Rapid development of vaccine protection in macaques by live-attenuated simian immunodeficiency virus

Christiane StahI-Hennig, Ulf Dittmer, Thomas Nißlein, Harald Petry, Elke Jurkiewicz, Dietmar Fuchs, Helmut Wachter, Kerstin Mätz-Rensing, Eva-Maria Kuhn, Franz-Josef Kaup, Erling W. Rud and Gerhard Hunsmann

1 German Primate Centre, Kellnerweg 4, D-37077 Goettingen, Germany
2 Institute of Medical Chemistry and Biochemistry and Ludwig-Boltzmann-Institute for AIDS Research, University of Innsbruck, Innsbruck, Austria
3 Health Canada, Bureau for HIV/AIDS, Laboratory Centre for Disease Control, Ottawa, Canada

Convincing data on experimental vaccines against AIDS have been obtained in the simian immunodeficiency virus (SIV) macaque model by preinfection with a virus attenuated by a nef deletion. To investigate the efficacy of a nef deletion mutant of SIVmac32H called pC8 as a live-attenuated vaccine after shorter preinfection periods and to learn more about the nature of the immune protection induced, eight rhesus monkeys were infected intravenously with the pC8 virus. All monkeys became persistently infected, exhibiting low cell-associated viral loads, but strong cellular and, in terms of binding antibodies, strong humoral antiviral responses. Two of eight pC8-infected monkeys developed an immunodeficiency and were not challenged. Sequence analysis of their nef revealed complete replenishment of the deletion. The other six monkeys, two preinfected for 42 weeks and four for 22 weeks, were challenged with pathogenic spleen-derived SIV. Complete protection was achieved in four vaccinees. Virus was consistently detected in two vaccinees from the 22-week-group challenge, however, they remained clinically healthy over a prolonged period. Protection from challenge virus infection or a delayed disease development seemed to be associated with a sustained SIV-specific T helper cell response after challenge. Thus, a sterilizing immunity against superinfection with pathogenic SIV can be induced even after a relatively short waiting period of 22 weeks. Nevertheless, such a vaccine raises severe safety concerns because of its potential to revert to virulence.

Introduction

The most effective protection against animal viruses is obtained with live-attenuated viral vaccines. Since such vaccine viruses are replication-competent, they activate the host's overall immune response leading to long-lasting immune protection. Inactivated whole virus, split particles, or purified viral polypeptides or peptides seldom induce long-term protection without repeated booster immunizations.

The simian immunodeficiency virus (SIV) and human immunodeficiency virus type 2 (HIV-2) macaque models are highly suitable for vaccine trials relevant for HIV vaccine development. So far, candidate vaccines consisting of inactivated whole virus or viral subunits have been investigated in the SIV or HIV-2 macaque model with limited success in terms of sterilizing immunity. When challenged at the peak of the vaccine-induced immunity, monkeys immunized with killed whole (Desrosiers et al., 1989; Murphy-Corb et al., 1989; Sutjipto et al., 1990) or detergent-disrupted virus (Desrosiers et al., 1989; Stahl-Hennig et al., 1992), recombinant-expressed (Ahmad et al., 1994; Giavedoni et al., 1993; Mills et al., 1992) or virion-derived subunit vaccines (Lüke et al., 1993; Stahl-Hennig et al., 1994; Lüke et al., 1996) or animals primed with vector-based vaccines and boosted with recombinant subunit preparations (Franchini et al., 1995; Hu et al., 1992) were protected against challenge with SIV or HIV-2 to a varying extent. However, usually the protective immunity lasted for a few months only, in case of the prime/boost regimen up to 6 months (Franchini et al., 1995), and could be overcome by a heterologous virus challenge as well as high viral doses. Moreover, resistance against challenge virus not grown in human T cell lines has rarely been described (Stahl-Hennig et al., 1994; Lüke et al., 1996). Furthermore, none of the cited
studies convincingly demonstrated clear-cut immune correlates of protection, but in one report protection seemed to depend on the presence of neutralizing antibodies (Lüke et al., 1996).

