**Mycobacterium saopaulense** sp. nov., a rapidly growing mycobacterium closely related to members of the *Mycobacterium chelonae–Mycobacterium abscessus* group

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Five isolates of non-pigmented, rapidly growing mycobacteria were isolated from three patients and, in an earlier study, from zebrafish. Phenotypic and molecular tests confirmed that these isolates belong to the *Mycobacterium chelonae–Mycobacterium abscessus* group, but they could not be confidently assigned to any known species of this group. Phenotypic analysis and biochemical tests were not helpful for distinguishing these isolates from other members of the *M. chelonae–M. abscessus* group. The isolates presented higher drug resistance in comparison with other members of the group, showing susceptibility only to clarithromycin. The five isolates showed a unique PCR restriction analysis pattern of the *hsp65* gene, 100 % similarity in 16S rRNA gene and *hsp65* sequences and 1–2 nt differences in *rpoB* and internal transcribed spacer (ITS) sequences. Phylogenetic analysis of a concatenated dataset including 16S rRNA gene, *hsp65*, and *rpoB* sequences from type strains of more closely related species placed the five isolates together, as a distinct lineage from previously described species, suggesting a sister relationship to a group consisting of *M. chelonae*, *Mycobacterium salmoniphilum*, *Mycobacterium franklinii* and *Mycobacterium immunogenum*. DNA–DNA hybridization values >70 % confirmed that the five isolates belong to the same species, while values <70 % between one of the isolates and the type strains of *M. chelonae* and *M. abscessus* confirmed that the isolates belong to a distinct species. The polyphasic characterization of these isolates, supported by DNA–DNA hybridization results, demonstrated that they share characteristics with *M. chelonae–M. abscessus* members, but constitute a different species, for which the name *Mycobacterium saopaulense* sp. nov. is proposed. The type strain is EPM 10906T (=CCUG 66554T=LMG 28586T=INQCS 0733T).

**Abbreviations:** DDH, DNA–DNA hybridization; ITS, internal transcribed spacer; PFGE, pulsed-field gel electrophoresis; PRA, PCR restriction analyses.

The GenBank/EMBL/DDBJ accession numbers for the 16S rRNA gene sequence of strains EPM 10906T, EPM 10695, IAL 3785, JAN1 and JAN2 are KM973037, KM973036, KM973038, DQ866774 and DQ866775, respectively; for the *hsp65* sequence are KM973026, KM973025, KM973027, DQ866786 and DQ866787, respectively; for the *rpoB* sequence are KM973029, KM973028, KM973030, KM973031 and KM973032, respectively; and for the internal transcribed spacer (ITS) sequence are KM973034, KM973033, KM973035, DQ866774 and DQ866775, respectively.

Three supplementary tables and a supplementary figure are available with the online Supplementary Material.
Nontuberculous mycobacteria are ubiquitous environmental organisms and several species can cause opportunistic infections in humans, in particular the members of the Mycobacterium chelonae–Mycobacterium abscessus group. This group comprises closely related, rapidly growing mycobacteria that can cause a broad spectrum of infections mainly affecting lung, skin and soft tissue (Simmon et al., 2011; Wallace et al., 1983). The ubiquitous distribution of these organisms facilitates the contamination of medical equipment and solutions that, associated with the growing number of therapeutic interventions, generate nosocomial infections and outbreaks representing a serious public health problem in some settings (Leão et al., 2010; Tortoli, 2009).

Until recently, the M. chelonae–M. abscessus group was composed of M. chelonae, M. abscessus (Kusunoki & Ezaki, 1992), Mycobacterium immunogenum (Wilson et al., 2001), Mycobacterium massiliense (Adékambi et al., 2004, 2006b), Mycobacterium bolletii (Adékambi et al., 2006a) and Mycobacterium salmoniphilum (Whipps et al., 2007). Taxonomic changes have been proposed and M. abscessus, M. massiliense and M. bolletii have been assigned to a single species (M. abscessus) with two subspecies, M. abscessus subsp. abscessus and M. abscessus subsp. bolletii, the latter including those isolates previously identified as M. massiliense and M. bolletii (Leão et al., 2011; Leão et al., 2009). Two novel members of this group have been recently described, Mycobacterium franklinii (Nogueira et al., 2015; Simmon et al., 2011) and ‘Mycobacterium fukienense’ (Zhang et al., 2013).

