Selenomonas bovis sp. nov., isolated from yak rumen contents

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Two strictly anaerobic, crescent-shaped bacterial strains, designated WG7 and Ycb08, were isolated from a cellulose-degrading mixed culture enriched from yak rumen contents. The strains were Gram-negative, non-spore-forming and motile, with four to six flagella situated at the centre of the concave side of the cell. The cells were 0.9–1.1×4–6 μm. Growth was observed at 27–46 °C (optimum 39 °C) and pH 4.2–8.3 (optimum pH 7.0–7.2). Arabinose, glucose, mannose, cellobiose, lactose, sucrose, trehalose, melibiose, raffinose, salicin and aesculin were fermented. The end products of glucose fermentation were acetate, propionate and CO2. The G+C contents of strains WG7 and Ycb08 were respectively 63.9 ± 0.2 and 62.5 ± 0.2 mol% (Tm). Phylogenetic analysis based on 16S rRNA gene sequences revealed that the two strains were related to the genera Mitsuokella and Selenomonas at similarity levels below 97%; however, they differed from members of the genus Mitsuokella in their flagellar arrangement. On the basis of phenotypic, genotypic and physiological evidence, strains WG7 and Ycb08 are identified as members of a novel species of the genus Selenomonas, for which the name Selenomonas bovis sp. nov. is proposed. The type strain is WG7 (=CGMCC 1.5073T =JCM 15470T).

The yak (Bos grunniens) is a ruminant found mainly on the Qinghai–Tibet Plateau, China, at altitudes over 3000 m above sea level, with grasses as their main food. Previous work in our laboratory (An et al., 2005) based on a culture-independent approach determined that more than 60 % of the bacteria in the yak rumen clustered in not-yet-cultured groups, displaying <90 % 16S rRNA gene sequence similarity with cultured species. In addition, a relatively large proportion of the species detected in the yak rumen grouped with fibrolytic bacteria isolated previously from rumen contents, suggesting that the yak rumen could harbour novel types of fibrolytic bacteria. Therefore, by using filter paper as the sole carbon source, we obtained a cellulose-degrading mixed-culture enrichment with yak rumen content as inoculant; a few fibrolytic and non-fibrolytic bacterial strains were isolated from the mixed culture. In this study, we describe two non-fibrolytic bacteria strains that represent a novel species of the genus Selenomonas.

Yak rumen content was inoculated into a modified basal medium (Hungate, 1966) with filter paper as the sole carbon source and grown under a gas phase of N2/CO2 (80:20). The medium contained 20 % rumen fluid and 80 % basal medium. A fibrolytic mixed culture was obtained by subculturing the enrichment in the same medium for eight to ten transfers. After serial dilution in peptone-yeast-extract-glucose (PYG) broth (Holdeman et al., 1977) and the use of the Hungate roll-tube technique (Hungate, 1969), single colonies were observed after incubation at 39 °C for 2 days. Colonies were picked and transferred to the same medium. The roll-tube procedure was repeated several times before strains WG7 and Ycb08 were obtained. The purity of the isolates was examined by light microscopy. All inoculations and transfers were done with syringes and needles, and the cultures were incubated at 39 °C unless indicated. Selenomonas ruminantium subsp. ruminantium DSM 2150T, S. ruminantium subsp. lactilytica DSM 2872T, Selenomonas sputigena DSM 20758T and Mitsuokella jalaludinii DSM 13811T were purchased from the Deutsche Sammlung von Mikroorganismen und Zellkulturen (Braunschweig, Germany) and served as reference strains in physiological characterization tests.

Cell morphology was examined under a light microscope (Olympus BH-2) and an electron microscope (Hitachi H-600A). For electron microscopy studies, bacterial cells were negatively stained with uranyl acetate. Spore formation was examined by phase-contrast microscopy at the end of growth as well as by heat treatment in a water bath at 80 °C for 10 min.
Cells were Gram-negative, typical curved rods, 0.9–1.1 × 4–6 μm, and occurred singly or in pairs. Spores were never observed, and the strains could not survive heat treatment at 80°C for 10 min. Negatively stained cells revealed the presence of four to six flagella in a tuft situated near the centre of the concave side (Supplementary Fig. S1, available in IJSEM Online).

Growth was determined by monitoring the OD₆₀₀ of the culture in PYG medium at 39°C. The strains were strictly anaerobic; no growth occurred when exposed to air. The generation time of strain WGᵀ was determined as 2.3 h by monitoring the OD₆₀₀ at 30 min intervals for 24 h. The temperature profile of strain WGᵀ was determined in a water bath from 20 to 50°C at 1°C intervals (pH 7.0) and growth was observed between 27 and 46°C, with optimal growth at 39°C. The pH range for growth was pH 4.2–8.3, with optimal growth around pH 7.0–7.2, as determined in PYG broth at various pH values adjusted with Na₂HPO₄ and NaH₂PO₄.

