**Bombyx mori** nucleopolyhedrovirus ORF9 is a gene involved in the budded virus production and infectivity

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The ORF9 of **Bombyx mori** nucleopolyhedrovirus (BmNPV) (Bm9) is conserved in all completely sequenced lepidopteran nucleopolyhedroviruses. RT-PCR analysis demonstrated that Bm9 is an early and late transcribed gene that is initiated at 3 h post-infection, and immunofluorescence microscopy showed that Bm9 is localized mainly in the cytoplasm of infected cells. To determine the role of Bm9 during virus infection, Bm9 was knocked out by recombination in a BmNPV genome propagated as a bacmid in *Escherichia coli*. The budded virus (BV) production of Bm9-deleted bacmids was reduced more than 10-fold compared with wild-type (wt) bacmid; however, the kinetics of viral DNA replication were unaffected. The defect in BV production was recovered by the Bm9 rescue bacmid. In addition, electron microscope observations revealed that polyhedra formation was not affected by the deletion of Bm9. Bioassays showed that the Bm9-deleted bacmid took approximately 14–22 h longer to kill fifth instar *B. mori* larvae than wt bacmid, and the LD50 was about 15 times higher than that of the wt bacmid. In conclusion, Bm9 is an important but not essential factor in virus production and infectivity in vivo and in vitro.

**INTRODUCTION**

Baculoviruses are double-stranded DNA viruses that are pathogenic for invertebrates, particularly insects of the order Lepidoptera. The family Baculoviridae is taxonomically divided into two genera, Nucleopolyhedrovirus and Granulovirus (Theilmann et al., 2005). Baculovirus gene expression is regulated mainly at the transcriptional level, and involves sequential expression of early, late and very late genes. The function of early genes is the initiation of infection, and they are required to accelerate replication events and to prepare the host cell for virus multiplication (Friesen, 1997). The late stage occurs following the initiation of viral DNA replication. Two virion phenotypes are produced: occlusion derived virus (ODV) and budded virus (BV) (Funk et al., 1997). The ODV transmits infection from insect to insect by infecting midgut columnar epithelial cells, whereas the BV is responsible for causing systemic infection within the host (Keddie et al., 1989).

Baculoviruses are being extensively studied for their potential use as bioinsecticides around the world and as expression vectors for heterologous gene expression in insect-derived cells, as well as in host caterpillars. Recently, **Bombyx mori** nucleopolyhedrovirus (BmNPV) has been successfully developed for surface display of recombinant proteins (Rahman & Gopinathan, 2003), and baculoviruses are also regarded as potentially useful gene therapy vectors (Condrey & Kost, 2007; Huser & Hofmann, 2003; Tani et al., 2003). These applications drive the study of the molecular basis of baculovirus infection.

Since the complete genome of BmNPV was determined (Gomi et al., 1999), extensive studies have focused on the function of individual genes. The open reading frame 9 (ORF9) of BmNPV (Bm9), homologous to ORF17 of *Autographa californica* multiple nucleopolyhedrovirus (AcMNPV), is considered to be a gene specific for lepidopteran nucleopolyhedroviruses (NPVs), suggesting that the gene plays an important role in the infection cycle of lepidopteran baculovirus. A previous study showed that AcMNPV ORF17 was dispensable for trans-activation in expression of a late gene (Guarino & Summers, 1988) and a recent study indicated that it was an early gene encoding a protein located in the cytoplasm of infected cells (An et al., 2006). However, its function is still unknown.

**METHODS**

Cell line, virus, bacterial strains, bacmids and plasmids. BmNPV (ZJ strain) and BmNPV bacmid virus were propagated in...
BmN (BmN-4) cells, maintained at 27 °C in TC-100 insect medium supplemented with 10% (v/v) fetal bovine serum. Routine manipulations were performed according to standard protocols (O’Reilly et al., 1992). The Escherichia coli strain BmDH10B, containing BmNPV bacmid (BmBac) DNA (Motohashi et al., 2005), was kindly provided by Dr Enoch Y. Park (Shizuoka University, Japan). The E. coli strain BW25113 (pKD46) was kindly provided by Dr Mary Berlyn (Yale University, USA). The plasmid pKD46 contains the phage λ Red system under the control of the arabinose promoter. The E. coli strain DH10Bac (Invitrogen) was used to isolate the helper plasmid (pMON7124), which encodes a transposase.