The aim of the present study was to define the taxonomic position of five mycobacterial isolates without conclusive species assignments (isolates EPM 10906T, EPM 10695, IAL 3785, JAN1 and JAN2), which share a PCR restriction analyses (PRA) profile of the hsp65 gene not present in the PRASITE database (http://app.chuv.ch/prasite/index.html). Our data indicate that these five isolates belong to a single taxon and represent a novel species of the M. chelonae–M. abscessus group.

The first two isolates (EPM 10906T and EPM 10695) were obtained in 1999 from corneal specimens of two patients with infectious crystalline keratopathy after LASIK surgery (laser-assisted in situ keratomileusis) performed in the same ophthalmological clinic, in São Paulo city (Brazil). These isolates were first misidentified as M. chelonae (Alvarenga et al., 2002) and subsequently as M. abscessus (Sampaio et al., 2006) on the basis of PRA of the hsp65 gene. Typing of these isolates by pulsed-field gel electrophoresis (PFGE) using a previously described protocol (Matsumoto et al., 2011), revealed that they share indistinguishable patterns and thus might belong to a single strain (Fig. 1). The third isolate (IAL 3785) was obtained in 2007 from a cervical abscess in the city of Ribeirão Preto, São Paulo (Brazil). The other two isolates (JAN1 and JAN2) were isolated from zebrafish (Danio rerio) (Kent et al., 2004) and initially categorized as M. chelonae. Using greater taxon sampling in a later phylogenetic analysis, these strains were recognized as phylogenetically distinct (Whipps et al., 2007), but they were not described as a novel species at the time. Isolates JAN1 and JAN2 share highly similar PFGE patterns, differing only in a single band, and were isolated from zebrafish at the same research facility about two months apart in 2003 (Kent et al., 2004); considering this, they could represent a single strain.

We investigated the classification of these five isolates, comprising three different strains, using a polyphasic approach that included microscopic and macroscopic morphological examination, cultural and biochemical tests, drug susceptibility testing, HPLC analysis of cell-wall mycolic acids, PRA, sequencing of three housekeeping genes and DNA–DNA hybridization. The results were compared with those displayed by M. abscessus subsp. abscessus ATCC 19977T, M. abscessus subsp. bolletii CCUG 50184T, M. chelonae ATCC 35752T, M. immunogenenum ATCC 700505T, M. salmoniphilum ATCC 13758T and M. franklinii DSM 45524T.

Cultures were grown on solid media [Löwenstein-Jensen (LJ), Middlebrook 7H10 supplemented with OADC (oleic acid, albumin, glucose and catalase) and Luria–Bertani agar], and in liquid Muller-Hinton medium or Luria–Bertani broth with 1 % Tween 80 at 28–30 °C for 5 days. Microscopic examination of colony smears by Ziehl–Neelsen staining confirmed that the isolates were acid-fast bacilli. Analysis of pigment production, single-source carbon utilization (mannitol, inositol and citrate), growth at 26 °C and 37 °C, and tolerance to 5 % NaCl, 0.2 % picric acid, para-nitrobenzolic acid (PNB) and nitrite were performed on 7H10-OADC and LJ. Nitrate reduction, Tween 80 hydrolysis and arylsulfatase production were also examined. All tests were performed as described in standard protocols for the biochemical identification of mycobacteria (Kent & Kubica, 1985; Leão et al., 2004; Tsukamura, 1984). The five isolates exhibited indistinguishable phenotypic and biochemical characteristics, which are listed in Table S1 (available in the online

![Fig. 1. Pulsed-field gel electrophoresis (PFGE) patterns of Dral digested DNA of the five isolates studied in this work. PFGE images were analysed with the BioNumerics program v. 7.1 (Applied Maths). The band-based Dice unweighted-pair group method using average linkages was used to prepare a dendrogram of PFGE profiles, based on 1.5 % optimization and position tolerance.](image-url)
Supplementary Material). These cultural and biochemical tests were not helpful for distinguishing these isolates from other members of the M. chelonae–M. abscessus group (Table S1).

