Legionella steelei sp. nov., isolated from human respiratory specimens in California, USA, and South Australia

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Legionella-like bacteria were isolated from the respiratory tract of two patients in California, USA, and South Australia, but were not thought to cause disease. These bacteria, strains F2632 and IMVS-3376T, were found to have identical Legionella macrophage infectivity potentiator (mip) gene sequences and were therefore further characterized to determine their genetic and phenotypic relatedness and properties. Both of these Gram-negative-staining bacterial strains grew on buffered charcoal yeast extract medium, were cysteine auxotrophs and made a characteristic diffusible bright yellow fluorescent pigment, with one strain making a late appearing colony-bound blue–white fluorescent pigment. The optimal in vitro growth temperature was 35 °C, with very poor growth at 37 °C in broth or on solid media. There was no growth in human A549 cells at either 35 or 37 °C, but excellent growth in Acanthamoeba castellani at 30 °C and poorer growth at 35 °C. Phylogenetic analysis of these bacteria was performed by sequence analysis of 16S rRNA, mip, ribonuclease P, ribosomal polymerase B and zinc metalloprotease genes. These studies confirmed that the new strains represented a single novel species of the genus Legionella for which the name Legionella steelei sp. nov. is proposed. The type strain is IMVS-3376T (=IMVS 3113T=ATCC BAA-2169).

Members of the genus Legionella are Gram-negative bacteria that can cause a type of pneumonia termed Legionnaires’ disease and rarely non-pneumonic infectious diseases (Edelstein & Cianciotto, 2010). Legionella pneumophila is the primary cause of Legionnaires’ disease and 19 other species of the genus Legionella have been shown to cause the disease, mostly in immunocompromised patients. At the time of writing, there were 53 members of the genus Legionella with validly published names, 33 of which have been isolated only from environmental sources (http://www.bacterio.cict.fr/index.html). All of the recognized species of the genus Legionella that have been examined for intracellular growth have been found to be facultative or obligate intracellular parasites of free living amoebae, and those causing human disease are facultative intracellular parasites of human cells (Cianciotto et al., 2006).

Identification of members of the genus Legionella was originally based on DNA–DNA hybridization, supplemented by serological identification, cellular fatty acid profiles and ubiquinone synthesis and other phenotypic tests (Brenner, 1987). However, serological cross-reactivity and the non-availability of antibodies to many recently described species makes serotyping a difficult and frequently a non-specific method of species identification. DNA–DNA hybridization is performed at only a few laboratories worldwide. Finally, cellular fatty acid and ubiquinone analyses are rarely performed for species identification, as typical patterns are shared for some species and are unavailable for several recently described Legionella species. Genomic DNA sequence analysis based on multiple gene targets has become the standard for the identification of members of the genus Legionella because of its portability, specificity and ease of performance (Lück et al., 2010).
In this study, the discovery of a novel species of the genus *Legionella* is described. The novel species was found following an examination of *Legionella*-like bacteria from the respiratory tracts of two patients from different continents who were thought not to have Legionnaires' disease. Independent identification of these clinical isolates by macrophage infectivity potentiator (*mip*) gene sequencing led to further testing to determine whether these two strains represented a single novel species.

**Methods**

**Phenotypic testing.** Bacteria were grown on laboratory-made MOPS buffered charcoal yeast extract medium supplemented with x-ketoglutarate (BCYEz agar), ACES buffered yeast extract broth (BYEx broth) and ACES buffered charcoal yeast extract broth (BCYEz broth), and incubated in humidified incubators with or without 5% CO₂ for plates, or in shaking incubators for broth cultures (Edelstein & Edelstein, 1991, 1993). Tests for growth on selective media used commercially prepared BCYEz media with added polymyxin B, anisomycin and cefamandole (BBL PAC; BD Diagnostics), or with added polymyxin B, anisomycin and vancomycin (BBL PAV). To detect flagella, bacteria were grown on a BCYEz plate at 35°C, then harvested into sterile water. The suspended bacteria were kept at room temperature for two to five days and then stained for flagella using the Ryu stain (Kodaka et al., 1982). Bacterial motility in amoebae was determined by microscopic examination of intra-amoebic bacteria. Gram reaction, oxidase, gelatinase and hippurate reactions, and glucose utilization were determined as described previously (Edelstein et al., 1982). Diffusible and colony fluorescence of plate-grown bacteria was detected by illumination with a long wave UV light in the dark. Reactivity with direct fluorescent antibodies to several different *Legionella* species was tested as described previously (Edelstein, 2004). Analysis of cellular fatty acid composition by gas liquid chromatography of fatty acid methyl esters was performed by MIDI Laboratories using 24 h growth on heavily inoculated BCYEz plates incubated in humidified ambient air at 35°C (Sherlock version 6.1, MIDI Laboratories, Inc.).

