The picornavirus replication inhibitors HBB and guanidine in the echovirus-9 system: the significance of viral protein 2C

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HBB [2-(α-hydroxybenzyl)-benzimidazole] and guanidine are potent inhibitors of picornavirus replication. Among other evidence, limited cross-resistance and a synergistic effect of both inhibitors suggest similar but not identical mechanisms of antiviral action. Echovirus-9 variants resistant to each of these drugs were characterized and sequenced. Complete resistance to HBB or guanidine was shown to be due to single but different point mutations in the non-structural protein 2C. Protein 2C was expressed as GST fusion and His-tagged proteins for the wild-type and various mutants. Although three mutations were located in or near conserved NTP binding motifs, NTPase activity was not altered in the presence of HBB or guanidine.

Introduction

It is well established that 2-(α-hydroxybenzyl)-benzimidazole (HBB) and guanidine selectively inhibit the replication of certain enteroviruses (Caligiuri & Tamm, 1973; Eggers, 1982). Neither of these substances affects processes like adsorption, uncoating or viral protein synthesis. However, it has been known for decades that they block viral RNA synthesis (Eggers & Tamm, 1963a; Eggers, 1982).

Many investigations on poliovirus-1 have revealed that guanidine targets the non-structural protein 2C (Baltera & Tershak, 1989; Pincus et al., 1986; Pincus & Wimmer, 1986) and recently, the determinant for dependence on HBB was found to be located within the 2C region of echovirus-9 (E9) (Hadaschik et al., 1999). Therefore, inhibitors like HBB or guanidine may serve as important tools to extend our knowledge of protein 2C and, in consequence, picornavirus replication.

Protein 2C is one of the most conserved polyproteins of picornaviruses (Argos et al., 1984). However, the function of 2C is still not well understood and seems to be complex (Rueckert, 1996; Wimmer et al., 1993). It has been shown that this protein plays an important role in RNA replication, and the presence of 2C in enterovirus replication complexes comprised of membrane particles, viral RNA and other virus-encoded proteins has been demonstrated (Bienz et al., 1990). Functions like membrane binding, protein–protein interactions, NTPase activity and RNA binding have been revealed for poliovirus 2C by different in vitro techniques (Cuconati et al., 1998; Echeverri & Dasgupta, 1995; Mirzayan & Wimmer, 1994; Pfister & Wimmer, 1999; Rodriguez & Carrasco, 1993, 1995). NTPase activity was also observed in E9 (Klein et al., 1999).

In this report, resistance to HBB and guanidine was defined at the genetic level, and the influence of these antiviral substances on known functions of E9 2C was investigated.

Methods

Virus, replication inhibitors and cells. E9 strain Barty (E9B), originally isolated in Cincinnati, Ohio (Eggers & Sabin, 1959), was passaged as described (Zimmermann et al., 1996). 2-(α-Hydroxybenzyl)-benzimidazole (HBB) was synthesized by Hoechst and guanidine hydrochloride was obtained from Sigma. For cell culture experiments and virus propagation, green monkey kidney (GMK) monolayer cultures were used (Eggers, 1977). Virus titres were determined by plaque assay.

Sensitivity assay. GMK monolayers were infected with the respective virus and the virus-induced cell damage (CPE) was estimated by microscopic examination (Eggers & Tamm, 1961). The average of at least two independent values, differing by not more than 50%, was taken. Infecting virus doses were standardized to approximately 100 TCID_{50} values (median 50% tissue culture infective doses).

Northern blot analysis. Total RNA of single-cycle infected GMK cells was isolated using the guanidium thiocyanate protocol (Ausubel et
Fig. 1. Synergistic inhibitory effect of HBB and guanidine. Wild-type E9 (100 TCID₅₀ per culture) was propagated in the presence of increasing amounts of HBB (a) or guanidine (b) in order to determine concentrations exhibiting comparable inhibitory potential. Thereafter, the action of both HBB and guanidine was studied using low inhibitor concentrations in combination (c).

**NTPase assay.** Echovirus 2C wild-type and mutants were purified as GST fusion proteins and the ATPase reaction was carried out as previously described (Klein et al., 1999). In brief, approximately 1 µg of protein was incubated for 60 min at 37 °C in reaction buffer (20 mM HEPES–KOH (pH 7.5), 5 mM MgCl₂, 2.5 mM DTT, 0.1% BSA, 0.05% Triton X-100, 50 µM ATP and 10 µCi/ml [γ-³²P]ATP (3000 Ci/mmol; Amersham)). For detection, one tenth of the reaction volume was subjected to TLC followed by autoradiography.

