Aspergillus brasiliensis sp. nov., a biseriate black Aspergillus species with world-wide distribution

János Varga,¹, ¹ Sándor Kocsubé,¹ Beáta Tóth,² Jens C. Frisvad,³ Giancarlo Perrone,⁴ Antonia Susca,⁴ Martin Meijer⁵ and Robert A. Samson⁵

¹Department of Microbiology, Faculty of Sciences, University of Szeged, PO Box 533, H-6701 Szeged, Hungary
²Cereal Research Non-Profit Company, PO Box 391, H-6701 Szeged, Hungary
³Center for Microbial Biotechnology, BioCentrum-DTU, Building 221, Technical University of Denmark, DK-2800 Kgs Lyngby, Denmark
⁴Institute of Sciences of Food Production, CNR, Via Amendola 122/O, 70126 Bari, Italy
⁵CBS Fungal Biodiversity Centre, Uppsalaalan 8, 3584 CT Utrecht, The Netherlands

A novel species, Aspergillus brasiliensis sp. nov., is described within Aspergillus section Nigri. This species can be distinguished from other black aspergilli based on intergenic transcribed region, β-tubulin and calmodulin gene sequences, by amplified fragment length polymorphism analysis and by extrolite profiles. A. brasiliensis isolates produced naphtho-γ-pyrones, tensidol A and B and pyrophen in common with Aspergillus niger and Aspergillus tubingensis, but also several unique compounds, justifying their treatment as representing a separate species. None of the isolates were found to produce ochratoxin A, kotanins, funalenone or pyranonigrins. The novel species was most closely related to A. niger, and was isolated from soil from Brazil, Australia, USA and The Netherlands, and from grape berries from Portugal. The type strain of Aspergillus brasiliensis sp. nov. is CBS 101740T (＝IMI 381727＝IBT 21946).

INTRODUCTION

Black aspergilli (Aspergillus section Nigri; Gams et al., 1985) have a significant impact on modern society. Many species cause food spoilage, and several are used in the fermentation industry to produce hydrolytic enzymes, such as amylases or lipases, and organic acids, such as citric acid and gluconic acid (Varga et al., 2000). They are also candidates for genetic manipulation in the biotechnology industries as Aspergillus niger used under certain industrial conditions has been granted the GRAS (generally regarded as safe) status by the Food and Drug Administration of the USA government. Although the main source of black aspergilli is soil, members of this section have been isolated from various other sources (Kozakiewicz, 1989; Abarca et al., 2004; Samson et al., 2004). Besides their economical importance, black aspergilli are also important as ochratoxin-producing organisms that contaminate several agricultural products including grape-derived products, coffee and cocoa (Cabanes et al., 2002; Samson et al., 2004).

Black aspergilli are one of the more difficult groups regarding classification and identification. The taxonomy of Aspergillus section Nigri has been studied by many taxonomists and was recently reviewed by Abarca et al. (2004). Nuclear and mitochondrial DNA (mtDNA) polymorphisms and PCR-based techniques led to the recognition of at least two species within the A. niger species complex (Aspergillus niger, Aspergillus tubingensis) (Kusters-van Someren et al., 1991; Varga et al., 1994). Regarding other black Aspergillus species, phylogenetic analyses of sequences of the intergenic transcribed spacer and the 5.8S rRNA gene (ITS region) and the D1–D2 region of the 28S rRNA gene indicated that, apart from those mentioned earlier, at least five other species belong to section Nigri: Aspergillus heteromorphus, Aspergillus ellipiticus, Aspergillus carbonarius, Aspergillus japonicus and Aspergillus aculeatus (Varga et al., 2000; Parenicova et al., 2001). Several other black Aspergillus species have been

Abbreviations: AFLP, amplified fragment length polymorphism; ITS, intergenic transcribed spacer; SEM, scanning electron microscopy.

The GenBank/EMBL/DDBJ accession numbers for the β-tubulin, ITS and calmodulin gene sequences determined in this study are shown in Table 1.

The Mycobank accession number for Aspergillus brasiliensis sp. nov. is MB510581 (http://www.mycobank.org).

Neighbour-joining trees based on ITS and calmodulin gene sequence data of Aspergillus section Nigri, a dendogram based on cluster analysis of AFLP data and tables listing the Aspergillus section Nigri strains used in AFLP analysis and the extrolites produced by the Aspergillus brasiliensis isolates are available with the online version of this paper.
described recently, including *Aspergillus vadensis* (de Vries et al., 2005), *Aspergillus costaricaensis*, *Aspergillus piperis*, *Aspergillus lacticoffeatus* and *Aspergillus sclerotioniger* (Samson et al., 2004) and *Aspergillus ibericus* (Serra et al., 2006).

During a survey of black *Aspergillus* isolates collected worldwide, we discovered some strains that did not fit into any species of *Aspergillus* section *Nigri*. We have used a polyphasic taxonomic approach in order to determine the delimitation and variability of this novel species. For phenotypic analyses, macro- and micromorphologies of the isolates were examined, and secondary metabolite and enzyme profiles were studied. For genotypic studies, partial sequences of the β-tubulin and calmodulin genes and the ITS region of the rRNA gene cluster and amplified fragment length polymorphism (AFLP) profiles were analysed (also used by Geiser et al., 1998; Varga et al., 2000; Hong et al., 2006; Perrone et al., 2004, 2006).