The pRADZ3 plasmid, containing the chloramphenicol resistance gene (CmR), was kindly provided by Dr Yuejin Hua (Zhejiang University, China). The pFastBac1-PG plasmid (Wu et al., 2006), harbouring the green fluorescence protein gene (gfp) under the control of AcMNPV ie1 promoter and polyhedrin gene (polh) under the control of the polh native promoter, was kindly provided by Dr Yi Pang (Sun Yat-sen University, China).

**RNA isolation and RT-PCR.** Monolayers of BmN cells (1 × 10^6) were infected with BmNPV BV at an m.o.i. of 5. Total RNA from mock- or BmNPV-infected cells was extracted at 3, 6, 12, 24, 48 and 72 h post-infection (p.i.). After treating with RNase-free DNase I (TaKaRa), cDNA was synthesized by using the PrimeScript reverse transcriptase (TaKaRa) with oligo-dT(18) according to the manufacturer’s protocol. The coding region of the Bm9 gene was amplified by PCR with Bm9-specific primers Bmorf9F and Bmorf9R (Supplementary Table S1). This PCR product was digested with PstI and ligated with pFB1-PH-GFP to generate the donor plasmid, pFB1-De9Re-PH-GFP. Briefly, a 1562 bp fragment containing the Bm9 gene and its native promoter and poly(A) signal was PCR amplified with primers DeBm9F and DeBm9R, and subsequently used for marker gene insertion and polh-negative. To examine the effects on occlusion morphogenesis derived from the Bm9 deletion, we constructed a donor plasmid, pFastBac-Bmp6 (Xu et al., 2008), to facilitate introduction of the BmNPV polh gene cassette into the BmNPV bacmid and the Bm9-deleted BmBac. pFastBac-Bmp6 was transformed into BmDH10B and DH10B/pBmBacA9 helper competent cells to construct wt BmNPV bacmid and a knockout bacmid containing the polh gene, respectively, designated BmBacWT-PG and BmBacKO-PG, respectively. The constructed plasmids were verified by PCR with pUC/M13 forward and reverse primers (Supplementary Table S1).

**Construction of BmNPV, knockout and repair bacmids containing gfp and polh.** To facilitate examination of virus infection, we introduced a donor plasmid pFB1-PH-GFP, which was generated by inserting AcMNPV polh and gfp (under the control of AcMNPV ie1 promoter) genes into pFastBac1 (Invitrogen) (Wu et al., 2006). pFB1-PH-GFP was transformed into BmDH10B and DH10B/pBmBacA9/helper competent cells to construct wt BmNPV bacmid and a knockout bacmid containing the polh gene, respectively, designated BmBacWT-PG and BmBacKO-PG, respectively. The constructed plasmids were verified by PCR with pUC/M13 forward and reverse primers.

To construct Bm9-rescued BmBac, we generated another donor plasmid, pFB1-De9Re-PH-GFP. Briefly, a 1562 bp fragment containing the Bm9 gene with its native promoter and poly(A) signal was PCR amplified with primers DeBm9F and DeBm9R, and subsequently used for marker gene insertion and polh-negative. To examine the effects on occlusion morphogenesis derived from the Bm9 deletion, we constructed a donor plasmid, pFastBac-Bmp6 (Xu et al., 2008), to facilitate introduction of the BmNPV polh gene cassette into the BmNPV bacmid and the Bm9-deleted BmBac. pFastBac-Bmp6 was transformed into BmDH10B and DH10B/pBmBacA9 helper competent cells to generate BmBacWT-PG and BmBacKO-PG, respectively. The constructed plasmids were verified by PCR with pUC/M13 forward and reverse primers.

**Viral growth curves.** To investigate viral growth, monolayers of BmN cells (1 × 10^6) cultured in 1 ml serum-free TC-100 medium were infected with BmBacKO-PG, BmBacKO-PG and BmBacWT-PG bacmids in 35 mm dishes with an m.o.i. of 5. After 1 h incubation, the inoculum was removed and the cells were washed with TC-100 medium three times, and exchanged with TC-100 medium supplemented with 10% serum; this was defined as time zero. Each infection was done in triplicate. The supernatant was collected at the indicated times p.i. and the viral titre was determined by the end point dilution method (TCID50) in triplicate (O’Reilly et al., 1992).