The ability of the bacteria to grow in amoebae was determined by inoculating the plate-grown (35°C) bacteria into tissue culture plates containing axenic *Acanthamoeba castellani* (ATCC 30234) in autoclaved dechlorinated tap water as described previously (Boulanger & Edelstein, 1995). The bacterial-amoebal co-cultures were incubated at 30°C in ambient air for up to seven days and followed microscopically and by supernate cultures placed onto BCYEz plates incubated at 35°C. Control strains included *L. pneumophila* serogroup 1 strain F889, a human isolate that is highly virulent for amoebae, human and guinea pig macrophages and guinea pigs (Edelstein et al., 1987); and F2352, a non-virulent *L. pneumophila dotA* mutant. Growth of the bacteria in the human respiratory alveolar epithelial cell line A549 was studied in tissue culture wells incubated at 35°C as described previously (Higa & Edelstein, 2001). Intracellular growth was monitored both microscopically and by viable plate counts.

**Genotypic analysis.** Crude bacterial DNA preparations were made by boiling heavy suspensions from plate-grown bacteria in EDTA (5 mM) or in Tris/HCl (10 mM) and EDTA (1 mM) (TE) for 10 min. The boiled supernatant was used in polymerase chain reactions (PCRs) and PCR products were sequenced on both strands by standard cycle sequence methods, as previously described (Ratcliff et al., 1998). Primers and amplification conditions were as previously described for: macrophage infectivity potentiator (*mip*) (Ratcliff et al., 1998); ribonuclease P (*rnpB*) (Rubin et al., 2005); and ribosomal polymerase B (*rpoB*) (Ko et al., 2002). Small subunit RNA gene (16S rRNA) used primers Leg-16S-F (5'-GGCTCAGATTGACGTCGGC-3') and Leg-16S-R (5'-ACCACCTCCCCATGTTGACGG-3') using 40 cycles of 94°C (30 s), 55°C (1 min) and 72°C (2 min); additional sequencing primers used were Leg16S-Int-R (5'-TTTACGCCAGTAAATCCG-3'), LEG16s-Int-F (5'-CAGGCCGCAATACG-3') and Leg16s-Int2-F (5'-TGGTCAGTCGCTGTCGTG-3'); zinc metalloprotease (*proA*) used LegProA-F (5'-TGATTTTAYGCNGNAYGNTAT-94C(30s),50°C(1min)and72°C(2min)). Sequences assembly, alignment and analysis were performed using Kodon (Applied Maths) and the phylogenetic analysis (maximum-likelihood) was performed using PAUP 4.0 (Sinauer Associates). Modeltest 3.7 (Posada & Crandall, 1998) was used to assess the best-fit model (GTR + I + G) used for the analysis and relationship trees were drawn with FigTree v.1.1.2 (Andrew Rambaut, Institute of Evolutionary Biology, University of Edinburgh, UK, http://tree.bio.ed.ac.uk/software/figtree/). Bootstrap analysis (1000 replicates) of the maximum-likelihood dendrogram was performed using RAxML accessed through the web-based CIPRES (Miller et al., 2010) portal (http://www.phylo.org/portal2). SplitsTree analysis was performed using SplitsTree4 (Huson & Bryant, 2006; http://wwwplitsTree.org/).

