Therapeutic effects of bovine enterovirus infection on rabbits with experimentally induced adult T cell leukaemia

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A bovine enterovirus, MZ-468, showed cytopathic effects on cell line F-647a, which was established by coculture of human T cell lymphotropic virus type 1-transformed MT-2 cells and X-irradiated rabbit lymphocytes. Microcalorimetric assay showed that residual, viable, MZ-468-infected F-647a cells produced less heat than non-infected cells. The therapeutic effects of MZ-468 infection were examined in rabbits in which adult T cell-like leukaemia (ATL) had been induced by inoculation of F-647a cells (1 x 10⁸ cells). Six newborn rabbits were separated into three groups: group A was inoculated with F-647a cells only; group B was treated with the same amount of virus 24 h after the inoculation with cells and then once every 4 days. Both of the animals in group A and one in group C died 10 and 11 days, and 22 days, respectively, after the inoculation with cells. Both rabbits in group B and one in group C survived for more than 4 months. The rabbits that died were examined pathologically; leukaemic infiltrations were found in the lungs of the group B rabbits, and in the lungs, spleens and livers of both group A rabbits. Two identical experiments produced almost the same findings. These results suggest that bovine enterovirus might be used clinically to prolong the life-span of ATL patients.

Anticancer therapy in humans is one of the most difficult medical problems; a great number of chemical and biological agents have been used for this purpose, none of which is completely effective. The problems involved include the non-specificity of the action of the anticancer agents, which is accompanied by severe side-effects. The use of viruses for therapy of malignant tumours is one of the strategies which employ biologically active agents (Casto & Hammon, 1970; Hunter-Craig et al., 1970; Taylor et al., 1971). Our laboratory has tried to identify those viruses which show strong oncolytic effects and minimal side-effects (Shingu, 1976, 1981; Kaneko et al., 1979), and we have found that bovine enteroviruses lytically infect mouse fibroblast cells only when the cells are transformed with simian virus 40 (Shingu, 1976). We have also found that some bovine enteroviruses have an oncolytic effect on solid tumours and prolong the life-span of mice bearing human carcinoma cells (Fujimaru, 1978).

Seto et al. (1988) recently have succeeded in inducing a disease closely resembling an acute type of adult T cell-like leukaemia (ATL) in rabbits by inoculating syngeneic human T cell lymphotropic virus type 1 (HTLV-1)-transformed cells into newborn rabbits. The HTLV-1-transformed T cell line, F-647a, was established from a 2-month-old female (B/J x Chbb:HM) F1 rabbit, which had been infected with the virus neonatally. In brief, peripheral blood lymphocytes were cultured in RPMI 1640 medium supplemented with 10% foetal calf serum (FCS), in the absence of T cell growth factor. The established cell line had the karyotype of a normal female rabbit, and was positive for the HTLV-1 p19 antigen, the pan T cell marker antigen and the Ia antigen, which were detectable using monoclonal antibodies. The F-647a cell line was able to kill newborn rabbits within 7 days when a dose of 1 x 10⁸ cells was inoculated intraperitoneally. In this study we demonstrated that bovine enterovirus has effects against a rabbit model of ATL, a disease caused by HTLV-1 in which patients usually die within about 4 months.

A bovine enterovirus of type 1 subtype 1, MZ-468, was isolated from calf faeces (Tamada, 1977; Urakawa & Shingu, 1987). The virus was propagated in HeLa cells, purified by equilibrium centrifugation in a CsCl gradient and plaque assayed in Wilms tumour cells originally established in this laboratory (Shingu & Chibana, 1981). F-647a cells (Seto et al., 1988) were kindly provided by Professor A Seto, Siga University School of Medicine, Japan, and passaged in RPMI 1640 medium (Nissui) containing 10% FCS (Gibco).

MZ-468 (5.0 p.f.u./cell) was inoculated and adsorbed onto suspended F-647a cells (4 x 10⁴) for 1 h at 37 °C in plastic dishes (Falcon), and incubated at 37 °C under 5% CO₂. Virus propagation was examined every 6 to 12 h by
plaque assay in monolayers of Wilms tumour cells. The average virus titre was determined from four or five cultures and MZ-468 replication in F-647a cells was confirmed using a one-step growth curve (Fig. 1).