**Quantitative PCR (qPCR) analysis of viral DNA replication.** To assess viral DNA replication, we quantified viral genomic DNA accumulation by using qPCR. The infection was performed as described above. At designated times p.i., cells were collected and bacmid DNA was extracted by using the method described by King & Possee (1992). Primers QR and QF (Supplementary Table S1) were designed for qPCR, and the reaction was carried out using the SYBR Premix Ex Taq kit (TaKaRa) on the iCycler machine (BioRad) using
the following conditions: 95 °C for 10 s and 40 cycles of 95 °C for 5 s
and 55 °C for 20 s.

We generated a recombinant plasmid (pGEX4t-2-118) with an
insertion in BmNPV ORF118 and used a dilution series of this
plasmid to generate a standard curve for qPCR. The number of
genomic copies of each sample were determined using this standard
curve.

**Electron microscopy (EM).** Monolayers of BmN cells were infected
with BmBacWT and BmBacKO at an m.o.i of 5. At 96 h p.i., the cells
were harvested by centrifugation (5000 g, 5 min, 4 °C) and the pellet
was fixed in 2.5% glutaraldehyde for 1 h at 4 °C, followed by fixing
with 1% osmium tetroxide for 1 h at room temperature. Next, the
pellets were dehydrated in graded ethanol (50–100%), and then
soaked in acetone for 20 min. Infiltration was performed in graded
Spurr (50–100%) (Sigma) and exposed for 16 h at 70 °C. Ultrathin
sections were cut with a diamond knife. After staining with uranyl
acetate and lead citrate, the ultrathin sections were viewed under a
JEM-1230 transmission electron microscope (JEOL) at an accelerating
voltage of 80 kV.

**B. mori larvae bioassay.** The LD₅₀ and median lethal time (LT₅₀) of
BV were estimated by haemocoelically injecting different doses of BVs
diluted in PBS (50, 500, 5000 and 50 000 p.f.u.) into B. mori larvae,
BaiYu × Qiufeng strain, within 8 h of moulting to the fifth instar.
Twenty-five larvae per dose were used and each dose was repeated in
triple. The mortality was counted every 4 h.

The computation processes, including modelling, test for the
goodness of fit and estimation of LD₅₀ and LT₅₀, were conducted
using DPS Data Processing System software (Tang & Feng, 2002),
adopting Probit analysis (Finney, 1971).

To investigate the effects on LD₅₀ and LT₅₀ comparisons between the
viruses were performed using one-way ANOVA followed by LSD test
to identify significant differences (P<0.01).

**RESULTS**

**Sequence and transcriptional analysis of Bm9**

The ORF9 of BmNPV (Bm9) is located between nt 8725 and
9355 in the genome, encoding a putative protein of 210 aa with a predicted molecular mass of 24.1 kDa; the
gene is transcribed in the same orientation as the polh gene
(Gomi et al., 1999). Sequence analysis revealed that a
TATA box, a baculovirus consensus early promoter motif
CAGT and two late transcriptional motifs, GTAAG and
ATAAG, were located at 56, 436, 220 and 444 nt upstream
of the start codon ATG, respectively. All of the early and
late motifs are far from the start codon. The prediction of
domains, motifs and post-translational modifications in
the amino sequence were carried out by Motifscan online
(http://www.expasy.org). An N-myristoylation site, a
putative viral immunoglobulin (Ig)-like domain 1
(Big-1) domain profile, an asparagine-rich region profile,
two putative casein kinase II phosphorylation sites, two
putative protein kinase C phosphorylation sites and three
putative N-glycosylation sites were predicted in the
encoded amino acid sequence.

To determine the transcriptional profile of Bm9, the BmN
cells were infected with BmNPV at an m.o.i of 5, and then
collected at various times p.i. for RT-PCR analysis.
Transcriptional analysis by RT-PCR revealed that a
product with predicted size of 592 bp was detectable at
3 h p.i., and was still stable at the very late phase (72 h p.i.)
(Supplementary Fig. S1, available in JGV Online). This
observation indicated that Bm9 was transcribed through-
out all stages of infection. In contrast, no PCR product was
detected in the mock-infected sample (Supplementary Fig.
S1). Also, no amplicon was obtained when the RT-PCR
step was omitted, which ruled out the possibility of
contamination of the cDNA sample with BmNPV genomic
dNA (data not shown).

