analysis was normal and qualitative PCR performed in a clinical laboratory was negative for JCV DNA. Quantitative PCR (QPCR) performed at the Virology Laboratory at Instituto de Medicina Tropical (Sao Paulo, Brazil) was positive for JCV, establishing the diagnosis of JCV GCN. She was treated with antiretroviral medications and 2 months later, the neurological examination was stable, CD4<sup>+</sup> T-cell count was up to 24 mm<sup>−3</sup> and the HIV plasma viral load was down to 916 copies ml<sup>−1</sup> (2.9 log<sub>10</sub>). However, she died soon thereafter from community-acquired pneumonia 4 months after the onset of neurological symptoms. A post-mortem examination was not performed.

Case report patient #5 (Table 1)

A 34-year-old HIV-infected Brazilian woman with a history of poor compliance to antiretrovirals presented with 6 months history of dizziness, unsteady gait and right-sided clumsiness, as well as recent cough and fever. Neurological examination showed a predominantly right cerebellar syndrome. Her absolute CD4<sup>+</sup> T-cell count was 116 cells mm<sup>−3</sup> and HIV viral load was 47 492 copies ml<sup>−1</sup> (4.7 log<sub>10</sub>). A CT scan of the brain demonstrated isolated cerebellar atrophy (Fig. 1b) confirmed by MRI (Fig. 1c). CSF analysis was normal and qualitative PCR was negative for JCV DNA. QPCR performed at the Virology Laboratory at Instituto de Medicina Tropical (Sao Paulo, Brazil) was positive for JCV, however, consistent with the diagnosis of JCV GCN. She was restarted on cART and 9 months after her initial presentation, she had stable CT alterations, but her CD4<sup>+</sup> T-cell count had risen to 221 mm<sup>−3</sup> and her HIV viral load became undetectable (<50 copies ml<sup>−1</sup>). She was lost to follow up 18 months after onset of neurological symptoms. Among 10 reported JCV GCN cases, we could access clinical samples of five cases (1, 2, 3, 4 and 5; Table 1). The JCV VP1 C terminus of each patient was PCR-amplified, cloned and sequenced as described previously (Dang & Koralnik, 2006). GCN-type JCV could be detected in every JCV GCN case available for testing and were named sequentially JCV<sub>GCN1</sub> to JCV<sub>GCN4</sub>. Among these five cases, three (#2, 4 and 5) had no white matter lesions at presentation and therefore, no evidence of PML in addition to JCV GCN (Table 1). GCN-type JCV was the only JCV strain detected in these JCV GCN cases without PML lesions. Alignment of DNA and primary protein sequences of all GCN types, the reference strain JCV<sub>Mad1</sub> and the JCV<sub>HWM</sub> found in the hemispheric white matter of our initial patient are shown in Fig. 2.

As reported previously, JCV<sub>GCN1</sub> has an out of frame 10 nt mutation, resulting in an alteration of the sequence of the
Table 1. Clinical characteristics of all reported JCV GCN cases

MA, Massachusetts; CA, California; NY, New York; cART, Combined Antiretroviral Therapy; NA, not applicable.