Deletions in some auxiliary genes have been shown to weaken (Lang et al., 1993; Hoch et al., 1995) or abrogate SIV pathogenicity (Kestler et al., 1991; Rud et al., 1994a). The most convincing results with experimental AIDS vaccines have so far been produced by preinfecting macaques with live-attenuated SIV containing a nef gene deletion. In a first experiment with a virus lacking 182 bp in nef (Kestler et al., 1991) protection was achieved against a high dose of challenge virus after a 2-2.5 year waiting period (Daniel et al., 1992). Another attenuated SIV, a molecular clone of SIVmac32H called pC8, contains an in-frame deletion of four amino acids in nef (Rud et al., 1992) and obviously induced protection against cell-free and cell-associated virus challenge after much shorter preinfection periods (Almond et al., 1995; Whatmore et al., 1995; Norley et al., 1996). So far almost nothing is known about whether an immunological mechanism mediates the observed protection. Recent results on vaccine protection with a triple-deleted SIV (SIVd3) suggest a correlation with the level of specific neutralizing and binding antibodies (Wyand et al., 1996). On the other hand, data on the pathogenicity of this SIVd3 in neonatal macaques (Baba et al., 1995) and on the reversion to virulence of an SIV mutant attenuated by a smaller nef deletion (Whatmore et al., 1995) question the potential use of similar live-attenuated HIV in man. Nevertheless, studying both the host’s cellular and humoral immune response to such attenuated experimental vaccines should provide important clues to their mechanism of immune protection.

As part of a concerted European AIDS vaccine experiment using attenuated SIV and involving a total of 49 macaques for immunization, we have infected eight rhesus macaques (*Macaca mulatta*) with the above-mentioned naturally occurring nef-deletion mutant, pC8 (Rud et al., 1992). At the time of inoculation the vaccine virus was shown to have an attenuated phenotype in both Chinese rhesus monkeys (Rud et al., 1992) and cynomolgus macaques (*Macaca fascicularis*) (Rud et al., 1992, 1994a). Some time later we learned that the vaccine virus may revert to virulence in rhesus monkeys of Indian origin (Whatmore et al., 1995). Six preinfected monkeys were exposed to challenge with a highly pathogenic spleen-derived SIVmac virus swarm 22 or 42 weeks after pC8 infection. Primary infection led to complete protection in four out of six animals. The remaining two monkeys became infected with the challenge virus, but disease development seemed to be delayed. However, in two of the pC8-infected monkeys a reversion to virulence by a repair of the nef deletion was also observed.

**Methods**

- **Animals, clinical observation and necropsy.** Twelve purpose-bred rhesus monkeys of Indian origin between 2 and 3 years old with a body weight of 2.5–3.5 kg were obtained either from the breeding colony of the German Primate Centre or from the Cayo Santiago Island, Puerto Rico, USA. All monkeys tested negative for antibodies against STLV-1, simian type-D retrovirus and SIV. Physical examinations as well as bleedings including the determination of white blood cell (WBC) counts were performed on all macaques at regular intervals following first and second virus inoculation. Necropsy was conducted on monkeys developing severe AIDS-like symptoms and included routine gross pathology and histological examinations. One inguinal or axillary lymph node from each vaccinee was removed surgically under ketamine xylazine anaesthesia at 47 weeks post-challenge and subjected to virological, immunological, histological and molecular biological investigations.

- **Immunization and challenge.** Four rhesus monkeys were inoculated intravenously with 10⁸⁵ TCID₅₀ in 1 ml RPMI culture medium supplemented with 1% human AB-serum of a cell-free preparation of the molecular clone pC8 of SIVmac32H. This virus represents a naturally occurring SIV deletion mutant with a 12 bp deletion in the nef gene and three further nucleotide substitutions in the 3’ LTR overlapping nef encoding two conservative amino acid changes and one silent change (Rud et al., 1992). Twenty weeks later another set of four monkeys was inoculated with the pC8 virus as described above. The inoculum was grown in the human T cell line C81–66. Its attenuated phenotype for rhesus and cynomolgus macaques has been reported recently (Rud et al., 1994a). After another 22 weeks six of the eight preinfected monkeys, all four from the second group (22-week-group) and two from the group infected first (42-week-group) together with four naive control monkeys were challenged intravenously with 50 median monkey infectious doses (MID₅₀) of SIVmac32H/spl. This challenge virus was prepared from the spleen of a vaccinated but unprotected rhesus monkey after challenge with SIVmac32H (Cranage et al., 1990) and was never kept in vitro. Details of this virus stock are described elsewhere (Lüke et al., 1996). The remaining two monkeys from the 42-week-group were not challenged because they had developed signs of immunodeficiency (Dittmer et al., 1995b).