**Immunofluorescence microscopy**

To investigate the subcellular localization of the Bm9
protein, we carried out an indirect immunofluorescence
assay to examine its distribution. Cells were collected for
immunofluorescence analysis at different times p.i.
Fluorescence signals were detected throughout infection
within the cytoplasm but not in the nucleus, as shown at
72 h p.i. (Fig. 1), indicating that Bm9 localizes mainly
within the cytoplasm of infected cells. In the control
samples, no fluorescence was detected with anti-Ac17
serum (Fig. 1). Preimmune sera did not react with
uninfected or infected samples (data not shown). The
cross-reactivity of anti-Ac17 antibody to Bm9 was shown
by Western blot analysis (Supplementary Fig. S2, available
in JGV Online).

**Construction of Bm9 knockout, repair and wt
BmNPV bacmid**

To determine the function of Bm9 in viral replication, we
generated a Bm9-deleted BmNPV bacmid via the λ Red
homologous recombination system in E. coli. A short
sequence of 23 bp (nt 8985–9007) within the Bm9 locus
was replaced with the Cmr6 gene cassette (Fig. 2a). A portion
(260 bp) of the 5’ end and 350 bp of the 3’ end of the Bm9
ORF were retained in order to preserve transcription of the
adjacent genes, ORF8 and ORF10. The precise deletion was
confirmed by PCR; as expected, a PCR product of 1.5 kb
(which includes the inserted Cmr6 cassette) was amplified
from the Bm9-deleted bacmid, while a 0.59 kb product was
amplified from the original bacmid (Fig. 2d), demonstrating
that Bm9 was successfully replaced with the Cmr6 cassette by
homologous recombination.

Since the BmNPV bacmid was phenotypically polh-
negative, we constructed the donor plasmid pFastBac-
Bmhp to transpose the polh gene cassette into the original
bacmid and the Bm9-deleted bacmid to generate BmBacWT
(Fig. 2b) and BmBacKO (Fig. 2c), respectively. The BmNPV
polh transposition was verified by PCR with pUC/M13
forward and reverse primers, which resulted in a 3.7 kb
product as expected (Fig. 2d). The BmBacWT and BmBacKO
bacmids were used to transfect BmN cells for further ODV
observation.
To facilitate observation of viral infection, the donor plasmid pFB1-PH-GFP, containing AcMNPV polh and gfp, was transposed into the polh locus in the original bacmid and Bm9-deleted bacmid to produce BmBac\textsuperscript{WT-PG} (Fig. 2b) and BmBac\textsuperscript{KO-PG} (Fig. 2c), respectively. The PCR with pUC/M13 forward and reverse primers confirmed successful transposition, which yielded the expected 4.2 kb products (Fig. 2d). The BmBac\textsuperscript{WT-PG} and BmBac\textsuperscript{KO-PG} bacmids were used to transfect BmN cells for viral DNA replication assay and BV titre assay.

To confirm the phenotype resulting from the Bm9 deletion, we constructed a Bm9 rescue bacmid. The donor plasmid pFB1-De9Re-PH-GFP, containing polh, egfp and Bm9 (with its native promoter), was transposed into the polh locus in the Bm9-deleted bacmid to create a rescue bacmid, named BmBac\textsuperscript{RePG} (Fig. 2c). The successful transposition was verified by PCR with primer pUC/M13 forward and reverse primers, and a 5.7 kb product was amplified as expected (Fig. 2d).