Antimicrobial drug-susceptibility testing was performed using a microdilution method in cation-supplemented Mueller–Hinton broth, according to the recommendations of the Clinical and Laboratory Standards Institute (CLSI, 2011) for rapidly growing mycobacteria. Amikacin, cefoxitin, ciprofloxacin, clarithromycin, doxycycline, minocycline, moxifloxacin and tobramycin were tested. The five isolates were more drug resistant than the other members of the M. chelonae–M. abscessus group, showing susceptibility only to clarithromycin at 3 and 14 days incubation. The isolates were resistant to doxycycline, cefoxitin and tobramycin, and resistant or intermediate to amikacin, ciprofloxacin, minocycline and moxifloxacin (Table 1). These results are consistent with susceptibility testing previously conducted for strain JAN1 (Chang & Whippis, 2015). The drug resistance profile of the novel species described in this study highlights the importance of its correct identification for patient management.

For HPLC analysis of cell-wall mycolic acids, two strains of the panel characterized here were selected (the proposed type strain EPM 10906T and JAN1) and three reference strains belonging to the closely related M. chelonae–M. abscessus group (M. abscessus subsp. abscessus ATCC 19977T, Mycobacterium massiliense CCUG 48898T and M. abscessus subsp. bolletti CCUG 50184T). The cells of these strains, grown in culture on Middlebrook 7H10 agar, were saponified, extracted and derivatized as recommended by the Sherlock Mycobacteria Identification System (SMIS; MIDI) and separated using a gradient of methanol and 2-propanol. All the strains analysed produced nearly identical HPLC patterns characterized by two late emerging clusters of peaks (Fig. 2). The Sherlock software (version Myco 1.0) identified all the strains as M. chelonae–M. abscessus with very high similarity indexes.

Table 1. Antimicrobial susceptibility results for isolates and type strains included in this study

<table>
<thead>
<tr>
<th>Drug</th>
<th>MIC (µg ml⁻¹)</th>
<th>isolate EPM 10906T</th>
<th>isolate JAN1</th>
<th>isolate M. abscessus subsp. abscessus ATCC 19977T</th>
<th>isolate M. massiliense CCUG 48898T</th>
<th>isolate M. abscessus subsp. bolletti CCUG 50184T</th>
</tr>
</thead>
<tbody>
<tr>
<td>Amikacin</td>
<td>32 I</td>
<td>128 R</td>
<td>32 I</td>
<td>≤0.5 S</td>
<td>≤0.5 S</td>
<td>≤0.5 S</td>
</tr>
<tr>
<td>Ciprofloxacin</td>
<td>2 I</td>
<td>2 I</td>
<td>2 I</td>
<td>≤0.5 S</td>
<td>≤0.5 S</td>
<td>≤0.5 S</td>
</tr>
<tr>
<td>Clarithromycin</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3 days</td>
<td>≤0.5 S</td>
<td>≤0.5 S</td>
<td>≤0.5 S</td>
<td>≤0.5 S</td>
<td>≤0.5 S</td>
<td>≤0.5 S</td>
</tr>
<tr>
<td>Doxycycline</td>
<td>&gt;32 R</td>
<td>&gt;32 R</td>
<td>&gt;32 R</td>
<td>&gt;32 R</td>
<td>&gt;32 R</td>
<td>&gt;32 R</td>
</tr>
<tr>
<td>Cefoxitin</td>
<td>512 R</td>
<td>&gt;512 R</td>
<td>&gt;512 R</td>
<td>&gt;512 R</td>
<td>&gt;512 R</td>
<td>&gt;512 R</td>
</tr>
<tr>
<td>Tobramycin</td>
<td>16 R</td>
<td>32 R</td>
<td>32 R</td>
<td>32 R</td>
<td>32 R</td>
<td>32 R</td>
</tr>
<tr>
<td>Minocycline</td>
<td>32 R</td>
<td>16 R</td>
<td>16 R</td>
<td>16 R</td>
<td>16 R</td>
<td>16 R</td>
</tr>
<tr>
<td>Moxifloxacin</td>
<td>2 I</td>
<td>8 R</td>
<td>4 R</td>
<td>8 R</td>
<td>8 R</td>
<td>8 R</td>
</tr>
</tbody>
</table>