**Non-denaturing purification of His-tagged 2C.** The coding region of echovirus 2C was amplified by PCR using the primers ATG GCT GAG CTC GAG AGT AAT GGC TGG CTC AAG (positions 4081–4113) and GTA GAC CTC GAG TTA TTG GAA CAG AGC TTC AGT AAG (positions 5096–5061), which contain manually inserted XhoI restriction sites (in bold). After digestion with XhoI, the PCR product was cloned into a XhoI-linearized pET-14b (Novagen) expression vector. Proper orientation and ligation were controlled by sequence analysis, and the plasmid was transformed into E. coli BL21 (DE3). Expression of His–2C in LB medium was started after induction with 1 mM IPTG at an A₆₀₀ of 1. After an incubation time of 2 h at 25 °C, the bacteria were harvested and resuspended in lysis buffer [0.5 M NaCl, 20 mM Tris–HCl (pH 7–9), 10 mM imidazole, 2 mM β-mercaptoethanol, 0.1% PMSF, 10% glycerol, 1 µg/ml pepstatin A, 1 µg/ml leupeptin, and 0.2% each of Tween 20, Nonidet P-40 and Triton X-100]. Cells were lysed by three freeze–thaw cycles followed by mild sonication. Further purification steps were performed by Ni²⁺ affinity chromatography according to the manufacturer’s recommendations (Qiagen). After step-wise washing and elution, 2C-containing fractions were analysed by SDS–PAGE, concentrated with spin columns (Amicon), and dialysed against 20 mM HEPES–KOH (pH 7.5), 100 mM KCl, 5 mM DTT and 25% glycerol.

**Results**

**Synergism of HBB and guanidine**

Sensitivity assays using different concentrations of HBB or guanidine separately as well as in combination were performed. Concentrations of 30 µM HBB or 0.5 mM guanidine did not inhibit multiplication of E9B significantly (Fig. 1a, b). When half of the respective concentrations of each drug were used together, virus multiplication was delayed for more than 2 days (Fig. 1c). This strongly suggests synergism based on
Cross-resistance was investigated and it could be shown that the multiplication of E9-H.res is possible even at high inhibitor concentrations, indicating that the detected mutations in the 2C gene are relevant for drug resistance.

In order to characterize the significance of each mutation, single nucleotide exchanges were introduced separately into rE9B-fl.20 by site-directed mutagenesis. Sensitivity assays of the resulting viruses revealed that the recombinants bearing mutation A_{1775}C (rE9-H.res and rE9-A\textsubscript{1775}C) exhibited the same phenotype as E9-H.res (Fig. 4b). Hence, this nucleotide exchange appears to be the determinant for HBB resistance. However, the second mutation found in the HBB-resistant isolate, C\textsubscript{1793}U, turned out to introduce dependence on HBB or alternatively on guanidine (Fig. 4c). Analogous experiments with the mutations of the guanidine-resistant isolates revealed that mutation G\textsubscript{4492}A leads to a complete guanidine-resistant phenotype and mutation A\textsubscript{4286}G causes drug resistance with an obvious tendency to guanidine-dependence (Fig. 4b, c).

Like the originally isolated HBB-resistant variant, the recombinant mutants derived from E9-H.res were able to multiply well in the presence of guanidine (Fig. 4). On the other hand, the mutants derived from isolate E9-G.res were only partially resistant to HBB.

**Effect of HBB on viral RNA replication**

To investigate whether the mutations in protein 2C directly influence RNA replication, both positive- and negative-strand RNA synthesis by the virus variants were analysed using single-cycle infection experiments followed by Northern blotting. When using the HBB-sensitive E9 wild-type, no positive-strand RNA was detectable in the presence of HBB

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**Influence of HBB and guanidine on echovirus protein 2C**

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**Isolation and characterization of HBB- and guanidine-resistant E9 variants**

E9B MP5 (Zimmermann et al., 1996) was cultivated under selective pressure of increasing amounts of HBB and guanidine, respectively. Selection was started either with 50 µM HBB followed by two passages each with 150 µM and 220 µM HBB, respectively, or with 0.5 mM guanidine followed by passages with 1.5 mM and 2 mM guanidine. To get rid of dependent viruses also selected under the chosen conditions, one passage without an antiviral agent was performed. Finally, resistant viruses were plaque-purified in the presence of one passage without an antiviral agent was performed. Finally, dependent viruses also selected under the chosen conditions, passages with HBB turned out to be weak (Fig. 2a).

Cross-resistance was investigated and it could be shown that the multiplication of E9-H.res is possible even at high guanidine levels but, in contrast, the resistance of E9-G.res to HBB turned out to be weak (Fig. 2b).