To determine the oncolytic activity of MZ-468 against F-647a cells in tissue culture, two different techniques were chosen to measure cell activity: measurement of heat production, and the number of cells present after 1, 2 and 3 days of cultivation.

A flow-type microcalorimeter ( thermoactive cell analyser ESCO-3000, Denshi Kagaku) was used to measure heat production. Washing medium (1.8 ml), guide medium (1.8 ml) and sample medium (0.6 ml) containing 2 × 10^6 suspended cells were added, in that order, to the microcalorimeter. Cell suspension (0.6 ml) was transferred into the microcalorimetric tube which was then introduced into the reaction vessel and heat production was recorded over a period of 15 min. The same medium used for cell culture was used for the measurement of heat production and oxygen consumption. The heat produced by MZ-468-infected (m.o.i. 13) or uninfected F-647a cells was measured after 24, 48 and 72 h of culture at 37 °C. Heat production, i.e. thermal power, was expressed as μW/10^6 cells, and the result presented as a percentage of the rate of heat production.

Cells were counted in a Bürker chamber. Cell viability was checked by the trypan blue exclusion test during culture and before microcalorimetry.

Infection of F-647a cells in RPMI 1640 by MZ-468 reduced the viability of and production of heat by the residual viable cells, as shown in Table 1. A good correlation existed between the number of cells and the results of the microcalorimetry experiments, indicating that bovine enterovirus can kill F-647a cells.

Six newborn New Zealand white rabbits (3 days old) from one litter were separated into three groups which were treated as shown schematically in Fig. 2. Group A was injected intraperitoneally with cells only (1 × 10^8); group B was injected with the same number of cells and 2.8 × 10^7 p.f.u. MZ-468; group C was injected with the same number of cells, and 24 h later virus (2.8 × 10^7 p.f.u.) was injected intraperitoneally and this was repeated once every 4 days. All the rabbits were allowed to suckle milk from their mother in a special, separate box.

Results of this experiment are shown schematically in Fig. 2. The group A animals (receiving no treatment with MZ-468) died 10 and 11 days after inoculation with cells. Pathologically, leukaemic infiltrations were found in the lungs, spleens and livers. In contrast, both group B

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Table 1. Effect of virus infection on viability of and heat production by viable F-647a cells

<table>
<thead>
<tr>
<th>Time post-infection (h)</th>
<th>Native F-647a</th>
<th>Virus-infected F-647a</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Viable cells (×10^6)</td>
<td>µW (%)</td>
</tr>
<tr>
<td>0</td>
<td>4.3</td>
<td>100</td>
</tr>
<tr>
<td>24</td>
<td>7.2</td>
<td>100</td>
</tr>
<tr>
<td>48</td>
<td>7.1</td>
<td>100</td>
</tr>
<tr>
<td>72</td>
<td>16.5</td>
<td>100</td>
</tr>
</tbody>
</table>

* Figures in parenthesis represent the number of viable cells expressed as a percentage of the number of viable, uninfected cells.
animals (which were simultaneously treated with MZ-468 and the ATL-derived cells) survived for more than 4 months. One of the animals from group C (which was treated 1 day after inoculation with cells) died showing leukaemic infiltration, but the other rabbit survived for longer than 4 months. Two other similar experiments produced almost the same results and pathological findings (data shown in Fig. 3). In this study, bovine enterovirus is effective in the treatment of ATL in experimentally treated animals.

The host range of a virus is generally restricted as to species and this restriction is dependent on the existence of cellular receptors. Bovine enteroviruses do not infect primary cultured human cells, but do infect malignant human cell lines (Shingu, 1976). A possible mechanism for this change in sensitivity may be that the cell membrane changes molecularly during transformation, a phenomenon derived from derepression and amplification of proto-oncogenes, as found for c-erbB-2 (Semba et al., 1985). Thus, expression of cell proto-oncogenes, especially those of the receptor type may produce targets for the adsorption of virus. Another mechanism for a viral oncolytic effect involves unmasking or xenozenzina-

References


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