**Observation of BmBac\textsuperscript{KO-PG}, BmBac\textsuperscript{WT-PG} and BmBac\textsuperscript{RePG} transfection in BmN cells**

To examine the effect of Bm9 deletion upon virus replication, the BmN cells were transfected with Bm9-deleted (BmBac\textsuperscript{KO-PG}), Bm9 rescue (BmBac\textsuperscript{RePG}) and wt (BmBac\textsuperscript{WT-PG}) bacmids. At 72 h post-transfection, these cells all presented cytopathic effects (CPE) and green fluorescent protein (GFP) expression (Supplementary Fig. S3, available in JGV Online). To further confirm the infectious capability of the BmBac\textsuperscript{WT-PG}, BmBac\textsuperscript{KO-PG} and BmBac\textsuperscript{RePG}, a passage assay was performed. The supernatants from these cells were added to fresh BmN cells and incubated for 72 h. A large amount of GFP was observed in the majority of cells infected with the three bacmids, indicating that infectious BV spread among cells. The knockout virus remains infectious after passing through several generations. Thus, the viral passage experiments suggested that the Bm9 deletion does not abort BV production in cultured BmN cells.

**Analysis of viral growth curves**

To quantitatively assess the effect of the Bm9 deletion on BV production, a viral growth curve was generated for supernatants from BmN cells infected with BmBac\textsuperscript{WT-PG}, BmBac\textsuperscript{KO-PG} and BmBac\textsuperscript{RePG}. At designated times p.i., the supernatants were collected in order to determine BV titre by TCID\textsubscript{50} assay. By 16 h p.i., the BV titre of BmBac\textsuperscript{KO-PG} was approximately one log (10-fold) lower than that of BmBac\textsuperscript{WT-PG}. However, the titre of BmBac\textsuperscript{RePG} could reach the BmBac\textsuperscript{WT-PG} level at 96 h p.i. (Fig. 3a), though there was a delay in BV production from 16–72 h p.i. Therefore, the data showed that the Bm9 deletion significantly reduced BV production in cultured BmN cells, and such a defect could be rescued by inserting the Bm9 gene cassette back into the knockout bacmid.

**Viral DNA replication**

To examine whether the reduced BV was due to viral DNA replication, we used qPCR to monitor viral genomic DNA accumulation in infected cells. The results revealed that, throughout the infection process, the Bm9-deleted bacmid presented similar kinetic growth curves of viral DNA replication to the wt and rescued bacmids. From 0 to 3 h, a phase prior to DNA replication, low numbers of viral
Fig. 2. Construction of BmBacKO, BmBacWT, BmBacKO-PG, BmBacWT-PG and BmBacRePG, and PCR identification of the constructs. (a) The scheme for construction of a Bm9 knockout bacmid. A 23 bp portion of the Bm9 locus was replaced with CmR via homologous recombination. (b) Schematic diagram of the wt bacmids BmBacWT-PG and BmBacWT. BmBacWT-PG was constructed by inserting AcMNPV polh and gfp into the BmNPV bacmid’s polh locus by Tn7-mediated transposition in the Bac-to-Bac system. BmBacWT was generated by inserting only BmNPV polh into the BmNPV bacmid’s polh locus. (c) Schematic diagram of the constructed bacmids BmBacRePG, BmBacKO-PG and BmBacKO. The rescue bacmid, BmBacRePG, was constructed by inserting AcMNPV polh, Bm9 gene cassette and gfp into the Bm9-deleted bacmid’s polh locus by Tn7-mediated transposition in the Bac-to-Bac system. The AcMNPV polh and gfp were inserted into the Bm9-deleted bacmid’s polh locus to generate BmBacKO-PG. The BmNPV polh was inserted into the Bm9-deleted bacmid’s polh locus to generate BmBacKO. (d) PCR identification of the Bm9 deletion and reconstructed bacmids of BmBacKO, BmBacWT, BmBacWT-PG, BmBacKO-PG and BmBacRePG. The sizes (bp) of the PCR products are indicated.

Fig. 3. Time-course analysis of BV production (a) and viral DNA replication (b) from Bm9-deleted bacmid (BmBacKO-PG; —) and wt bacmid (BmBacWT-PG; …..) and the rescue bacmid (BmBacRePG; -.-). (a) BmN cells were infected with an m.o.i of 5, and supernatant was collected at the indicated times. The BV titres were determined by TCID50. (b) BmN cells were infected with an m.o.i of 5; cells were collected at the indicated times and viral DNA was extracted for qPCR analysis. Results are shown as mean ± SD.
genomes per cell (<10) were detected (Fig. 3b). By 12 h p.i., approximately 100–500 genomes per cell were detected in cells transfected by all three bacmid-generated viruses (Fig. 3b). By 36 h p.i., the accumulation of viral DNA reached a plateau, with approximately 3000 genomes per cell (Fig. 3b). Thus, the assay illustrated that the Bm9 deletion does not affect viral DNA replication in cultured BmN cells.