**Cloning and sequencing of E9-H.res and E9-G.res**

Viral RNA of E9-H.res was prepared and transcribed and the longest fragment [position 3893 to the 3’ poly(A) tail] was cloned into the infectious full-length wild-type clone rE9B-fl.20 (Zimmermann et al., 1996), giving rise to the recombinant clone rE9-H.res (Fig. 3b). Sequence comparison of the HBB-resistant cDNA clones and the wild-type clone revealed amino acid exchanges: A\textsubscript{1775}C and C\textsubscript{1793}U. Both mutations are located in the coding region of 2C (Table 1).

Since the mutations responsible for the guanidine resistance of E9B are also supposed to be located within the 2C gene, viral RNA extending from position 3226 to 5474 was amplified by RT–PCR. The resulting DNA was cloned into the rE9B-fl.20 plasmid giving rise to the construct rE9-G.res (Fig. 3c). Sequence analysis also revealed two mutations: A\textsubscript{4286}G and G\textsubscript{4492}A (Table 1).

**Influence of the point mutations on the replication of E9**

After transcription of the plasmids rE9-H.res and rE9-G.res in vitro, transfection experiments were performed using the DEAE method (Ausubel et al., 1994), and the resulting viruses were propagated in GMK cells. Sensitivity assays of these recombinants revealed a resistant phenotype as found for the original isolates, E9-H.res and E9-G.res (Fig. 4a). In both cases, virus replication was possible even at high inhibitor concentrations, indicating that the detected mutations in the 2C gene are relevant for drug resistance.
Fig. 3. Construction of infectious HBB- and guanidine-resistant E9 clones.
(a) Schematic diagram of the E9B wild-type clone (rE9B-fl.20). (b) HBB-resistant full-length clone (E9-H.res). The product of cDNA synthesis was cloned into rE9B-fl.20 using a unique ScaI restriction site. (c) Guanidine-resistant full-length clone (E9-G.res). The RT–PCR product was cloned into rE9B-fl.20 using unique NdeI and BglII sites. Genomes are arranged proportionally and the 2C coding region is marked by broken lines. The indicated mutations (a–d) are explained in detail in Table 1.

Table 1. Mutations of HBB-resistant (rE9-H.res) and guanidine-resistant (rE9-G.res) clones compared to the wild-type E9 2C region

<table>
<thead>
<tr>
<th>Variant</th>
<th>Mutation*</th>
<th>nt position†</th>
<th>Sequence divergence</th>
<th>aa position‡</th>
<th>aa divergence</th>
<th>Affected functional domain</th>
</tr>
</thead>
<tbody>
<tr>
<td>E9-H.res</td>
<td>a</td>
<td>4775</td>
<td>A → C</td>
<td>227</td>
<td>I → L</td>
<td>NTPase motif ‘C’</td>
</tr>
<tr>
<td></td>
<td>b</td>
<td>4782</td>
<td>C → U</td>
<td>229</td>
<td>A → V</td>
<td>NTPase motif ‘C’</td>
</tr>
<tr>
<td>E9-G.res</td>
<td>c</td>
<td>4286</td>
<td>A → G</td>
<td>64</td>
<td>E → G</td>
<td>—</td>
</tr>
<tr>
<td></td>
<td>d</td>
<td>4492</td>
<td>G → A</td>
<td>133</td>
<td>A → T</td>
<td>NTPase motif ‘A’</td>
</tr>
</tbody>
</table>

* See Fig. 3.
† Positions relative to whole virus genome.
‡ Positions relative to protein 2C.

Fig. 4. Sensitivity assays of the recombinant viruses containing both mutations (a) or single mutations (b and c).

(a) No inhibitor; ○, 73 µM HBB; ■, 146 µM HBB; ▲, 220 µM HBB; □, 1 mM guanidine; △, 2 mM guanidine. See text for nomenclature of constructs.

after a replication time of 4 or 6 h (Fig. 5a). Also, in contrast to the uninhibited samples, negative-strand RNA could not be detected at these time-points (Fig. 5b). In an analogous study, the replication assay was carried out using the recombinant viruses. The resistant mutants rE9-H.res and rE9-A1775C were able to replicate the RNA in the absence and presence of HBB,
Influence of HBB and guanidine on echovirus protein 2C

Recently, it has been shown that E9 protein 2C possesses ATPase/GTPase activity (Klein et al., 1999). With one exception, A\_133 G, the mutations of the HBB- and guanidine-resistant E9 variants are located within or near conserved protein domains of 2C which are proposed to be involved in NTP binding or splitting (Table 1). Therefore, the influence of HBB and guanidine on the NTPase activity of several GST–2C fusion protein constructs (GST2C-wt, GST2C-H.res, GST2C-A\_177 C, GST2C-C\_179 U and GST2C-G.res) was tested. In the presence of HBB, however, the ATPase activity of wild-type 2C, GST2C-H.res, GST2C-A\_177 C, and GST2C-C\_179 U was not altered, even in the presence of 1 mM HBB (Fig. 6). Also, guanidine did not abolish ATPase activity at physiologically relevant concentrations.