Fig. 2. Representative mycolic acid pattern of isolate EPM 10906T paired with the reference profile of Mycobacterium chelonae–Mycobacterium abscessus (Sherlock database). LMMIS, Low molecular mass internal standard; HMMIS, high molecular mass internal standard.
For molecular identification, PRA of the hsp65 gene and of the ITS, and partial sequencing of the small subunit (16S) rRNA gene, rpoB, hsp65 and ITS were performed. The DNA was prepared by boiling one loop full of bacteria for 10 min in 300 μl TET (10 mM Tris, 1 mM EDTA, 1 % Triton X-100; pH 8.0) followed by centrifugation at 14 000 g for 2 min. For PRA-hsp65, a 441 bp fragment of the hsp65 gene was amplified using primers Tb1 and Tb12 (Table S2), and the amplicon was digested in two separate tubes with BstEII and HaeIII restriction enzymes (Telenti et al., 1993). For PRA-ITS, amplicons generated with primers Sp1 and Sp2 (Table S2) were digested with TaqI restriction enzyme (Roth et al., 2000). PRA-hsp65 and PRA-ITS digestion products were visualized in 3 % agarose gels stained with ethidium bromide after electrophoresis, using the 50 bp ladder as the molecular size standard. The restriction fragment sizes were estimated using the BioNumerics program version 7.1 (Applied Maths) and compared to the patterns included in the PRASITE database for PRA-hsp65 and published by Roth et al. (2000) for PRA-ITS. The five isolates showed identical PRA-hsp65 patterns – BstEII [bp] (235, 210) and HaeIII [bp] (145, 60, 50), which differs from patterns of other members of the M. chelonae–M. abscessus group. This profile was not registered in the PRASITE database. The isolates showed the same PRA-ITS pattern – TaqI [bp] (225, 30), which is common to M. abscessus and M. franklinii. Therefore, only PRA-hsp65 was useful for differentiation of this novel species.

For DNA–DNA hybridization (DDH), high-molecular-mass DNA was prepared from 2 g cell mass using the protocol described by Pitcher et al. (1989) with modifications. Bacterial cells were centrifuged, inactivated at 90 °C for 30 min and resuspended in 3 ml lysis buffer containing 1 ml⁻¹: 200 μg RNase, 25 mg fresh lysozyme and 100 U mutanolysine. The suspensions were incubated overnight at 37 °C and the DNA was extracted with chloroform/isooamyl alcohol, treated with RNase and precipitated with ethanol as described by Marmur (1961). DDH was performed as described by Ezaki et al. (1989), using
photobiotin-labelled probes in microplate wells. Fluorescence was measured in a HTS7000 Bio Assay Reader (Perkin-Elmer). DNA–DNA hybridization values are presented as means of reciprocal experiments, performed in quadruplicate hybridization reactions. The DNA G+C content, estimated as described by Mesbah & Whitman (1989), was 64.6 mol% (EPM 10906T), 64.8 mol% (EPM 10695), 64.7 mol% (IAL 3785), 64.5 mol% (JAN1) and 64.7 mol% (JAN2). These values were used for calculation of the 50 °C hybridization temperature, and are consistent with the DNA G+C contents of the genus Mycobacterium, between 59 mol% and 66 mol% (Devulder et al., 2005). DDH experiments performed with the five isolates yielded hybridization values above 70% (data not shown), confirming that they belong to the same species. Isolate EPM 10906T was selected to perform DDH reciprocal experiments with the type strains of the M. chelonae–M. abscessus group. All values were below 70%, confirming that EPM 10906T and the other four isolates belong to a distinct species of the M. chelonae–M. abscessus group (Table 2).

