Expression of the herpes simplex virus type 1 glycoprotein E in human cells and in *Escherichia coli*: protection studies against lethal viral infection in mice

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The objective of this study was to examine the protective efficacy of purified recombinant herpes simplex virus type 1 (HSV-1) glycoprotein E (gE-1) in the mouse lethal challenge model. A secreted form of gE-1 (hgE-1s) protein, containing amino acids 1-406, was produced in human cells by using the episomal replicating pRP-RSV expression vector. In addition, a portion of the gE-1 (bgE-1t) protein corresponding to amino acids 90-406, was expressed in *Escherichia coli* as a fusion protein with maltose binding protein using the pMAL-c2 expression vector. Mice vaccinated with hgE-1s developed high serum titres of HSV-1-neutralizing antibodies and were significantly protected from intraperitoneal lethal HSV-1 challenge, whereas mice vaccinated with bgE-1t exhibited only moderate levels of protective immunity. These results demonstrate that the expression of gE-1 in human cells has a strong impact on its protective efficacy and that most importantly the hgE-1s protein could be of value as a component of an HSV multi-subunit vaccine.

Herpes simplex virus (HSV) glycoproteins appear to be promising candidates for subunit vaccine development against HSV infection (Burke, 1993; Stanberry et al., 1989). Of the eleven known glycoproteins, gD, gB and gC were the first antigens to be expressed in various expression systems and to be shown to provide encouraging results in animal models of HSV disease (Laurence et al., 1984; Pachl et al., 1987; Krishna et al., 1989; Weir et al., 1989; Kino, 1990; Manservigi et al., 1990; Burke, 1991; Ghiasi et al., 1991, 1992a). Moreover, a number of clinical trials using gB and/or gD are in progress and promising results have been reported (Straus et al., 1994; Stanberry, 1995; Langenberg et al., 1995). On the other hand, it has been suggested that better adjuvants and incorporation of other glycoproteins in addition to gD, gB and gC might improve the efficacy of subunit vaccine formulation in humans (Burke, 1993). Consequently, it would be essential to evaluate the antiviral immunity induced by other HSV glycoproteins and to conduct comparative studies.

The HSV-1 glycoprotein E (gE) may be of particular value for the development of an HSV subunit vaccine. Glycoprotein E, alone or in combination with gI, acts as a viral Fc receptor that can bind the Fc portion of immunoglobulin G (Baucke & Spear, 1979; Para et al., 1982; Johnson & Feenstra, 1987; Johnson et al., 1988; Frank & Friedman, 1989; Bell et al., 1990; Dubin et al., 1994). This activity may protect infected cells from antibody-dependent cellular cytotoxicity (Dubin et al., 1991) by blocking some functions requiring Fc binding (Adler et al., 1978). Most interestingly, recent studies have indicated the gE-1 protein to be responsible for cell-to-cell virus spread (Balan et al., 1994; Dingwell et al., 1994).

The efficacy of the HSV-1 gE protein as a potential component of a subunit vaccine has been tested previously. At first, Blacklaws et al. (1990) reported that a gE-1 protein, expressed in vaccinia virus recombinants, failed to induce protective immunity and gave only a weak neutralizing antibody response when tested in mice. Subsequent studies, based on baculovirus recombinants expressing gE showed that extracts of whole insect cells infected by the gE/baculovirus recombinants can confer protection against HSV-1 challenge infection...
in animals and induce high levels of neutralizing antibodies (Ghiasi et al., 1992b, 1994a). Provided that the results from those studies were variable and none of them utilized purified gE-1 recombinant protein, we considered evaluating the immunogenicity and protective efficacy of gE by the use of a secreted form of gE expressed in human cells and a purified truncated form of gE expressed in Escherichia coli, utilizing the mouse lethal challenge model. Various expression systems, including mammalian cell lines and E. coli, have been used for the evaluation of the antiviral immunity provided by immunization with other HSV glycoproteins, such as gD, gB and gC. However, parallel experiments to evaluate the efficacy of an antigen produced by different systems has never been conducted (Burke, 1993). This represents the first report on the evaluation of the antiviral properties of a recombinant gE antigen expressed in human cells and in E. coli.