Sequences of isolate IMVS-3376T were deposited in GenBank/EMBL/DDBJ with accession numbers HQ398202 (16S rRNA), HQ398203 (*mip*), HQ398204 (*proA*), HQ398205 (*rnpB*) and HQ398206 (*rpoB*).

**Results**

**Case histories.** Patient 1, an adult male, who was previously healthy, had suspected bronchitis in February 1999, in Adelaide, Australia. Cultures for *Legionella* were performed from sputum. A slowly growing legionella-like organism was isolated at the Microbiology and Infectious Diseases Laboratories, Institute of Medical and Veterinary Science, Adelaide, Australia. The patient was thought not
to have a legionella infection based on clinical grounds. Of note was the fact that the patient had been making and using compost for gardening prior to the onset of his bronchitis. The bacterial isolate was designated isolate number IMVS-3376T. The isolate appeared to be a novel species based on the absence of reactivity with available antisera as well as mip gene sequence analysis.

Patient 2, an adult male, had suspected pneumonia in September 2008, in Los Angeles, California, USA, while hospitalized for severe congestive heart failure. Bronchoalveolar lavage specimen at the Hospital of the University of Pennsylvania, USA, grew a legionella-like bacterium on BCYE medium at 35 °C in humidified air after four days incubation. Consultants felt that the patient was unlikely to have Legionnaires’ disease but treated the patient regardless with a variety of antimicrobials known to be active against L. pneumophila. The patient died several months later of his underlying disease.

Identification of the bacterial isolate by 16S rRNA sequencing (500 bp) revealed 96 % sequence similarity to Legionella anisa. The isolate was referred to a research laboratory at the hospital of the University of Pennsylvania, USA, for definitive identification and was given isolate number F2632. The isolate appeared to represent a novel species based on absence of reactivity with available antisera; mip gene sequencing revealed the similarity of the strain to the corresponding gene sequence of isolate IMVS-3376T.

**Phenotypic characterization.** Both strains IMVS-3376T and F2632 were characterized in one laboratory (Philadelphia, USA) to determine their phenotypic properties. Both isolates were Gram-negative-staining rods that were cystone auxotrophs. They both possessed a single monopolar flagellum and grew on BCYE medium in ambient air.

The bacterial colonies were round, 0.5–2 mm in diameter, entire and convex, with an opalescent ground glass appearance when examined using a dissecting microscope. A large amount of a diffusible yellow–green fluorescent pigment was produced by the bacteria growing on BCYE medium, such that the entire plate appeared yellow when illuminated by long wave UV light. In addition, individual colonies of strain IMVS-3376T, but not strain F2632, were brightly electric blue–white fluorescent; this fluorescence was not diffusible into the surrounding medium and appeared to be maximal for isolated colonies and suppressed when there was confluent bacterial growth. Both isolates grew well on the selective BCYE media PAC and PAV at 35 °C.

Neither isolate utilized glucose and both were oxidase-, hippurate-, catalase-, β-lactamase- and gelatinase-positive. There were no serological reactions with a species-specific L. pneumophila monoclonal antibody and no reactivity with antibodies to Legionella bozemanii serogroups 1 and 2, Legionella dumoffii serogroups 1 and 2, Legionella feeleii, Legionella gormanii serogroup 1, Legionella jordanis, Legionella longbeachae serogroups 1 and 2, Legionella micdadei serogroups 1 and 2, Legionella oakridgensis, Legionella rubrilucens and Legionella sainthelenis. Immunoblot staining for the presence of L. pneumophila IcmX and DotA showed no reactivity with either isolate.

The cellular fatty acids of strain IMVS-3376T that comprised more than 1 % of the identified fatty acids were iso14:0 (3.9 %), 14:0 (2.1 %), antiso15:0 (20.4 %), 15:0iso0c (2.0 %), 15:0 (3.9 %), iso16:0 (9.0 %), 16:1o7c and 16:1o6c (combined 20.1 %), 16:0 (23.9 %), antiso17:0 (3.9 %), cyclopropane 17:0 (2.6 %), 17:0 (2.0 %) and 18:0 (1.5 %). A repeat analysis from independently grown bacteria gave close results with 10 % or less variation from the initial analysis. No Legionella species in the MIDI database had matching cellular fatty acid compositions; the closest matches were L. feeliei, L. steigerwaltii, L. birminghamensis and L. adelaidensis, with MIDI similarity indices of 0.41, 0.35, 0.28 and 0.26, respectively. MIDI similarity indices of 1.0 and 0.4 indicate complete identity and species level relatedness, respectively.