**Examination of polyhedra formation**

EM observation was performed to examine the distribution of nucleocapsids of BmBacKO and BmBacWT in infected cells. The results showed that there was no discernible morphological effect on nucleocapsid distribution for the two bacmids (Fig. 4). The BmBacKO-infected cells displayed features characteristic of baculovirus infection, including an enlarged nucleus, the presence of an electron-dense virogenic stroma, lots of nascent nucleocapsids accumulated within the stromal matter and polyhedra occluded with nucleocapsids (Fig. 4b, c). Thus, no obvious discrimination of polyhedra formation was observed in the Bm9 deletion bacmid.

**Effect of Bm9 deletion on BV infectivity in B. mori larvae**

To determine whether the Bm9 deletion has an effect on infectivity in *B. mori* larvae, fifth instar larvae were haemocoelically injected with BmBacKO-PG, BmBacWT-PG and BmBacRepG BVs. Firstly, LD50 was determined by injecting with various doses of BV (50, 500, 5000 and 50 000 p.f.u.). The data revealed that there were significant differences in LD50 for BmBacKO-PG and BmBacWT-PG, which yielded 1987.5 p.f.u. and 137.5 p.f.u. (Table 1), respectively. No significant difference in LD50 was observed between BmBacWT-PG and BmBacRepG (Table 1). Thus, the LD50 assay indicated that the Bm9 deletion substantially decreased the infectivity of BmNPV budding virus in *B. mori* larvae.

To determine the LT50, fifth instar *B. mori* larvae were haemocoelically injected with various doses of BV (50, 500, 5000 and 50 000 p.f.u.). We observed that the LT50 of BmBacKO-PG was about 14–22 h longer than that of BmBacWT-PG and BmBacRepG at every dose tested (Table 2). Additionally, we examined the BV production in *B. mori* larvae after injection with BmBacWT-PG and BmBacKO-PG (5000 p.f.u. per larva) for 6 days. The BV titre of BmBacWT-PG was approximately 19 times higher than that of BmBacKO-PG, with 5.62 × 107 and 3.03 × 108 TCID50 (ml haemolymph)−1, respectively (Supplementary Fig. S4, available in JGV Online).

**DISCUSSION**

*Bm9* and its homologues are found specifically in lepidopteran NPVs (Herniou *et al.*, 2003), suggesting that in many other baculovirus host systems, the function of this gene either is not required or is replaced by other viral and/or cellular factors.

In this study, we examined transcription of the *Bm9* gene of BmNPV and the cellular localization of the gene product, as well as its effect on viral replication of BmN cultured cells. Transcriptional analysis via RT-PCR demonstrated that *Bm9* is an early and late transcribed gene, with transcripts initially observed at 3 h p.i. (Supplementary Fig. S1); this is identical to observations of its homologue Ac17 (An *et al.*, 2006; Guarino & Summers, 1988). To detect the Bm9 distribution in virus-infected cells, we performed immunofluorescence assays with polyclonal antibody against Ac17. The results showed that Bm9 protein was found exclusively in the cytoplasm at 72 h p.i. (Fig. 1).

To further investigate the function of Bm9, we constructed a *Bm9*-deleted bacmid virus. The deletion of *Bm9* did not affect viral DNA replication (Fig. 3b). However, the *Bm9*-null virus was deficient in the production of infectious virions, with BV titres reduced to 10% of wt levels (Fig. 3a). This defect could be restored by insertion of the *Bm9* gene cassette into the *polh* locus, indicating that such a phenomenon is derived from *Bm9* deletion.