<table>
<thead>
<tr>
<th>Case no.</th>
<th>Geographical location</th>
<th>Age (years)</th>
<th>Gender</th>
<th>Underlying diagnosis</th>
<th>PML</th>
<th>JCV strains</th>
<th>VP1 C terminus gene PCR product size (bp)*</th>
<th>Clinical evolution since onset of neurological symptoms</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Boston, MA</td>
<td>39</td>
<td>M</td>
<td>HIV+</td>
<td>+</td>
<td>JCV$<em>{GCN1}$, JCV$</em>{HWM}$</td>
<td>104, 114</td>
<td>Died 6 months later</td>
<td>Du Pasquier et al. (2003)</td>
</tr>
<tr>
<td>2</td>
<td>New York City, NY</td>
<td>43</td>
<td>F</td>
<td>HIV+</td>
<td>−</td>
<td>JCV$_{GCN2}$</td>
<td>108</td>
<td>Wheel-chair dependent, on cART, 13 years later</td>
<td>Koralnik et al. (2005)</td>
</tr>
<tr>
<td>3</td>
<td>San Francisco, CA</td>
<td>15</td>
<td>M</td>
<td>CD-40 ligand deficiency</td>
<td>+</td>
<td>JCV$<em>{GCN3}$, JCV$</em>{SF}$</td>
<td>131, 114</td>
<td>Worsening of cerebellar syndrome. PML lesions in frontal lobes after 21 months with CSF JC viral load $12 \times 10^{6}$ cps ml$^{-1}$ and died after 23 months</td>
<td>Hecht et al. (2007)</td>
</tr>
<tr>
<td>4</td>
<td>Sao Paulo, Brazil</td>
<td>25</td>
<td>F</td>
<td>HIV+</td>
<td>−</td>
<td>JCV$_{GCN4}$</td>
<td>104</td>
<td>Died 4 months later</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>Sao Paulo, Brazil</td>
<td>34</td>
<td>F</td>
<td>HIV+</td>
<td>−</td>
<td>JCV$_{GCN1}$</td>
<td>104</td>
<td>Lost to follow-up 18 months later</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>Springfield, MA</td>
<td>52</td>
<td>F</td>
<td>HIV+</td>
<td>+</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>Otis &amp; Moral (2005)</td>
</tr>
<tr>
<td>7</td>
<td>Seoul, South Korea</td>
<td>37</td>
<td>F</td>
<td>HIV+</td>
<td>−</td>
<td>NA</td>
<td>NA</td>
<td>Bed-ridden 3 years later</td>
<td>Shin et al. (2008)</td>
</tr>
<tr>
<td>8</td>
<td>Sydney, Australia</td>
<td>49</td>
<td>F</td>
<td>Sarcoidosis</td>
<td>+</td>
<td>NA</td>
<td>NA</td>
<td>Died</td>
<td>Granot et al. (2009)</td>
</tr>
<tr>
<td>9</td>
<td>Sydney, Australia</td>
<td>42</td>
<td>M</td>
<td>HIV+</td>
<td>−</td>
<td>NA</td>
<td>NA</td>
<td>Stopped cART and died 2 years later</td>
<td>Tan &amp; Brew (2009)</td>
</tr>
<tr>
<td>10</td>
<td>Providencia, Chile</td>
<td>37</td>
<td>M</td>
<td>HIV+</td>
<td>+</td>
<td>NA</td>
<td>NA</td>
<td>Alive with severe cerebellar dysfunction 2 years later</td>
<td>Bustamante et al. (2009)</td>
</tr>
</tbody>
</table>

*Using PCR primers CJS2465 and CJR2578 described in Dang & Koralnik (2006).
last 13 aa of the protein and was found in both our initial case #1 (Table 1) from Boston (Du Pasquier et al., 2003) as well as case #5 from Brazil. JCVGCN4 from Brazil has the same 10 bp deletion as JCVGCN1 starting at nt 2503 and ending at nt 2512 in the VP1 gene. In addition, JCVGCN4 has five single nucleotide mutations after the 10 bp deletion, leading to 4 aa changes compared with JCVGCN1. JCVGCN2 from New York City has a 6 bp in-frame deletion from nt

Fig. 1. Brain imaging studies of two HIV-seropositive patients with JCV GCN. Head CT scan of patient #4 in Table 1 (a) and head CT and MRI of patient #5 in Table 1 (b and c) showing cerebellar atrophy (arrows) and no demyelinating lesions of PML.

Fig. 2. Alignment of DNA and amino acid sequences of VP1 C terminus from JCV strains identified from different JCV GCN cases. The nucleotide and amino acid position numbers correspond to JCVMad1 prototype sequence. HWM, Hemispheric white matter; GCN1–4, granule cell neurons strains; SF, San Francisco. Dashed lines denote sequence homology. Line interruption denotes deletion. The sequence underlined and in italics in JCVGCN3 is the one that is duplicated in the 17 bp insert at nt 2502.
2511 to 2515, resulting in the loss of aa 348 and 349. Finally, JCV_{GCN3} from San Francisco has a duplication of a 17 bp sequence inserted at nt 2502, leading to a premature stop codon and a truncated VP1 protein lacking the last 9 aa. This patient also had a wild-type JCV strain in his CSF, named JCV_{SF}.