Both isolates had identical restricted growth temperatures in comparison with L. pneumophila, with minimal and very delayed (5–7 days) growth at 37 °C and no growth at 42 °C, on BCYE medium, with or without 5 % CO2 incubation. In contrast, the bacteria grew well on BCYE medium at 35 °C, with heavy growth after three days, with a very slight growth enhancement with 5 % CO2 supplementation. Growth at 30 °C was as fast as at 35 °C, though with slightly smaller colonies. There was a one- to two-day growth delay at 25 °C in comparison with growth at 30 or 35 °C. Growth of both isolates in BYE broth medium was minimal at 35 °C, with no growth above or below that temperature. BCYE broth supported significantly better growth of both isolates at 35 °C than found for BYE broth, but no growth occurred at the other temperatures tested.

**Intracellular growth in amoebae and human cells.** Isolates IVMS-3376T and F2632 both grew equally well in cultures of Acanthamoeba castellanii at 30 °C, with a 3 log10 increase in bacterial concentrations over a four day period (Fig. 1). In contrast, intra-amoebal growth was less robust at 35 °C than found for BYE broth, though with slightly smaller colonies. There was a one- to two-day growth delay at 25 °C than at 30 or 35 °C. The virulent F889 strain grew well in amoebae at 35 °C, but failed to multiply, but were not killed, in the human respiratory tract cell line A549, at 35 and 37 °C (Fig. 2). The virulent control strain, L. pneumophila F889, grew well at both temperatures, while the avirulent dotA mutant strain, F2352, was killed by the A549 cells.

**Genotypic characterization.** Sequence was recovered from both strains (IVMS-3376T and F2632) from the following...
gene targets: mip (614 nt), 16S rRNA (1362 nt), rpoB (330 nt), rnpB (309 nt) and proA (517 nt). For each gene target, identical sequence was recovered from the two strains, confirming their close genetic similarity despite their diverse origins. The phylogenetic analysis determined that the sequences were not congruent with respect to their diverse origins. The phylogenetic analysis determined that the sequences were not congruent with respect to the inferred evolutionary relatedness of individual species, so the gene sequences were analysed separately. The maximum-likelihood analysis of the mip gene sequences is presented in Fig. 3. Only the clade containing the

Fig. 1. Growth of legionella bacteria in Acanthamoeba castellani at 30 and 35 °C. Axes show bacterial concentration versus days of incubation. Not shown for graphical clarity are the data for bacterial strain F2632, which grew almost identically to strain IMVS-3376T in the amoebae. Results are from triplicate culture wells. Graph represents data from two different experiments. 889, L. pneumophila strain F889; 3376, strain IMVS-3376T.

Fig. 2. Growth of legionella bacteria in A549 cells at 35 and 37 °C. Axes show bacterial concentration versus days of incubation. Not shown for graphical clarity are the data for bacterial isolate F2632, which grew almost identically to strain IMVS-3376T in the cells. Results are from triplicate culture wells. Graph represents data from two different experiments. 3376-37 and 3376-35, isolate IMVS-3376T at 37 and 35 °C, respectively; 2352-35, L. pneumophila dotA mutant F2352 at 35 °C; 889-37 and 889-35, L. pneumophila strain F889 at 37 and 35 °C, respectively.

Discussion

Based on their clear genotypic differences from previously described species of the genus Legionella, strains IVMS-3376T and F2632 are proposed as representing a novel species, Legionella steelei sp. nov. For each of the five genes studied, the corresponding sequences for the strains of L. steelei sp. nov. were distinct from all other species of the

Legionella steeli sp. nov.