For the expression of gE-1 in E. coli, the pMAL-c2 (New England Biolabs) expression vector was used to construct plasmid pHPI427. For this purpose plasmid pHPI400 was first constructed by cloning the 2.1 kb NruI–BamHI fragment (nt 140579–142746) from pRB123 (Post et al., 1980), carrying most of the gE coding sequences (aa 1–501) into the HincII–BamHI site of pGEM-3Zf+ (Promega). Subsequently, the 1.3 kb ApaI–ApaI fragment (nt 141131–142463) from pHPI400, containing nucleotides from -40 to +1291 relative to the transcriptional initiation site of gE, was cloned into the SmaI site of pUC19 to yield pHPI402. Finally, the 980 bp SphI–SphI fragment from pHPI402, containing nucleotides from +338 to +1291 from the transcription start of the gE-1 gene coding for aa 90–406 of the protein, was ligated into the XmnI site of the pMAL-c2 vector in frame to the C-terminal portion of the maltose binding protein (MBP) coding gene (malE), to yield pHPI427. This part of the gE protein forms most of the extracellular domain of the protein and lacks the signal peptide, which might be toxic for E. coli (Rose & Shafferman, 1981). The expression of the fusion protein was first tested by separation of E. coli protein cell extracts on a Coomassie blue-stained SDS-PAGE gel. As shown in Fig. 1(a), a band of 78 kDa, corresponding to the predicted molecular mass of the MBP–gE fusion protein, was clearly detected in lysates of E. coli cells harbouring the pHPI427 plasmid but was absent in cell lysates harbouring the pMAL-c2 vector alone (lanes 3 and 1 respectively). The identity of the 78 kDa band was further confirmed on a Western blot probed with the gE monoclonal antibody II 481-A6, kindly provided by P. Spear, (Fig. 1b, lane 1) verifying the correct translation of the chimeric protein. Subsequently, for the large-scale preparation of the E. coli expressed MBP–gE-1t (bgE-1t) fusion protein, E. coli XL1-Blue cells harbouring plasmid pHPI427 were induced to synthesize the MBP–gE-1t fusion protein by the addition of IPTG and the MBP–gE-1t fusion protein was purified from the bacterial cell extracts by employing an amylose resin affinity column (New England Biolabs). The eluted protein was
dialysed against PBS and its purity was checked on a 10% SDS–PAGE gel by Coomassie blue staining (Fig. 1c). Purified protein was quantified using the Bradford assay. The yield was 15 mg of purified protein per litre of bacterial culture. The purified fusion protein was treated with factor Xa (New England BioLabs) in order to cut the gE-1 portion of the chimeric protein from the bacterial MBP. The MBP–gE-1t protein was used to produce an anti-gE polyclonal antiserum (designated as V3) in New Zealand white rabbits which reacted specifically with the HSV-1 gE in immunoblot and immunoprecipitation tests. Both the intact MBP–gE-1t and the MBP–gE-1t treated with factor Xa proteins were used in the vaccination experiment.

It has been suggested that the proper post-translational modification and folding of a recombinant protein are critical for the effectiveness of a subunit vaccine. In addition it has been reported that gB and gD expressed in mammalian cells are properly folded and readily secreted into the culture medium when the transmembrane region is deleted (Berman et al., 1984; Manservigi et al., 1990) and this results in a biologically active, easily purified antigen for subunit vaccine formulation. Thus, to achieve high levels of expression and secretion of an antigenically authentic form of gE-1, we chose to express a truncated derivative of gE (hgE-1s) in human cells using the episomal replicating vector pRP-RSV (Manservigi et al., 1990). The 1·3 kb ApaI–ApaI fragment from pHPi400, described above, was ligated into the BamHI cloning site of the pRP-RSV vector under the control of the long terminal repeat (LTR) of the Rous sarcoma virus (RSV). The resulting plasmid, pHPi404, contains a truncated gE-1 gene that lacks the last 430 nucleotides and codes for the first 406 amino acids of gE-1 protein.