- **Determination of viral load.** For virus recovery from citrated blood, monkey peripheral blood mononuclear cells (MPBMC) were separated by centrifugation on Ficoll density gradients (Histopaque; Sigma-Aldrich) at 1000 g for 15 min in Leucoprep separation tubes (Greiner, Tübingen, Germany) without predilution of the blood. To determine the cell-associated viral load, cocultures were initiated by mixing 5 x 10⁶, 1 x 10⁶ and 3 x 10⁶ MPBMC with 3 x 10⁶ C81–66 cells each in 50 ml tissue culture flasks. Twenty-four-well tissue culture plates were used when 2.5 x 10⁵ to 122 MPBMC were cocultured with 3 x 10⁶ C81–66 cells in 2 ml medium per well. Cells were diluted every 3 to 4 days (cultures split in flasks in a ratio of 1:3, the well cultures 1:2) and assessed for virus-induced cytopathic effect (CPE). Viral replication obvious by CPE was confirmed by detection of viral antigen in culture supernatants with an HIV-1 antigen-capture assay (Innogenetics). Cultures were regarded as virus-negative if after 4 weeks of cultivation neither CPE nor supernatant antigen could be detected. The lowest MPBMC dilution leading to virus replication was the endpoint, expressed as the number of infectious cells per 1 x 10⁶ MPBMC. Low viral load was defined as 0.33–2 infectious cells per 10⁶ MPBMC whereas 4–64 and 128–4096 per 10⁶ MPBMC indicated moderate or high viral load.

Mononuclear cells from lymph nodes were obtained by passing the organs through 200 μm stainless steel mesh. One part of the recovered lymphocytes was subjected to the virus isolation protocol described above; another part was stained for flow cytometry analysis and a third part was analysed by PCR.

Antigenaemia was determined by a commercial SIV p27-capture assay (Coulter).
virions were measured by an ELISA using disrupted SIVmac32H in optimal concentrations for antigen coating (Stahl-Hennig et al., 1992).

Antibodies against structural viral proteins of SIVmac were evaluated by Western blotting (WB) as described previously (Stahl-Hennig et al., 1992) applying the antigen used for ELISA.

Nef antibodies were detected by WB performed with lysates of CV-1 cells infected at an m.o.i. of 1 p.f.u. per cell of a recombinant vaccinia virus (VV) encoding the nef gene of the full-length molecular clone pJ5 of SIVmac32H (kindly provided by M. Mackett and B. E. Clarke, Department of Molecular Sciences, Wellcome Research Laboratories, Langley Court, Beckenham, UK). Infected cells were harvested after 24 h, resuspended in PBS, washed and lysed by three freeze-thaw cycles. Lysates were separated by SDS-PAGE and processed as described previously (Stahl-Hennig et al., 1992). Monkey sera were diluted 1:10 and incubated with Nef-containing nitrocellulose strips. As one of the positive controls a mouse monoclonal antibody (MAB) reacting with pJ5 Nef (KK75; kindly provided by Karen Kent, NIBSC, UK) was included.

A sensitive MT4 cell-killing assay was employed to detect SIV-neutralizing serum antibodies (Harada et al., 1986; Jurkiewicz et al., 1992).

Cell-mediated immunity, T cell subsets and urinary neopterin concentrations. During the investigation period both the T helper (TH) cell and cytotoxic T lymphocyte (CTL) response were investigated regularly (Dittmer et al., 1994, 1995b). For proliferation assays MPBMC were stimulated with purified heat-inactivated whole SIVmac32H. The CTL response of the pC8-infected monkeys was measured in a commercial CytoTox98 non-radioactive assay (Promega).

Autologous B-lymphoblastoid cell lines infected either with recombinant VV expressing GagPol, Env or Nef proteins of SIVmac32H(pJ5) (Rud et al., 1994) or with wild-type VV served as target cells. Lymphocyte subsets were determined by labelling Ficoll-separated MPBMC with fluorescein- or phycoerythrin-conjugated mouse MABs specific for CD4 (OKT4; Ortho Diagnostic Systems) or CD8 (Leu 2a; Becton Dickinson). Such labelled cells were analysed with an EPICS Profile 1 flow cytometer (Coulter).

Urinary neopterin is a marker of immune activation (Fuchs et al., 1988) and has been shown to peak between 10 and 15 days after SIV infection (Stahl-Hennig et al., 1992; Dittmer et al., 1995a; Hoch et al., 1995). It rises steeply in most SIV-infected monkeys shortly before AIDS-related death (Dittmer et al., 1995a; Hoch et al., 1995; C. Stahl-Hennig, unpublished data). Neopterin was determined in early morning urine (Fendrich et al., 1995a; Hoch et al., 1995; C. Stahl-Hennig, unpublished data). The early detection of virus in blood coincided with peak urinary neopterin levels between 10–15 days p.i. (Fig. 2).

The two- to fourfold neopterin increases of individual baseline levels were lower than those 7- to 20-fold ones observed after infection with pathogenic SIV (Fig. 2, and C. Stahl-Hennig, unpublished data).