L. santicucis ATCC 35301T (U92220)
L. longbeachae ATCC 33462T (X83026)
L. sainhelensi ATCC 35248T (U92219)
L. cincinatiensis ATCC 43753T (U91636)
L. graminea ATCC 49413T (U92206)
L. gormanii ATCC 33297T (U91638)
L. tucsonensis ATCC 49180T (U92224)
L. bozemanae ATCC 33217T (U91609)
L. anisa ATCC 35329T (U91607)
L. parisiensis ATCC 353299T (U92215)
L. wadsworthii ATCC 33877T (U92225)
L. steigerwaltii ATCC 35802T (U92223)
L. cherri ATCC 35252T (U91635)
L. dumoffii ATCC 33279T (U91637)
L. bozemanae ATCC 33297T (U91638)
L. anisa ATCC 35292T (U91639)
L. gormanii ATCC 33297T (U91638)
L. cincinnatiensis ATCC 35248T (U92219)
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Fig. 3. A maximum-likelihood phylogenetic dendrogram of mip sequences from members of the genus Legionella that were genetically closest to L. steelei sp. nov. All Legionella species were included in the analysis, using a GTR + I + G evolutionary model. Coxiella burnetii was used as an outgroup. Bootstrap values (1000 replicates) ≥70 % are displayed. The complete dendrogram is presented in Figure S1 (available in IJSEM Online) and a listing of all the species tested is given in the figure legend.

The similarity of the novel strains to the nearest sister species, compared with that between the closest species pair, for each gene target, is presented in Table 1. Note that for each genetic target, the novel species is less similar to other Legionella species than the most closely related species pair. The lack of congruency of the inferred evolutionary relationships between species from each genetic target suggests complex evolutionary processes such as recombination events have also occurred. To accommodate this evolutionary model, a SplitsTree network analysis was performed on the concatenated sequences (2973 nt). This analysis confirmed that the novel species was located in a clade that comprises L. anisa, L. bozemanae, L. cherrii, L. dumoffii, L. gormanii, L. parisiensis, L. steigerwaltii, L. tucsonensis and L. wadsworthii (Fig. S2).
Table 1. Sequence similarity data for *mip*, 16S rRNA, rpoB, rnpB and proA gene sequences

Values in parentheses are percentage nucleotide similarity for either one or both of the shown species.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Closest sister species to <em>L. steelei</em> sp. nov.*</th>
<th>Most closely related <em>Legionella</em> species†</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>mip</em></td>
<td><em>L. cherrii</em> and <em>L. steigerwaltii</em> (93.6)</td>
<td><em>L. rubrilucens</em> and <em>L. taurinensis</em> (96.3)</td>
</tr>
<tr>
<td>16S rRNA</td>
<td><em>L. bozemanae</em> and <em>L. steigerwaltii</em> (98.5)</td>
<td><em>L. erythra</em> and <em>L. rubrilucens</em> (99.3)</td>
</tr>
<tr>
<td>rpoB</td>
<td><em>L. steigerwaltii</em> (92.2)</td>
<td><em>L. rubrilucens</em> and <em>L. taurinensis</em> (93.6)</td>
</tr>
<tr>
<td>rnpB</td>
<td><em>L. dumoffii</em> (97.4)</td>
<td><em>L. santicricus</em> and <em>L. cincinnatiensis</em> (98.7)</td>
</tr>
<tr>
<td>proA</td>
<td><em>L. cherrii</em> (88.0)</td>
<td><em>L. longbeachae</em> and <em>L. sainheleni</em> (96.2)</td>
</tr>
</tbody>
</table>

*The most closely related species to *L. steelei* sp. nov. for each gene target.
†The most closely related *Legionella* species pair for each gene target.