As far as BV production was concerned, *Bm9* had a similar function to *pe38* and *pp31* genes, which both greatly reduced BV yields (Milks *et al.*, 2003; Yamagishi *et al.*,...
However, the pe38-null virus reduced the viral DNA synthesis level, whereas the Bm9- and pp31-null viruses presented identical levels of DNA replication compared to wt bacmid virus. Subsequently, EM analysis indicated that the rod-shaped nucleocapsids, virogenic stroma, ODVs and polyhedral containing ODVs were formed in the BmBacKO virus-infected cells and showed no difference to those in BmBacWT virus-infected cells. Therefore, the results indicated that Bm9 was not essential for occlusion body morphogenesis in BmNPV-infected cells. Guarino & Summers (1988) found that the Ac17 gene, which was included in the PstI-G region, was not required for late gene expression. Their results and ours indicate that Bm9 is dispensable for DNA replication and some late or very late gene expression at the cellular level. Bioassay results showed that the deletion of Bm9 elongated the LT50 and increased the LD50 compared with the controls, which is consistent with the reduction of BV production detected in the in vitro assay, suggesting that both infectivity and virulence were reduced in the deletion bacmid. These results support the hypothesis that defects in BV production led to reduced virulence in infected larvae (Milks et al., 2003). To determine whether Bm9 is a component of BV or ODV structure, we carried out a Western blot of BV and ODV using anti-Ac17 serum (Supplementary Fig. S5, available in JGV Online). Our results showed that Bm9 was a component of neither BV nor ODV, suggesting that the reduction in the infectivity was not due to a lack of Bm9 in the budded virion structure, but a reduction in the number of budded virions produced. It is interesting to note that, based on sequence analysis, Bm9 has a Big-1 domain in the N-terminal region, which is present in bacterial adhesion molecules of the intimin/invasin family, and is involved in pathogenicity. The Big-1 proteins are surface-expressed proteins mediating mammalian host cell invasion or attachment (Hamburger et al., 1999; Kelly et al., 1999; Luo et al., 2000). However, it was not determined whether Bm9 was also expressed on the membrane surface. Whether this domain of Bm9 had an

### Table 1. Dose mortality of BmBacWT-PG, BmBacRePG and BmBacKO-PG for fifth instar B. mori larvae

All analyses used 2 degrees of freedom.

<table>
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<th>Virus</th>
<th>Regression equation</th>
<th>$\chi^2$ value</th>
<th>$P$ value</th>
<th>LD50 (p.f.u.)*</th>
<th>95% confidence limit (p.f.u.)</th>
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<td>$y=0.72 x - 3.66$</td>
<td>2.5401</td>
<td>0.2808</td>
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<td>92 183</td>
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<td>BmBacRePG</td>
<td>$y=0.53 x - 3.79$</td>
<td>0.1836</td>
<td>0.9123</td>
<td>142.2 a</td>
<td>105.4 191.9</td>
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<td>BmBacKO-PG</td>
<td>$y=0.81 x - 2.35$</td>
<td>0.8836</td>
<td>0.6429</td>
<td>1987.5 b</td>
<td>1319.5 3473</td>
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</table>

*LD50 values followed by a different letter are statistically different ($P<0.01$). If $P>0.05$, the model was determined to be statistically significant.

### Table 2. Time mortality of BmBacWT-PG, BmBacRePG and BmBacKO-PG for fifth instar B. mori larvae

NA, LT50 value (hence 95% confidence limit) not available.

<table>
<thead>
<tr>
<th>Virus concentration per larva (p.f.u.)</th>
<th>Virus</th>
<th>LT50 (h)*</th>
<th>95% confidence limit (h)</th>
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<td>Upper</td>
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<td></td>
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<td>142.6 150.4</td>
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*LT50 values followed by a different letter are statistically different ($P<0.01$).
effect on infectivity and virulence would be of significant interest in future studies.

In conclusion, we examined the transcription of Bm9 in infected cells and found that it was an early transcribed gene. Detection of the localization of Bm9 in BmNPV-infected cells showed that it is mainly localized in the cytoplasm. There was a significant reduction in infectious BV production in cells infected with the Bm9 null virus relative to wt virus; however, the kinetics of viral DNA replication were unaffected. The bioassays following inoculation with BV showed that the Bm9-deleted bacmid took approximately 14–22 h longer to kill the fifth instar larvae than wt bacmid, and that the LD50 was about 15 times higher than that of wt bacmid, demonstrating that Bm9 is not an essential gene for viral invasion and replication but affects the efficiency of virus infection to some degree in larvae. EM analysis of Bm9 knockout virus-infected cells did not highlight obvious differences from wt virus-infected cells. All these results suggested that Bm9 is not necessary for BmNPV propagation but affects infectivity and virulence in vivo and in vitro.

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REFERENCES