Besides the GST–2C fusion proteins, His-tagged proteins were constructed to minimize the possibility of unspecific reactions. Therefore, echovirus 2C was cloned into a pET-14b vector. After expression in E. coli BL21, the His-tagged proteins were purified to homogeneity by affinity chromatography under non-denaturing conditions. The His–2C proteins exhibited NTPase activity, and they also remained unaffected by HBB or guanidine (data not shown).

Discussion

In this study, we present data showing that the antiviral substance HBB inhibits at least positive-strand RNA synthesis of E9. In addition, an increase in negative strands could not be detected by Northern blotting techniques. Additionally, HBB and guanidine do not affect the NTPase activity of the non-structural protein 2C.

We have demonstrated that the picornavirus replication inhibitors HBB and guanidine seem to differ in terms of their precise mechanism of antiviral action. The limited or, more precisely, unidirectional cross-resistance of the two drugs and the fact that HBB and guanidine exhibit a synergistic action allow us to propose that the inhibitors act in a similar, but not identical way.

To elucidate the mechanisms of antiviral action of both substances at the molecular level, a panel of experiments was performed using drug-resistant E9B variants. For poliovirus, it is known that mutations crucial for resistance against or dependence on picornavirus replication inhibitors map to the coding region of 2C (Pincus et al., 1986; Pincus & Wimmer, 1986). In addition, previous work in our laboratory revealed that the only determinant for HBB dependence of E9B is located within the 2C gene (Hadaschik et al., 1999). Consequently, we focused our interest on this region of the viral genome. Indeed, two mutations were detected in the 2C coding region of each resistant variant and two of them are sufficient to induce a phenotype identical to the original resistant isolates. For guanidine-resistant or -dependent poliovirus, it has been revealed that most of the responsible amino acid substitutions are located at position 179 or 187, respectively (Baltera & Tershak, 1989; Tolskaya et al., 1994). However, in the case of E9, the crucial amino acid exchanges conferring resistance are located at different sites, i.e. position 133 (guanidine) and 227 (HBB), and may suggest different ways of drug inhibition (Table 1).
Fig. 6. ATPase activity of 2C proteins of E9 wild-type and the mutants in the presence of HBB or guanidine (Gua). After a standard ATPase assay procedure, the reactions were subjected to TLC followed by autoradiography. As controls, the reactions were driven with GST protein (lane GST) or no protein (lane --).

The exchange C<sub>4782</sub>U leads to an HBB-dependent phenotype (Fig. 4). It is noteworthy that the identical exchange has been determined in an independent project with the aim of isolating an HBB-dependent E9B variant (Hadaschik et al., 1999). Whether this constitutes mere coincidence – an unlikely possibility – or the only molecular mechanism for HBB dependence can only be elucidated by investigation of a larger number of dependent E9B isolates.

Three of the four mutations characterized in this project are located in the direct vicinity of two of three predicted NTP binding and splitting domains of 2C (Table 1). The NTPase activity of these domains was proven in previous work (Klein et al., 1999). However, 2C fusion proteins of both E9B wild-type and resistant mutants did not exhibit altered NTPase activity in the absence or presence of physiologically active inhibitor concentrations (Fig. 6). The weak inhibitory effect of 20 mM guanidine is probably due to the initiation of protein denaturation. Therefore, the NTPase activity of E9 2C does not appear to be a target for HBB or guanidine antiviral activity on E9. Recently, Pfister & Wimmer (1999) demonstrated that the NTPase function of poliovirus wild-type fusion protein GST–2C is inhibited in the presence of low guanidine levels, whereas the respective activity of a protein derived from a resistant mutant was unimpaired. These results may be a further hint that guanidine inhibits echoviruses and polioviruses in a different way.

Preliminary experiments allow the assumption that echovirus 2C binds ssRNA unspecifically, and again this 2C function does not seem to be influenced by HBB or guanidine (data not shown). Taken together, since neither the NTPase nor the RNA binding capacity of 2C could be affected by the inhibitors in vitro, our results support the conjecture that there may be an additional function of 2C acted upon by the drugs. A helicase activity as well as involvement in packaging have been discussed for poliovirus (Gorbalenya et al., 1990; Kadaré & Haenni, 1997; Li & Baltimore, 1990; Vance et al., 1997).

Because inhibition of RNA synthesis of E9 was proven at least for HBB (Fig. 5a, b), it is more likely that the helicase function is affected by the antiviral drugs, if such an activity is actually true for protein 2C.

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References


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