**JCV containing the GCN1 mutation are replication competent in vitro**

To determine whether mutation of JCV VP1 C terminus altered the kinetics of JCV replication *in vitro* we initially focused on the GCN1 mutation found in our index patient as well as one Brazilian case (Table 1). Prototype JCV_{Mad1}, PML isolate JCV_{HWM}, JCV_{GCN1} and prototype JCV_{Mad1D} engineered with the GCN1 deletion were transfected in medulloblastoma cell line DAOY, a tumour arising from the granule cell layer of the cerebellum. In addition, these strains were also transfected in the permissive human astroglial cell line SVG and the monkey kidney cell line COS-7, which have been transformed with JCV-related simian virus 40 (SV40) T Ag, and therefore can sustain JCV replication. Cellular and supernatant DNA were harvested over a period of 100 days. Control experiments showed that *DpnI* digestion of input viral DNA resulted in complete digestion of nucleic acids, ensuring that QPCR only amplified packaged viral DNA. Viral replication kinetics of the various JCV strains in different cell lines was measured by QPCR and is shown in Fig. 3. We used a regression model with pairwise contrast and Bonferroni correction for multiple comparisons to analyse differences in viral kinetics over time between the four viral strains in each cell line, and for each strain in the three different cell lines.

The DAOY cells could sustain replication of all four JCV strains at a similar low level, and allowed productive infection that could be detected in the supernatant throughout the experiment. The presence of the GCN1 mutation did not confer any significant advantage or disadvantage compared to the non-deleted strains (Fig. 3a, b). Conversely, in SVG cells however, JCV_{Mad1} had the highest peak viral load of all four viral strains reaching 8 logs both in the cells and supernatant. JCV_{Mad1} viral load was higher overtime in both cell lysate and supernatant compared with JCV_{GCN1} (*P*=0.004 and *P*<0.0001). Conversely, JCV_{Mad1D} engineered with the GCN1 deletion had similar viral load overtime in cell lysate and supernatant as JCV_{GCN1} (*P*=1.0 and *P*=1.0) and lower replication levels than the undeleted JCV_{Mad1} (*P*=0.09 and *P*=0.003) (Fig. 3c, d).

Finally, all four JCV strains could replicate at significantly higher levels overtime in COS-7 cells compared with the other two cell lines. But the JCV_{GCN1} initially had the slowest replication kinetic compared with the three other strains and a lower viral load in cell lysate than JCV_{Mad1} (*P*=0.03), (Fig. 3e, f).
GCN1 mutation remains stable overtime in vitro

To determine whether the GCN1 mutation persisted over time in various cell lines we amplified the VP1 gene fragment encompassing the mutated area from the cellular DNA after 30 passages in vitro. Compared to the undeleted JCV_Mad1 and JCV_HWM, the JCV_GCN1 and JCV_Mad1D mutation remained stable after 100 days in culture (Fig. 4a) and the size of the amplified fragment was the same as the one amplified from the input virus (not shown). Furthermore, the JCV VP1 protein could be detected by Western blot from cell lysates of all three cell lines infected with either strain, indicating a complete replication cycle and the presence of mature viral particles in these cells (Fig. 4b).

DISCUSSION

Although JCV GCN is a recently discovered CNS condition, it has now been recognized in patients from the USA (Du Pasquier et al., 2003; Hecht et al., 2007; Koralnik et al., 2005; Otis & Moral, 2005), Australia (Granot et al., 2009; Tan & Brew, 2009), Chile (Bustamante et al., 2009), South Korea (Shin et al., 2008), and in this report, Brazil. JCV GCN patients were from both genders, their age ranged from 15 to 52 years and all were immunosuppressed, either in the context of HIV infection, congenital or inflammatory conditions. Among all five JCV GCN cases reported in this study and five described by others (Bustamante et al., 2009; Granot et al., 2009; Otis & Moral, 2005; Shin et al., 2008; Tan & Brew, 2009), half of the cases had co-existent white matter lesions at the time of presentation (case #1, 3, 6, 8 and 10; Table 1). This indicates that JCV GCN can occur separately from, or in association with, PML.