In conclusion, phenotypic and genotypic tests indicated that isolates EPM 10906T, EPM 10695, IAL 3785, JAN1 and JAN2 belong to the M. chelonae–M. abscessus group. In addition, several results clearly indicated that these isolates form a uniform group separated from the other members of

Table 2. DNA–DNA hybridization values of isolate EPM 10906T against the type strains of members of the M. chelonae–M. abscessus group

<table>
<thead>
<tr>
<th>Strain</th>
<th>Mean DNA–DNA relatedness ± SD (%)</th>
<th>Reciprocal values</th>
</tr>
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<tbody>
<tr>
<td>M. abscessus subsp. abscessus ATCC 19977T</td>
<td>41.5 ± 2.5</td>
<td>39;44</td>
</tr>
<tr>
<td>M. abscessus subsp. bolletii CCUG 50184T</td>
<td>40 ± 1</td>
<td>39;41</td>
</tr>
<tr>
<td>M. chelonae ATCC 35752T</td>
<td>47 ± 5</td>
<td>42;52</td>
</tr>
<tr>
<td>M. immunogenum ATCC 700505T</td>
<td>40 ± 7</td>
<td>33;47</td>
</tr>
<tr>
<td>M. salmoniphilum ATCC 13758T</td>
<td>44.5 ± 1.5</td>
<td>43;46</td>
</tr>
<tr>
<td>M. franklinii DSM 45524T</td>
<td>51 ± 4</td>
<td>47;55</td>
</tr>
</tbody>
</table>
M. chelonae–M. abscessus group. Therefore, we propose to classify these isolates as a novel species of the genus Mycobacterium in the M. chelonae–M. abscessus group, with the name Mycobacterium saopaulense sp. nov.

Description of Mycobacterium saopaulense sp. nov.

Mycobacterium saopaulense (sa.o.paul.en’se. N.L. neut. adj. saopaulense of or pertaining to the Brazilian state of São Paulo, where the first strains were isolated).

Cells are acid-fast bacilli and visible growth on solid media requires 3–5 days at 28 °C. Colonies are non-pigmented and smooth. After some days, the medium often acquires a brown colour. Growth occurs in the presence of 5% NaCl, picric acid, para-nitrobenzoic acid (PNB) and nitrite. Growth is observed in the presence of citrate as a single source of carbon, but not in the presence of mannitol or inositol. Negative reactions are observed for nitrate reduction and Tween 80 hydrolysis. Conventional biochemical testing cannot distinguish this species from other members of the M. chelonae–M. abscessus group. The antimicrobial pattern is characterized by susceptibility to clarithromycin and resistance to doxycycline, tobramycin and cefoxitin. Variable results, intermediate or resistant, were obtained with amikacin, ciprofloxacin, minocycline and moxifloxacin. The mycolic acid profile is similar to that of M. chelonae–M. abscessus by HPLC analysis. Genotype CM shares the same profile as M. chelonae. The partial sequences of rpoB and hsp65 genes can distinguish Mycobacterium saopaulense from other members of M. chelonae–M. abscessus group. The PRA-hsp65 pattern that characterizes this species is BstEII [bp] (235, 210) and HaeIII [bp] (145, 60, 50).

The type strain is EPM 10906T (=CCUG 66554T=LMG 28586T=INCQS 0733T). The DNA G+C content of the type strain is 64.6 mol%.

Acknowledgements

This study received financial support from Fundação de Amparo à Pesquisa do Estado de São Paulo (www.fapesp.br) (FAPESP) (grant 2011/18326-4). C. L. N. and C. K. M. received fellowships from FAPESP (2012/13763-0 and 2013/16018-6). This work has been partially supported by International Cooperation UAM-Banco Santander and Latin America (CEAL-UAM). Contributions to this work by C. M. W. were funded in part by the Office of Research Infrastructure Programs of the National Institutes of Health (NIH) under award number R24OD010998. The content is solely the responsibility of the authors and does not necessarily represent the official views of the NIH.

References