All infected monkeys rapidly developed SIV-specific antibodies. By 4 to 8 weeks p.i. their sera already contained antibodies against all viral structural proteins when investigated by WB (data not shown). The increase in specific antibody titres was documented by titration of serially taken sera in a whole-virus ELISA. Titres rose steeply 8 to 12 weeks p.i. to 12,500–50,000 (Fig. 3).

In all macaques a SIV-specific TH cell reactivity appeared lasting from 4 to 20 or 40 weeks after pC8 infection. We have never observed such a reactivity in monkeys infected with pathogenic SIV. Moreover, CTL activities against different SIV polypeptides including Nef were detectable at a varying degree in 6 out of 7 animals examined (for details see Dittmer et al., 1995b). In short, all these data suggested a low-grade infection of rhesus monkeys well-controlled by a specific cellular immune response during the first 20 weeks following pC8 infection.

Two pC8-infected rhesus monkeys developed signs of immunodeficiency

These two monkeys (Mm 1820 and Mm 1823) belonged to the 42-week-group. The immunodeficiency appeared between week 28 and 40 after pC8 infection. It was diagnosed by several independent criteria and was associated with an early repair of the nef deletion (for details see Dittmer et al., 1995b). However, at the time of challenge, their cell-associated viral loads (Fig. 1a) and ELISA antibody titres (data not shown) were indistinguishable from those of the other six pC8-infected monkeys maintaining the nef deletion. Interestingly, only these two nef reverters developed Nef-specific antibodies between 8 and 50,000.
Fig. 1. Cell-associated viral load in rhesus macaques following (a) infection with the pC8 virus and (b) after challenge with the pathogenic spleen-derived SIV. Virus was isolated from MPBMC by a limiting dilution coculture technique and the endpoint was determined. Viral loads are expressed as the number of infectious cells per 10⁶ MPBMC. The respective animal groups as well as each individual animal number and death (d) of the animal are indicated. The dotted line marks the cut-off value of the assay. Cultures below the cut-off are considered negative. In animals marked with an asterisk challenge virus was detectable. At one time point not all animals were investigated (nd, not done).
Fig. 2. Urinary neopterin levels during the preinfection period and after challenge in preinfected and non-infected rhesus monkeys. Neopterin values obtained after first infection or challenge were related to preinfection data and expressed as the n-fold of the mean of six to ten neopterin concentrations determined independently before infection. The diagram shows the neopterin concentrations of the non-infected monkeys (Controls), of those preinfected for 42 weeks (42-week-group), and those preinfected for 22 weeks (22-week-group). Individual animal numbers are indicated. For the first 30 days mean values for 5 day intervals are shown; for days 31–40 mean values of a 10 day interval, for days 41–60 of a 20 day interval, for days 61–180 of a 30 day interval and then for 60 day intervals are shown. The dotted line indicates the time of challenge and ‘d’ the death of the animal. In animals marked with an asterisk challenge virus was detectable.

and 24 weeks after pC8 infection. These antibodies persisted for the remaining observation period until 98 weeks p.i. (Table 1). Neopterin concentrations were elevated only slightly (twofold in Mm 1820) or moderately (four- to fivefold over 360 days in Mm 1823). The latter animal also exhibited clinical alterations, e.g. progressing peripheral lymphadenopathy
Table 1. Nef-specific antibody response in pC8-infected rhesus macaques before and after challenge

The antibody responses were scored as follows: +++, strong positive; +, positive; −, negative by immunoblot.

<table>
<thead>
<tr>
<th>Reaction with Nef protein</th>
</tr>
</thead>
<tbody>
<tr>
<td>1820⁺</td>
</tr>
<tr>
<td>---</td>
</tr>
<tr>
<td>Post-infection</td>
</tr>
<tr>
<td>0</td>
</tr>
<tr>
<td>4</td>
</tr>
<tr>
<td>8</td>
</tr>
<tr>
<td>16</td>
</tr>
<tr>
<td>22/24</td>
</tr>
<tr>
<td>32</td>
</tr>
<tr>
<td>42</td>
</tr>
<tr>
<td>Post-challenge</td>
</tr>
<tr>
<td>4</td>
</tr>
<tr>
<td>8</td>
</tr>
<tr>
<td>12</td>
</tr>
<tr>
<td>16</td>
</tr>
<tr>
<td>24</td>
</tr>
<tr>
<td>36</td>
</tr>
<tr>
<td>48</td>
</tr>
</tbody>
</table>

* Macaques exhibiting signs of immunodeficiency and exempted from challenge.
† Macaques in which challenge virus was detectable.
‡ Sera obtained 90 weeks after infection with pC8.
ND, Not done; NA, data not available because of the shorter preinfection period.

beginning at 42 weeks p.i. and a splenomegaly observed by 62 weeks p.i. Both monkeys remained persistently virus positive over the whole observation period, but at invariably low levels (0–33–2 infectious cells/1 × 10⁶ PBMC). Since the alterations described indicated the development of immunodeficiency these two monkeys were not challenged.