Although clinically recognizable cases of JCV GCN remain rare, the prevalence of JCV infection of granule cell neurons is likely underestimated. A histopathological survey of archival cerebellar samples showed evidence of JCV infection in granule cell neurons of up to 51% of HIV-seropositive PML patients and 3% of HIV-seropositive individuals without PML (Wüthrich et al., 2009). This infection occurred regardless of whether classic PML lesions were also present in the nearby cerebellar white matter. It is therefore possible that many AIDS patients presenting with cerebellar atrophy and a cerebellar syndrome indeed have JCV GCN (Tagliati et al., 1998).

Interestingly, we found three novel GCN-type mutations, all occurring in the same area of the VP1 C terminus as the initial GCN1 deletion (Dang & Koralnik, 2006). JCV-coding region is highly conserved, and these changes had never been described in more than 400 full-length viral sequences deposited in GenBank. It is therefore extremely unlikely that they could have occurred by chance. Furthermore, mutations found in all but the GCN2 variant lead to the loss of a β-pleated sheet in this area as modelled by Expert Protein Analysis System (EXPASY) (data not shown). These changes may probably have important consequences on the structure of the viral capsid, which is formed by 72 pentamers of the VP1 protein, associated on their inner surface with either the VP2 or VP3 protein. Indeed, this region forms the principal interpentamer contact, since each pentamer extends five invading arms to surrounding pentamers and receives five invading arms in return (Stehle et al., 1996). Of note, this area is likely to be exposed on the exterior surface of the capsid, but in a position near the base of the canyons between pentamer knobs, which is remote from the known receptor-binding site on the surface of the pentamers (Neu et al., 2010).

Since mature viral particles were seen by electron microscopy in granule cell neurons of our initial patient (Du Pasquier et al., 2003) these changes may not prevent the formation of the capsid, but rather may influence viral tropism or post-entry events, including uncoating, transport and replication of JCV DNA, which could potentially enable productive infection in granule cell neurons.

Of note, the GCN-type mutations were not located in areas of single nucleotide mutations, leading to amino acid substitutions of the VP1 protein described in a recent study (Gorelik et al., 2011).

Regardless of the exact pathogenetic mechanism, our data suggest that the detection of GCN-type JCV will potentially help clinicians diagnose JCV GCN, using the PCR primers encompassing the area of interest. Interestingly, the patient...
with JCV_{GCN1} also had an undeleted wild-type JCV strain in his CSF. The concomitant presence of deleted and undeleted JCV strains was also present in our initial case (Dang & Koralnik, 2006; Du Pasquier et al., 2003), as well as in 25% of CSF and 68% of PBMC samples from PML patients, suggesting that the deletion may arise outside of the CNS, perhaps at the level of the bone marrow (Tan et al., 2009).

Studies of JCV pathogenesis in neurons are challenging since JCV only infects humans, and no suitable human neuronal cell line is available. We therefore used for the first time a cell line derived from a human medulloblastoma, a neuronal tumour arising from the cerebellar granule cell layer. The viral load of all JCV strains remained low in this cell line. However, the fact that JCV could replicate at all in cells of neuronal origin which, unlike SVG or COS-7, were not previously transformed and immortalized with the SV40 T-Ag gene, is remarkable. We had initially postulated that the isolates with GCN1 deletion would have a selective advantage in this cell line, which was not verified experimentally. These results highlight the difficulties associated with modelling neuronal infection by JCV in vitro. However, our data indicate that the GCN1 mutation does not prevent expression of the VP1 protein, and that the deletion remains stable over a long period of time in vitro, without reverting to wild-type. Furthermore, detection of the VP1 protein by Western blot in DAOY cell lysate indicates that despite a low level of viral replication, a full replicative cycle occurred in these cells with production of mature viral particles.

As expected, the prototype JCV_{Mad1} outperformed other JCV strains in astroglial cells, whereas the introduction of the GCN1 deletion in this strain conferred the same phenotype as JCV_{GCN1}. These results are consistent with the fact that an undeleted VP1 protein provides an advantage for replication in glial cells. It is not clear why JCV_{HWM} isolated from a PML lesion, did not replicate significantly better than JCV_{GCN1} in this cell line. Both viral isolates have a similar RR with tandem repeat pattern (Dang & Koralnik, 2006) and share the same backbone and differ only in the RR, which may dampen its replication in SVG cells. Finally, the fact that an undeleted VP1 protein provides an advantage for replication in glial cells. It is not clear why JCV could replicate at all in cells of neuronal origin which, unlike SVG or COS-7, were not previously transformed and immortalized with the SV40 T-Ag gene, is remarkable. We had initially postulated that the isolates with GCN1 deletion would have a selective advantage in this cell line, which was not verified experimentally. These results highlight the difficulties associated with modelling neuronal infection by JCV in vitro. However, our data indicate that the GCN1 mutation does not prevent expression of the VP1 protein, and that the deletion remains stable over a long period of time in vitro, without reverting to wild-type. Furthermore, detection of the VP1 protein by Western blot in DAOY cell lysate indicates that despite a low level of viral replication, a full replicative cycle occurred in these cells with production of mature viral particles.