The genus on the basis of a maximum-likelihood phylogenetic analysis. Further, for every target, the sequence uniqueness was supported by a maximum-likelihood bootstrap analysis (1000 replicates). Such computationally hungry analyses have only recently been made possible by a rapid maximum-likelihood algorithm RAxML and web-based access to large computational resources (Miller et al., 2010). Although the nearest sister species varied for some of the targets, the genetic similarity of the novel strains to the nearest species was greater than that between other recognized species for each target (Table 1). The SplitsTree analysis (Fig. S2), which is based on a concatenation of the sequences from the five gene targets, confirms the genetic uniqueness of *L. steelei* sp. nov. within a tight cluster that contains *L. anisa*, *L. bozemanae*, *L. cherrii*, *L. dumoffii*, *L. gormanii*, *L. parisiensis*, *L. steigerwaltii*, *L. tucsonensis* and *L. wadsworthii*. With the exception of *L. wadsworthii*, all of species exhibit blue–white fluorescence, as does strain IMVS-3376.

This novel species is temperature growth restricted in vitro, with minimal growth in vitro at temperatures greater than 35 °C, and negligible growth in vivo in human cells at either 35 or 37 °C. It has a strong ability to grow well in amoebae at 30 and 35 °C, with superior growth at 30 °C. In contrast, *L. pneumophila* grows well on artificial media over a much broader temperature range, from 25 to 42 °C, and is fully virulent for human cells at 37 °C. The excellent growth of *L. steelei* sp. nov. in amoebae at 30 °C, and its inability to establish a productive infection in human cells suggests that this bacterium is primarily an amoebal pathogen, with little ability to cause disease in humans. Whether either of the patients from whom this bacterium was isolated had Legionnaires’ disease is uncertain, but it is unlikely due to the clinical course of the disease and the consultants’ opinions. These bacteria were probably isolated from humans because they were transient colonizers, or perhaps contaminants. It is however possible that this bacterium can cause infection in severely immunocompromised patients.

Specific laboratory identification of *L. steelei* sp. nov. using simple phenotypic methods is not possible, although an inability to grow well, or at all, at 37 °C should be a clue to its identity. In addition, production of large amounts of diffusible bright yellow fluorescent pigment when the isolate is grown on BCYEz medium, particularly if accompanied by cell-bound blue–white autofluorescence would be an additional clue to its identity. Cellular fatty acid analysis yields an apparently unique pattern and such an analysis could be useful for full species identification. Sequencing of the *mip* gene using a standardized method and published databases is the easiest and most specific means of species identification. Use of partial 16S rRNA gene sequencing may, in contrast, lead to incorrect species assignment.

Although not an apparent human pathogen, *L. steelei* sp. nov. may be a valuable model organism to study legionella pathogenesis in low temperature hosts, such as *Dictyostelium discoideum* and zebrafish, assuming that it is virulent at 20 to 25 °C in these hosts. It will also be important to determine whether *L. steelei* sp. nov. has a *dot/icm*-like type IV secretion system, as the failure to detect immunoreactive *L. pneumophila* DotA or IcmX in this study does not exclude such a possibility.

It is possible that *L. steelei* sp. nov. is commonly found in water because it grows well in amoebae at lower temperatures. It may be that its restricted growth temperature has hindered isolation from environmental specimens.

**Description of *Legionella steelei* sp. nov.**

*Legionella steelei* (steel’i. N.L. masc. gen. *steelei* of Steele, pertaining to the Australian microbiologist Trevor Steele, who performed pioneering work on the ecology and pathogenesis of *L. longbeachae* infection).

Gram-negative-staining rod. Grows on BCYEz agar, but not on tryptic soy blood agar or BCYEz agar without L-cysteine. Optimal growth temperature is 35 °C, with no or very poor growth at ≥37 °C. Produces large amounts of diffusible yellow–green fluorescent pigment. Variable production of colony-bound blue–white fluorescence. Positive in tests for activities oxidase, catalase, gelatinase, hippurate hydrolysis and β-lactamase. Negative result for glucose utilization. Motile, with monopolar flagella. Dominant cellular fatty acids are anteiso15:0, 16:1 and 16:0. Virulent in *Acanthamoeba*...
castellani. 16S rRNA, mip, rpoB, rnpB and proA gene sequences differ significantly from all recognized species of the genus Legionella.

The type strain, IMVS-3376T (=IMVS 3113T=ATCC BAA-2169T), was isolated from a human respiratory tract specimen.

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References