The immune status of the non-reverters was homogenous at the time of challenge

None of the virological or immunological parameters investigated in the six remaining pC8-infected monkeys discriminated them from each other at the time of challenge. They all had maintained the nef deletion as shown by sequencing of respective PCR products (data not presented). Four of the six monkeys were virus isolation positive but had low viral loads (Fig. 1b). Their antibody titres against whole SIV had reached 50 000 to 400 000, but neither neutralizing (data not shown) nor Nef antibodies (Table 1) were detectable. All monkeys were healthy and maintained normal CD4/CD8 ratios (above 1; data not shown). Their neopterin levels remained close to the baseline (Fig. 2) and all monkeys exhibited a considerable SIV-specific TH cell reactivity with stimulation indices ranging between 4 and 6 (Fig. 4). Unfortunately, CTL data were not available at time of challenge. However, 4 weeks before challenge two of five monkeys examined showed a clear CTL activity against the structural SIV proteins, in one case also against Nef (for details see Dittmer et al., 1995 b).

Challenged animals displayed three distinct patterns of reactivity

The six pC8-infected monkeys without signs of immunodeficiency were challenged along with four naive controls with 50 MIF₅₀ of the spleen-derived pathogenic SIVmac, four animals at 22 weeks p.i. and two animals at 42 weeks p.i. The four control monkeys became productively infected thereby presenting one pattern of reactivity. Two weeks post-challenge proviral DNA was recovered from their PBMCs containing high viral loads (Fig. 1b) and viral plasma antigen was detected (data not shown). All controls maintained the high cell-associated viral load until their death between 17 and 35 weeks post-challenge (Fig. 1b). Three of them were euthanized in advanced AIDS stages. Pathological findings were giant cell disease, encephalopathy and opportunistic infections with Pneumocystis carinii and Cryptosporidium. The remaining animal (Mm 7073) had to be killed due to an irreparable rectal prolapse. However, at necropsy this monkey also showed AIDS-related alterations, e.g. a severe generalized
Fig. 3. SIV-specific antibody titres of sera obtained during the preinfection period and after challenge from preinfected and non-infected rhesus macaques. Antibodies were determined by ELISA using whole lysed SIV as antigen. The diagram shows the titres of the non-infected monkeys (Controls), of those monkeys preinfected for 42 weeks (42-week-group), and of those preinfected for 22 weeks (22-week-group). Times of preinfection and challenge (dotted line), death (d) of the animal as well as symbols for individual animal numbers are indicated.

lymphadenopathy confirmed histologically by a severe hyperplasia of the germinal centres as well as opportunistic infections in the intestine. Furthermore, three of the controls seroconverted against SIV antigen. However, one monkey from this group (Mm 7071) remaining ELISA (Fig. 3) and WB (data not shown) antibody-negative was severely ill by 17
Fig. 4. For legend see facing page.
weeks p.i. and had to be euthanized. In addition, in this group a typical initial neopterin increase was recorded which was 9- to 12-fold higher than baseline levels (Fig. 2).

By contrast, only two of the six vaccinees (Mm 1798 and Mm 1799) were evidently superinfected, thus forming a second pattern of reactivity. They had been challenged 22 weeks p.i. The earliest indicator of replicating challenge virus was the complete replenishment of the nef deletion by 2 weeks p.i. shown by direct sequencing of nef PCR products. The respective nucleotide sequences were identical to those of the control monkeys and the original pJ5 sequence (Rud et al., 1994a) (data not shown). Despite the presence of full-length nef in these two monkeys, no other early signs of infection with the spleen-derived challenge virus were observed. The animals had low cell-associated viral loads (Fig. 1b) and did not show a neopterin increase (Fig. 2), antigenaemia (data not shown) or an anamnestic antibody response when compared with the controls. The slightly rising antibody titres in these two monkeys of 1–2 log₂ steps after challenge were attributable to still rising antibody titres after the primary infection. This became evident when the titres were compared with the antibody kinetics of the vaccinees from the 42-week-group.