The end products from glucose fermentation were determined by growing strains WGᵀ and Ycb08 in PYG medium; short-chain fatty acids were then analysed by using a gas chromatograph (GC-14B; Shimadzu) equipped with a flame-ionization detector (column temperature 230°C, injector 250°C, detector 280°C). The carrier gas was N₂ at a flow rate of 30 ml min⁻¹. It was determined that strain WGᵀ and Ycb08 produced acetate, propionate and CO₂ from glucose at a molar ratio of 0.7:1.1:1.2.

Substrate utilization was tested by growing the two strains in PY medium (Holdeman et al., 1977) separately containing each of the following substrates: lactate, glycerol, dulcitol, mannitol, sorbitol, xylose, arabinose, glucose, mannose, cellobiose, lactose, sucrose, trehalose, melibiose, raffinose, inulin, salicin and aesculin. Stock solutions of the test compounds were prepared anaerobically, sterilized by filtration and injected into PY medium; short-chain fatty acids were then analysed by GC-14B (Shimadzu) equipped with a consensus 1379 bp 16S rRNA gene sequence, a phylogenetic tree rooted with Bacillus subtilis DSM 10ᵀ was constructed (Fig. 1). It was shown that the two strains clustered with the members of the genera Selenomonas and Mitsuokella and were closest to M. jaluadauni M 9ᵀ (95.5% 16S rRNA gene sequence similarity), followed by S. ruminantium subsp. ruminantium GA192ᵀ (95.4%), S. ruminantium subsp. lactyltyca HD₄ (95.2%) and Mitsuokella multacidica NCTC 10934ᵀ (95.1%). According to the well-accepted bacterial species delimitation of less than 97% 16S rRNA gene sequence similarity (Wayne et al., 1987), strains WGᵀ and Ycb08 could represent a novel species affiliated either to the genus Selenomonas or Mitsuokella.

Based on the genus descriptions, members of Mitsuokella are characterized by a cell morphology of stout rods without flagella (Mitsuoka et al., 1974; Holdeman et al., 1984; Lan et al., 2002), while cells of the genus Selenomonas are curved to helical rods, motile with a typical flagellar arrangement as a tuft near the centre of the concave side (Bryant, 1984; Kingsley & Hoeningr, 1973). Strain WGᵀ presented crescent-shaped cells with flagella on the concave side; together with other differential characteristics listed in Table 1, the two strains can be classified in the genus Selenomonas. Furthermore, the two strains resembled other members of the genus Selenomonas in metabolic features, such as the use of a variety of carbohydrates as substrates and the production of acetate, propionate and CO₂ from glucose, as well as production of H₂S from L-cysteine but the absence of nitrate reduction (Table 1). However, strains WGᵀ and Ycb08 displayed <97% 16S rRNA gene sequence similarity with described Selenomonas species and differed from their closest relative, S. ruminantium subsp. ruminantium DSM 2150ᵀ, in their higher G+C content and by liquefying gelatin but not fermenting dulcitol, mannitol, sorbitol and xylose. Thus, on the basis of DNA G+C content and the physiological and phylogenetic traits described above, strains WGᵀ and Ycb08 represent a novel
species of the genus *Selenomonas*, for which the name *Selenomonas bovis* sp. nov. is proposed.

**Description of Selenomonas bovis** sp. nov.

*Selenomonas bovis* (bo′vis. L. gen. n. bovis of a cow, of a bovine, referring to the isolation of the first strains from yak rumen contents).

Gram-negative, crescent-shaped rods, 4–6 μm long and 0.9–1.1 μm wide. No spore formation is observed. Flagella (four to six) are arranged linearly as a tuft near the centre of the concave side of the cell. The temperature range for growth is 27–46 °C; optimum growth at 39 °C. Grows at pH 4.2–8.3, with optimum growth at pH 7.0–7.2. The generation time is 2.3 h when grown on glucose at 39 °C. The final pH in PYG medium is 4.5. Utilizes arabinose, glucose, mannose, cellobiose, lactose, sucrose, trehalose, melibiose, raffinose, salicin and aesculin but not lactate, xylose, starch, dulcitol, mannitol or sorbitol. The end products of glucose fermentation are acetate, propionate and CO₂ according to the equation 1 glucose → 0.7 acetate + 1.1 propionate + 1.2 CO₂. Liquefies gelatin and...

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**Table 1.** Differential characteristics of strains WG³ and Ycb08 and phylogenetically related type strains

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<tr>
<td><strong>DNA G+C content (mol%)</strong></td>
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<td>62.5 ± 0.2</td>
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<td>61</td>
<td>56.8</td>
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*CR, Curved rods; SR, stout rods.*

†LA, Linearly arranged as a tuft near the centre of the concave side; NF, no flagella.

‡Data from this study.
produces H₂S but not indole. Does not reduce nitrate. Voges–Proskauer and methyl red tests are negative. The G+C content of the genomic DNA of the type strain is 63.9 ± 0.2 mol%.

The type strain, WGT (=CGMCC 1.5073T =JCM 15470T), and a second strain, Ycb08, were isolated from yak rumen contents.

Acknowledgements

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References