**Construction of JCV_{GCN1}.** Due to the low viral copy number, the full-length sequence of JCV_{GCN1} in the cerebellum sample of our index patient had been previously characterized after sequencing multiple PCR-amplified fragments covering the entire JCV genome (GenBank accession no. DQ875212) (Dang & Koralnik, 2006; Du Pasquier et al., 2003). Considering that JCV_{GCN1} and the JCV strain isolated from the hemispheric white matter of this patient, called JCV_{HWM} obtained by long PCR amplification (GenBank accession no. DQ875211), shared the same backbone and differ only in the RR, the C terminus of VP1-coding region, and the C terminus of the T-coding region (Dang & Koralnik, 2006), we used JCV_{HWM} genome to reconstruct a full-length clone of JCV_{GCN1}. The DNA template corresponding to JCV_{GCN1} RR located between two Nco sites (nt 5014–5169) was amplified using primer pair: JGCNco5 (5′-GAAATTTCCATGTTTCTC-3′, GGAATT-4958–4970 of JCV_{HWM}) and JGCNco3 (5′-CTTTGGC- TTGGTTTACTT-3′, 257–240 reverse complement sequence of JCV_{HWM}, designed by ‘Primer Express/version 2.0’; Applied Biosystems). The PCR product was cloned into TA vector (Invitrogen) and sequenced. One positive clone was selected to proceed to the next step. Plasmid containing the JCV_{Nos} fragment was purified from selected positive clone and was digested by Nos (New England Biolabs) to free the JCV_{Nos} fragment. The JCV_{Nos} fragment was recovered from agarose gel (QIAquick Gel Extraction kit; Qiagen) and used to replace the corresponding sequence of JCV_{HWM}. The JCV strain obtained after this step was a new JCV strain that had the backbone of JCV_{HWM} and RR of JCV_{GCN1}, this strain was called JCV_{GCN1-RR}.

DNA template of partial JCV_{GCN1} genome containing the deleted VP1 gene and intact C terminus of T gene with two single restriction enzyme sites of Pad and Bsu361 (nt 2369–3056, JCV_{HWM} sequence) was also synthesized by BlueHeron (nt 2097–3107, JCV_{HWM} sequence, 1099 bp) and named PG_{VP1-T}. Primer pair: JV2For (5′-AAGATATTTTGGGACACATCAACAGGAGGAGA-3′, 2097–2126 of JCV_{HWM}) and JV2Rev (5′-TTCAGGGCATGGCATAAGCAACC-3′, 3187–3165 RC sequence of JCV_{HWM}, designed by ‘Primer Express/ version 2.0’) were used to amplify partial JCV_{GCN1} sequence containing the C terminus of both VP1 and T gene from PG_{VP1-T}. The PCR product was then cloned into TA vector, screened and sequenced. The plasmid containing the PG_{VP1-T} fragment was purified and digested by Pad (2369 of JCV_{HWM}) (Bilcock et al., 1999; Haciglugu et al., 1997) and Bsu361 (3056 of JCV_{HWM}) (New England Biolabs). The 1086 bp PG_{VP1-T} fragment containing 10 bp VP1 deletion and intact C terminus of the T gene was recovered from agarose gel (QIAquick Gel Extraction kit; Qiagen) and then used to replace the counterpart of JCV_{GCN1-RR} to obtain the full-length genome of JCV_{GCN1}.