Only 4 to 8 weeks after challenge, cell-associated viral loads rose to moderate levels in the two vaccinees containing virus with intact nef (Fig. 1b). One monkey (Mm 1798) maintained this level until the end of the observation period, the other (Mm 1799) reached high levels by 36 weeks p.i. However, these elevated levels were still approximately eightfold lower than the mean viral load of the controls (1850 infectious cells per 1 × 10⁶ PBMC) determined over 36 weeks (Fig. 1b). Like the two pC8-infected but immunodeficient monkeys these two vaccinees replicating SIV with full-length nef also mounted anti-Nef antibodies by 8 to 16 weeks p.i. (Table 1). Additionally, in one of these two monkeys (Mm 1798) the CD4/CD8 ratio dropped slightly by 40 weeks p.i. (data not shown). In spite of these changes both vaccinees remained clinically healthy.

In the other four preinfected monkeys superinfection could not be detected, thus representing the third reactivity pattern. All of them were still replicating the nef deletion mutant by 47 weeks p.i. (data not shown). Their cell-associated viral load (Fig. 1b) and urinary neopterin concentrations (Fig. 2) remained invariably low. They showed no secondary antibody response (Fig. 3) and their clinical condition was normal. Unexpectedly, one monkey (Mm 1822) from this group in which only nef-deleted virus was detectable, generated persisting antibodies against Nef by 12 weeks post-challenge (Table 1) although all other parameters remained unchanged.

Monkeys sustaining a proliferative TH cell response after challenge were protected to various extents

To examine a possible influence of the TH cell immune response on the post-challenge period we continued measuring SIV-specific TH cell proliferation after challenge. All vaccinees maintained their virus-specific TH cell proliferative activity over 28 to 44 weeks post-challenge (Fig. 4). However, by 52 weeks post-challenge this TH cell function was no longer detectable. Unexpectedly, the two obviously superinfected vaccinees (Mm 1798 and Mm 1799) also showed a long-lasting proliferative response (Fig. 4), but each to a different extent. In one superinfected vaccinee (Mm 1799) even a sort of booster effect was observed. Its stimulation indices rose as high as 12 between 20 and 24 weeks post-challenge. Both monkeys showed no obvious clinical alterations except a mild lymphadenopathy and palpable splenomegaly in one monkey (Mm 1799) by 40 weeks post-challenge. The four completely protected animals kept their TH cell responsiveness on a low to moderate level up to 44 weeks post-challenge indicating an intact function of this cellular immune compartment.

Results from physical and blood examination reflected lymph node histology and viral status

A superficial lymph node from each vaccinee was surgically removed at 47 weeks post-challenge to test for viral load, genotypic characteristics of retained virus and for morphological alterations in the lymphatic periphery. Results from investigations with lymph node mononuclear cells (LNMC) were compared with those from simultaneously prepared MPBMC. The cell-associated viral load was invariably low (< 0.3–0.5 infectious cells per 10⁶ LNMC) in the lymph nodes of the four vaccinees obviously protected from challenge infection (Mm 1821, Mm 1822, Mm 1825 and Mm 1828). Generally, the viral load of these animals was similar in blood and lymph node. One animal (Mm 1828) was even virus negative in both PBMC and LNMC. nef sequence analysis of proviral DNA from LNMC only demonstrated the pc8 genotype, but no evidence for superinfecting virus. The lymph nodes had an intact histological architecture with a mild to moderate activation similar to healthy animals. In contrast, in the two vaccinees replicating challenge virus with full-length nef, cell-associated viral loads of LNMC were as high as or even ten times higher (Mm 1798) than those of the PBMC (256 and 128 infectious cells per 10⁶ LNMC versus 256 and 16 infectious cells per 10⁶ PBMC). Sequence analysis of proviral DNA from LNMC confirmed the presence of wild-type SIV. Clinically Mm 1798 developed a mild splenomegaly and
lymphadenopathy which was consistent with a histologically detectable moderate follicular hyperplasia of the germinal centres found in one lymph node of this animal. Apart from these mild alterations both monkeys were clinically stable until 56 weeks post-challenge (time point of sacrifice).

**Discussion**

Two different attenuated SIV nef deletion mutants and a triple-deleted SIV conferred protection against challenge after varying waiting periods of 8 weeks to 2.25 years (Daniel et al., 1992; Almond et al., 1995; Whatmore et al., 1995; Norley et al., 1996; Wyand et al., 1996). The mechanism responsible for protection still remains obscure. However, in none of these experiments had the cellular immune response been analysed. The present study using the attenuated SIVmac32H(pC8) (Rud et al., 1994a) for preinfection had a double purpose: firstly, to investigate whether shorter intervals after the initial infection would still be sufficient to allow development of protection against a pathogenic virus swarm; and secondly to elucidate the mechanisms mediating the protective immunity.