For the construction of the 293gE-1s stable cell lines, 293 cells (embryonal kidney adherent cell line; ATCC CRL 1573) were cotransfected with pHPi404 (20 µg) and pSV2neo (2 µg) plasmid DNA. Forty-eight h post-transfection the cells were split in selection medium containing G-418 (400 µg/ml). After 10 days, individual neomycin resistant clones were isolated and tested for constitutive expression of hgE-1s protein. Cells were labelled for 2 h with 30 µCi/ml [35S]methionine (Amersham; >1000 Ci/mmol) and the proteins from cell lysates and culture medium were immunoprecipitated using either the anti-gE monoclonal (H-600) or the polyclonal (V3) antiserum. Fig. 2(b) illustrates the results of representative 293gE-1s clones. A’3-9 and A’4-15 clones, expressing the highest amount of gE-1t were chosen for further studies. B’1-7 represents a negative clone. Both positive clones expressed a protein of 54 kDa which was present in the culture medium and reacted specifically and strongly with the anti-gE antibody. As expected, the relative mobility of this protein by SDS–PAGE was higher than that predicted from the amino acid sequence, indicating that the secreted gE-1s was modified during its transport through the exocytic pathway. In addition, a second band of approximately 45 kDa was consistently present in the immunoprecipitates derived from the cell lysates of the A’3-9 and A’4-15 clones but was absent in the mock-treated cells and the non-producing clone B’1-7. This band possibly represents a cell-associated immature form of the gE-1s protein.

In order to verify and determine the extent of the predicted N-glycosylation processes in the human cell line producing gE-1s, proteins from extracts of the A’4-15 cell line were treated with EndoH and PNGase F endoglycosidases (New England BioLabs). Autoradiographs obtained after separation of the proteins on SDS–PAGE showed that the secreted form of gE was sensitive to PNGase F but resistant to EndoH digestion (Fig. 2c), indicating that this form is N-glycosylated and contains primarily complex oligosaccharides, similar to the native gE. Furthermore, we examined the relative levels of the recombinant hgE-1s produced after several passages of the cell lines. As shown in Fig. 2(d) the amount of the protein was comparable after 10 passages, indicating the stability of the episomal vector in the cells and the absence of possible toxic effects.

For vaccination experiments the secreted hgE-1s protein was derived from serum-free culture medium of confluent 293gE-1s cells after incubation at 37°C for 24 h. The hgE-1s protein was detected in considerable quantities in the supernatant (Fig. 2a). The protein content was determined by the Bradford method and was estimated to be 360 µg/4 × 10⁶ cells in 24 h. This protein was used for the vaccination experiment described below.

In order to study the protective efficacy of the E. coli and the mammalian cell produced gE-1 proteins, 2-month-old female BALB/c mice were immunized with recombinant hgE-1s, bgE-1t and hgB-1s HSV-1 glycoproteins. The hgB-1s was prepared as previously described (Manservigi et al., 1990). This protein is known to be an effective subunit vaccine in mice and was used for comparative purposes. The mice received two intraperitoneal inocula which contained 5 µg (0.5 ml/dose) of hgE-1s, bgE-1t, bgE-1t/Xa (bgE-1t-MBP cut with factor Xa), hgB-1s and hgB-1s + hgE-1s mixed with an equal volume of Freund’s complete adjuvant on day 1 or with Freund’s incomplete adjuvant on day 25. The control animals were injected intraperitoneally with culture medium of normal 293 cells or with 5 µg of MBP in the same way as the animals receiving recombinant glycoproteins. A positive control group of mice was immunized on day 1 by inoculation of a non-
Fig. 2. Analysis of the hgE-1s protein expression in the 293gE-1 clones. (a) Coomassie-blue stained 8.5% SDS-polyacrylamide gel illustrating hgE-1s produced by the A'3-9 cell line. (b) Autoradiograph of immunoprecipitates of hgE-1s protein with anti-gE monoclonal antibody (H-600), from cell lysates and the supernatant culture medium after separation on a 10% SDS–polyacrylamide gel. A'3-9 and A'4-15 represent positive human cell clones, B'1-7 corresponds to a negative human cell clone and M corresponds to 293 cells. The 54 kDa and 45 kDa protein bands indicated by the arrows represent the secreted form and the cell-associated form of the gE-1 protein. (c) Analysis of glycosylation patterns of the hgE-1s protein. Supernatants from A'3-9 cells, were immunoprecipitated with the V3 polyclonal antiserum. Subsequently, the immunoprecipitated proteins were treated with EndoH or PNGaseF endoglycosidases, resolved on a 10% SDS–polyacrylamide gel and visualized by autoradiography. Autoradiograph of the immunoprecipitated proteins from the supernatant culture medium untreated, or treated with EndoH or PNGaseF. (d) Autoradiograph of hgE-1s protein, produced by the A'3-9 stable cell line after 1, 3, 7 and 10 passages, immunoprecipitated with anti-gE monoclonal antibody (H-600). F corresponds to 293 cells infected with HSV-1 (F strain) 15 h post-infection and M corresponds to 293 cells. All the immunoprecipitation experiments were carried out after labelling of the cells with 30 µCi/ml [³⁵S]methionine for 2 h between 12 and 48 h post-seeding.