**Construction of JCV_{Mad1D}.** A similar strategy was used to obtain JCV_{Mad1D}, a mutant of the JCV reference strain JCV_{Mad1} containing the 10 bp deletion in the VP1 gene found in JCV_{GCN1}. DNA template containing the VP1 gene of JCV_{Mad1D} with 10 bp deletion in the C terminus was synthesized by BlueHeron (nt 2336–3806 of JCV_{Mad1} strain with 10 bp deletion between 2503 and 2512). Primer pair: Mad1DS (5′-AACCCCTACCCAATTTTCCTTCCC-3′, 2336–2360 of JCV_{Mad1} strain) and Mad1DS (5′-GCTTATGGGCTATGAC-3′, 2398–2797 RC sequence of JCV_{Mad1} designed by ‘Primer Express/ version 2.0’) were used to amplify the synthesized template. The PCR product was cloned into TA vector and sequenced. Target JCV_{Mad1D} DNA sequence was freed from the vector using Pad (position 2373 of JCV_{Mad1}) and Pstl (position 3359 of JCV_{Mad1}) (New England Biolabs). The 977 bp target band (with 10 bp VP1 deletion) was recovered from agarose gel (QIAquick Gel Extraction kit; Qiagen) and then used to replace the counterpart of JCV_{Mad1D} to obtain full-length genome of JCV_{Mad1D}.

**METHODS**

This study was approved by the BIDMC institutional review board. Patients consent was obtained according to institutional IRB guidelines.
Plasmids containing full-length genome of different JCV strains were used to transform *Escherichia coli* INV110 for large-scale preparation, leading to methylation of *DpnII* sites.

**In vitro transfection of four JCV strains into three different cell lines.** In the *in vitro* transfection experiment, a total of four JCV strains; JCVMad1, JCVMad1D, JCVHVIM, JCVSCN, were used to transfect separately three different cell lines: human astroglial SVG (purchased from ATCC), simian kidney cell COS-7 (purchased from ATCC) and human medulloblastoma DAOY (a generous gift from Dr Scott L. Pomeroy, Children’s Hospital, Boston, MA, USA). Plasmids containing the full-length JCV genome were digested with EcoRI (New England Biolabs), the 5 kb band was recovered from the agarose gel using QIAquick Gel Extraction kit (Qiagen). Transfection was performed with a viral per cell ratio of 10² viral genome copies per cell using Effectene transfection reagents from Qiagen in a six-well culture plate. Cells transfected with the JCV genome were then transferred from a six-well plate to P25 flask 3 days post-transfection, and to P175 flask 6 days post-transfection. JCV-transfected cells were passaged into a new P175 flask at a ratio of 1:5 every 3.5 days, and DNA was extracted from one-fifth of the total cell number at each passage (QIAamp DNA Blood Mini kit; Qiagen). Supernatant was collected at each passage and stored at −20 °C. DNA was extracted from the supernatant using QIAamp DNA Blood Mini kit after cell culture was terminated. The experiment ended after 100 days and 30 passages *in vitro*.

**QPCR detection of JCV viral load in the *in vitro* transfection experiment.** DNA samples extracted from both JCV-transfected cells and the corresponding supernatant were quantified by SmartSpec Plus spectrophotometer (Bio-Rad). To remove any remaining contaminating input JCV DNA, samples were digested with *DpnII* (New England Biolabs) in a volume of 30 µl at 37 °C overnight. Since there are two *DpnII* sites (GA<sup>III</sup>TC, nt 841–844 and 970–973) located within the QPCR target sequence (nt 792–992), any methylated DNA sequence from input viral genome would be digested and could not be used as PCR template. After heat inactivation of *DpnII* at 80 °C for 20 min, 0.5 µg digested DNA was used as template in QPCR analysis. Primer pair: JCD792 (5′-TTGTGTTGTTGTGCTAGTGG-3′, position 792–815 of JCVMad1 amplifying a conserved sequence in all four JCV strains, and JCD992 (5′-AGCGGGCCCAATTGTCTAG-3′, reverse complement sequence of position 974–992) were used for QPCR detection. Probe JCV0824 (6FAM-AGATTTTTGTGGTGAAT-3′), which cross-reacts with JCV VP1 protein, was detected using mouse mAb anti-SV40 VP1 protein (PAB597; generous gift from Dr Walter Atwood), which cross-reacts with JCV VP1 protein. The presence of the protein was revealed with HRP-conjugated goat anti-mouse Ab (GAM-HRP) (Bio-Rad).

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**REFERENCES**