**The initial high viral load appears to be immunologically contained in pC8-infected macaques**

We inoculated eight rhesus monkeys with the pC8 virus. The deletion in Nef of four amino acids (Rud et al., 1992) is much smaller than that of the mutated SIV described earlier (Kestler et al., 1991) but still rendered the virus apathogenic (Rud et al., 1992). All monkeys became readily infected by the pC8 virus as assessed by several independent criteria. An unexpected finding was that despite attenuation of the virus by the nef deletion the initial cell-associated viral loads were as high as those measured in monkeys infected with pathogenic SIV. In an earlier report (Daniel et al., 1992) viral load had only been examined 14 weeks p.i. when those monkeys had obviously already cleared the virus from the periphery. The viral loads of our monkeys also dropped and virus isolation became only intermittently positive 12 weeks p.i. but three monkeys maintained a persistent low-level viraemia. Furthermore, we were able to measure a long-lasting cellular immune response in the pC8-infected macaques. Six out of seven monkeys examined had developed an antiviral CTL activity (Dittmer et al., 1995b) indicating virus replication (Stahl-Hennig et al., 1992). Most interestingly, all infected macaques generated an SIV-specific TH cell response lasting over the respective waiting periods in six of the eight monkeys. Therefore our finding fits the observation of early over-expression of T cell cytokines in pC8-infected monkeys compared to no or rather a down-regulation of expression in monkeys infected with pathogenic SIV (Benveniste et al., 1996). We had earlier observed a similar proliferative TH cell reactivity in animals infected with apathogenic HIV-2 (Dittmer et al., 1994) or SIV_AVR (Hoch et al., 1995) but never after infection with highly pathogenic SIV strains (Dittmer et al., 1994). This underlines the attenuation of the pC8 virus.

**Two monkeys with reverting pC8 virus are slow progressors**

Unexpectedly, two pC8-infected monkeys developed signs of immunodeficiency (Dittmer et al., 1995b) and therefore were exempted from challenge. However, in these two animals only mild clinical symptoms developing later than in the controls were observed during 2 years of follow-up. Sequential analysis revealed a partially incorrect but functionally complete reversion to full-length nef by 12 weeks p.i. (Dittmer et al., 1995b). In one monkey we were able to demonstrate a duplication of the regions flanking the deletion at 10 weeks p.i. (data not shown). Such reversion to virulence by molecular evolution has been demonstrated recently for rhesus monkeys of Indian origin (Whatmore et al., 1995) but not for Chinese rhesus monkeys (Rud et al., 1992, 1994a) or for cynomolgus macaques (Rud et al., 1994a). Thus, it seems that in Indian rhesus monkeys, which are known to control SIV replication less effectively than Chinese rhesus monkeys (Joag et al., 1994 and C. Stahl-Hennig, unpublished data), reversion of pC8 to virulence is a more likely event.

**A longer waiting period warrants better protection**

By contrast, the other six pC8-infected monkeys remained normal by all laboratory and clinical criteria examined and were challenged with the pathogenic ex vivo derived SIV (Stahl-Hennig et al., 1994; Lüke et al., 1996) at 22 or 42 weeks p.i.

All vaccinees were primarily protected against the pathogenic consequences of the challenge virus while the four control animals rapidly progressed to immunodeficiency. Three of them had to be euthanized in a moribund condition within six months after challenge. By 16 weeks post-challenge two vaccinees from the 22-week-group exhibited median levels of cell-bound viral load lasting throughout the observation period. In these two monkeys virus with full-length nef was already detected by 2 weeks post-challenge. Since the nef sequences were identical to that of the challenge virus and the nef repair takes several weeks (Whatmore et al., 1995) we conclude that the challenge virus rather than a late revertant replicated in these two vaccinees. However, in these two animals disease development was apparently delayed since only a few very mild pathogenic changes were observed in the later phase of their superinfection. Although such a conclusion is difficult to draw due to the small number of animals involved, a mortality rate of 60% in 20 historical control animals within the first year after infection with our spleen-derived SIVmac makes this assumption more likely.

We have observed one of the shortest waiting periods so far required for the induction of complete protective immunity after experimental infection with an attenuated SIV. Sterile
immunity was already present at 22 weeks p.i. whereby earlier results were confirmed and extended (Daniel et al., 1992; Almond et al., 1995; Whatmore et al., 1995; Norley et al., 1996; Wyand et al., 1996).