lethal dose of HSV-1 (F) into the foodpad. Three weeks after the final vaccination, mice were challenged intraperitoneally with 1.5 x 10⁶ p.f.u. (50 LD₅₀) of the virulent HSV-1 strain 13. Challenged mice were monitored for 3 weeks. To study the immunogenicity of recombinant hgB-1s and gE-1 proteins 3 weeks after the final vaccination and before challenge, mice were bled and pooled sera from each group of animals were tested for HSV-1 neutralizing activity in an in vitro neutralization assay. Sera were heat inactivated at 56 °C for 30 min. Duplicate twofold serial dilutions were mixed with 100 p.f.u. of HSV-1 (F) in MEM containing 5% FCS and 10% guinea-pig complement. Virus/serum mixtures were incubated for 1 h at 37 °C and then inoculated onto
Table 1. Protection of mice against lethal HSV-1 challenge by recombinant gE-1 and gB-1s

<table>
<thead>
<tr>
<th>Antigen</th>
<th>In vitro neutralization titre</th>
<th>No. dead/no. inoculated</th>
<th>Protection (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>hgE-1s</td>
<td>&gt; 1280</td>
<td>0/12</td>
<td>100</td>
</tr>
<tr>
<td>hgB-1s</td>
<td>&gt; 1280</td>
<td>0/12</td>
<td>100</td>
</tr>
<tr>
<td>hgE-1s + hgB-1s</td>
<td>&gt; 1280</td>
<td>0/12</td>
<td>100</td>
</tr>
<tr>
<td>bgE-1t</td>
<td>&gt; 1280</td>
<td>7/12</td>
<td>45</td>
</tr>
<tr>
<td>bgE-1t/Xa</td>
<td>&lt; 20</td>
<td>12/12</td>
<td>0</td>
</tr>
<tr>
<td>HSV-1 (F)</td>
<td>&gt; 1280</td>
<td>0/12</td>
<td>100</td>
</tr>
<tr>
<td>293 cell CM</td>
<td>&lt; 20</td>
<td>12/12</td>
<td>0</td>
</tr>
<tr>
<td>MBP</td>
<td>&lt; 20</td>
<td>12/12</td>
<td>0</td>
</tr>
</tbody>
</table>

Vero cell monolayers grown in 24-well microtitre plates. The virus was adsorbed for 1 h at 37 °C and then overlaid with MEM containing 1% FCS and 0.1% γ-globulin. The plates were incubated at 37 °C for 48 h and stained with Giemsa. The plaques were counted and the antibody titres (50% plaque reduction) were expressed as the reciprocal of the serum dilution.