The protection seems to be mediated by cellular immune responses and mostly correlates with lack of Nef antibodies

Several functional and non-functional immune parameters were examined before and after challenge in order to find immunological correlates of protection. In the present trial, neutralizing antibodies did not seem to be important for protection although our failure to detect neutralizing antibodies might be due to the method applied. However, the role of vaccine virus-induced neutralizing antibodies is contradictory (Daniel et al., 1992; Almond et al., 1995; Norley et al., 1996; Wyand et al., 1996). In contrast, we were able to repeatedly measure cytotoxic T cells against various SIV proteins in the pC8-infected animals before challenge (Dittmer et al., 1995b). These effector cells might contribute to the protection observed. Remarkably, the vaccine virus infection induced a long-lasting virus-specific TH cell response which, however, did not discriminate between protected and unprotected vaccinees. We presume that this immune function plays a crucial role in the protection either against primary challenge virus replication (Stahl-Flenig et al., 1992) or at least against the fatal consequences following infection with pathogenic challenge virus (Petry et al., 1995). Such a TH cell reactivity may also have suppressed disease development in those monkeys in which primary challenge virus replication was not completely prevented by vaccination (Shafferman et al., 1991; Israel et al., 1994) but was perhaps suppressed by neutralizing antibodies. Surprisingly, chronic infection of macaques with the pathogenic pJ5 virus also appeared to confer protection against superinfection with the attenuated pC8 virus. Unfortunately, the identity of virus reisolated after this challenge was unknown and no data on cellular immunity was presented (Rud et al., 1994b). However, the pJ5 virus has been reported to be only moderately pathogenic with a mortality rate of less than 15% within 1 year (Rud et al., 1994a). Therefore, even in such infected macaques a certain TH cell reactivity cannot be completely ruled out which might have contributed to the protection observed in the above-mentioned challenge experiment.

Our hypothesis is further supported by recent findings from retroviral infections of mice. Immunization of mice with VV recombinants containing TH and B cell epitopes from the envelope glycoprotein of Friend murine leukaemia virus conferred protection in mice. Additional inclusion of an immunodominant CTL epitope in the vaccine did not abrogate the fatal effects of systemic CD4+ cell depletion, thus underlining the critical role of TH priming for retrovirus immunity (Hasenkrog et al., 1996). In another experiment it was shown that a peptide from the gp70 C-terminal TH epitope alone was sufficient to protect mice against subsequent Friend virus infection (Miyazawa et al., 1995).

The pC8 virus only carries a small nef deletion but it was not clear whether Nef is expressed in infected monkeys. Sera serially taken from all eight pC8-infected monkeys after first or second inoculation were examined for Nef-specific antibodies. Formation of anti-Nef antibodies seemed to depend on the expression of full-length Nef in four monkeys, i.e. in the two monkeys repairing nef early and the two vaccine failures. Thus, for antibody production Nef must be either presented in its native tertiary structure which perhaps is altered in the deleted form. Alternatively, the immunodominant epitope was located in the deleted region. Surprisingly, one completely protected monkey also generated Nef-specific antibodies although neither in PBMC nor in the lymph node was there any evidence for detectable full-length nef. In this case, amplification of nef-specific proviral sequences always generated slightly smaller fragments compared to PCR products from animals infected with the challenge virus (data not shown) thus hinting at the presence of nef in its deleted form. However, one could argue that direct sequencing of PCR products might have failed to detect a minor virus subpopulation representing either late reverted pC8 virus or the challenge virus in this animal but which was nevertheless sufficient for Nef antibody induction. Such an assumed subpopulation could be contained by Nef-specific CTL which have previously been reported to influence viral replication (Gallimore et al., 1995). Another explanation of these unexpected anti-Nef antibodies could be that the induction of such an antibody response needs a persistent viral replication on a certain as yet unidentified level. However, if this proves to be so, such antibodies should have been detected in this obviously protected animal before and not only after challenge.

Nef-deleted viruses as candidate vaccines have been met with justified safety concerns since even attenuated SIV with a triple deletion appears to be pathogenic for neonatal macaques (Baba et al., 1995). Moreover, we and Whatmore et al. (1995) have observed that viruses with small deletions can more easily revert to virulence. Thus, a safe and effective AIDS vaccine should induce the protective immunity of live-attenuated virus including a long-lasting cellular immune response but should not contain replication-competent retroviruses.

We are grateful to Dr M.P. Cranage, Centre for Applied Microbiology and Research, Porton Down, UK for providing the pC8 virus. Dr. M. Mackett and Dr B.E. Clarke, Department of Molecular Sciences, Wellcome Research Laboratories, Langley Court, Beckenham, UK and the EU-Programme EVA are acknowledged for providing the vaccinia virus recombinants expressing Nef (VV9011), Env (VV9002) or Gag/Pol (VV9019) of SIVmac32H(pJ5). We thank K. Borchardt, K. Heinrichs, K. Wäse and U. Zedler for excellent technical assistance. This work was funded by the European Community, Concerted Action on Monkey Models for AIDS Research (grant No. MRA*-042-D).
References


Received 29 May 1996; Accepted 7 August 1996