Animals immunized with hgB-1s or with a combination of hgB-1s and hgE-1s achieved complete protection against a lethal challenge with HSV-1 strain 13 administered intraperitoneally and the level of protection was as good as that induced by vaccination with a non-lethal dose of HSV-1. Protection was almost as good when mice were immunized with hgE-1t alone (only 1 of 12 mice died, 92% protection). In contrast, immunization with bgE-1t either fused with MBP or separated by digestion with protease factor Xa resulted in a lower survival rate compared with that following immunization with hgE-1s. Nevertheless, the survival rate of the bgE-1t-vaccinated mice was higher (45%) than that of the mock-vaccinated controls (0%) (Table 1). Thus, mice vaccinated with the E. coli expressed gE-1t recombinant protein were partially protected from HSV-1 challenge. The neutralizing antibody titres in sera of mice immunized either with hgB-1s or with a combination of hgB-1s and hgE-1s were greater than 1:1280 and comparable to antibody titres of mice immunized with infectious virus (Table 1). Sera pooled from mice vaccinated with hgE-1s also had considerable in vitro neutralizing activity (1:970). On the other hand, the neutralizing antibody response of the mice immunized with bgE-1t was about eightfold lower than the response of the hgE-1s vaccinated mice.

In conclusion, mice vaccinated with the human cell-produced gE (hgE-1s) developed high titres of complement-mediated neutralizing antibodies and were highly protected against lethal infection with HSV, whereas immunization with E. coli produced gE (bgE-1t) gave low titres of complement-mediated neutralizing antibodies and low levels of protection. Thus, there appears to be a strong correlation between protective efficacy and titres of neutralizing antibodies. However, it should be noted that his does not apply for all of the HSV recombinant glycoproteins, since recent studies concerning the baculovirus-expressed gG (Ghiasi et al., 1994a) and gK (Ghiasi et al., 1994b) have shown that these proteins can induce protective immunity even though they elicit low levels of neutralizing antibodies. Treatment of the bgE-1t with the factor Xa, which cleaves away the MBP portion of the fusion protein, does not appear to affect the results suggesting that the low level of protective immunity is independent of the presence of the bacterial MBP. Thus, the above findings suggest the presence of immunogenic domains in the amino acid sequence of the extracellular portion of gE-1, capable of providing the observed 'partial' protection.

Vaccination studies using recombinant HSV glycoproteins other than gB, gD and gC are rather limited, consisting mainly of the use of baculovirus recombinants and live vectors such as vaccinia virus (Martin et al., 1989; Ghiasi et al., 1994a). Most importantly, some of the conclusions are rather variable. For example, vaccination studies based on vaccinia virus recombinants expressing HSV glycoproteins showed significant protection only with gB, gC and gD, partial protection with gE and no protection with gG, gH and gI (Martin et al., 1989; Blacklaws et al., 1990). In contrast, baculovirus-expressed antigens appear to be highly immunogenic even though several studies have reported that oligosaccharide structures synthesized by insect cells differ from those synthesized by mammalian cells (Lehman et al., 1993). Immunization with baculovirus recombinants showed significant protection with gB, gC, gD, gE or gI, partial protection with gG and gK and no protection with gH (Ghiasi et al., 1994a, b). These results suggested that other glycoproteins besides the well-studied gB, gC and gD may play important roles in the induction of protective immunity when presented by means other than the use of vaccinia virus vectors. However, the interpretation of these results should be qualified by the understanding that the vaccination experiments with the baculovirus-expressed HSV glycoproteins were not performed with purified recombinant proteins but rather with lysates of whole insect cells infected with baculovirus recombinants followed by a low dose of challenge virus (4 LD₅₀). Moreover, the animals vaccinated with cell lysates of insect cells (mock-vaccinated) exhibited unexpectedly high survival rates, ranging between 20 to 40%, which might be due to non-specific immunological response or the low challenge dose of the virus. Since the composition of a subunit vaccine is known to have a strong impact on the immunogenic properties of the antigen and its relative efficacy is dose dependent (Burke, 1993), studies with purified glycoproteins would be
rather important in determining the protective efficacy of the individual protein. Our results address these problems and demonstrate that the protection conferred by vaccination with hgE-1s protein is specific inasmuch as the formulation of the vaccine is well defined and the high dose of challenge virus (50 LD_{50}) does not permit the survival of control animals (0% protection).

In conclusion, the expression of the gE protein in human cells has enabled us to re-assess the relative importance of this subunit as a target of the immune system and has confirmed that the gE is capable of inducing high titres of virus-neutralizing antibodies and good levels of protection against lethal intraperitoneal HSV challenge infection in animals. Therefore, gE is an important inducer of protective immunity to HSV infection and might thus be a useful and important component of a (multi-)subunit vaccine.

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