**METHODS**

The strains examined are listed in Table 1 and were maintained on malt extract autolysate (MEA) agar slants.

**Table 1. Origin and GenBank accession numbers of sequences of the isolates examined in this study**

<table>
<thead>
<tr>
<th>Isolate</th>
<th>Origin</th>
<th>Accession no.</th>
</tr>
</thead>
<tbody>
<tr>
<td>CBS 101740T=IMI 381727T=JHC 614T</td>
<td>Soil, Pedreira, São Paulo, Brazil</td>
<td>AJ280010 AM295175</td>
</tr>
<tr>
<td>JHC 601</td>
<td>Soil, St Rosa do Viterbo, Brazil</td>
<td>DQ900599 AM295186</td>
</tr>
<tr>
<td>JHC 603</td>
<td>Soil, St Rosa do Viterbo, Brazil</td>
<td>DQ900601 DQ900610 †</td>
</tr>
<tr>
<td>JHC 605</td>
<td>Soil, Araçatuba, São Paulo, Brazil</td>
<td>† †</td>
</tr>
<tr>
<td>JHC 606</td>
<td>Soil, A. Nogueira, São Paulo, Brazil</td>
<td>†</td>
</tr>
<tr>
<td>JHC 607</td>
<td>Soil, Campinas, São Paulo, Brazil</td>
<td>DQ900600 DQ900611 †</td>
</tr>
<tr>
<td>CBS 246.65=IBT 28083</td>
<td>Soil, Sydney, New South Wales, Australia</td>
<td>DQ900597 DQ900607 †</td>
</tr>
<tr>
<td>CBS 733.88=IBT 28084</td>
<td>Soil, North Carolina, USA</td>
<td>DQ900598 DQ900612 †</td>
</tr>
<tr>
<td>CBS 116970=IBT 28085</td>
<td>Soil, The Netherlands</td>
<td>DQ900596 DQ900613 †</td>
</tr>
<tr>
<td>ITEM 4540</td>
<td>Grapes (Cabernet Sauvignon), Ribatejo Region, Portugal, 2001</td>
<td>AM295178 AM295179</td>
</tr>
<tr>
<td>ITEM 4544</td>
<td>Grapes (Tinta Miúda), Ribatejo Region, Portugal, 2001</td>
<td>‡ AM295177</td>
</tr>
<tr>
<td>ITEM 4544T</td>
<td>Grapes (Vinhão), Vinhos Verdes Region, Portugal, 2001</td>
<td>‡ AM295174</td>
</tr>
<tr>
<td>ITEM 4539</td>
<td>Grapes (Tinta Barroca), Douro Region, Portugal, 2002</td>
<td>‡ AM295179</td>
</tr>
</tbody>
</table>

*CBS, Centraalbureau voor Schimmelcultures, Utrecht, The Netherlands; JHC, James H. Croft’s culture collection, Birmingham, UK; IBT, BioCentrum-DTU, Kgs. Lyngby, Denmark; IMI, CABI Bioscience Genetic Resource Collection, Egham, UK; ITEM, Agri-Food Toxigenic Fungi Culture Collection, Institute of Sciences of Food Production, Bari, Italy.
†100% identity with the deposited sequence AM295175.
‡100% identity with the deposited sequence AM295180.

**Morphological analysis.** For macromorphological observations, Czapek yeast autolysate (CYA), MEA agar, Czapek yeast autolysate with 5% NaCl (CYAS) agar, yeast extract-sucrose (YES) agar, oatmeal agar (OA) and Czapek agar (CZA) were used (Samson et al., 2004). The isolates were inoculated at three points on each plate of each medium and incubated at 25 °C in the dark for 7 days. For micromorphological observations, microscopic mounts were made in lactic acid from MEA colonies and a drop of alcohol was added to remove air bubbles and excess conidia. Scanning electron microscopy (SEM) was performed using a Hitachi S570 microscope. For SEM preparation, conidia were transferred to aluminium stubs using double-sided adhesive tape. A small drop of 10 mM ACES buffer containing 0.05% Tween 80 was added to the conidiophores. The suspension was air-dried and coated with platinum. The strains were also inoculated on creatine-sucrose agar (CREA), CYA at 37 °C and on CYA with 5% NaCl (Frisvad & Samson, 2004).

**Extrolite analysis.** Extrolites were analysed by HPLC using alkylphenone retention indices and diode array UV-VIS detection as described by Frisvad & Thrane (1987), with modifications as described by Smedsøgaard (1997). Standards of ochratoxin A and B, aflavinines, asperazine, astudiol, kotanin and other extrolites from the collection at BioCentrum-DTU (Denmark) were used to compare the extrolites from the species under study, Pyrononigrin A, tensidol A and B and pyrophlen were identified by comparison with literature UV and MS data (Hiort et al., 2004; Fukuda et al., 2006).
Isolation and analysis of nucleic acids. Total nucleic acids were isolated according to the literature (Leach et al., 1986). Fragments containing the region encoding the intergenic transcribed spacer 1 (ITS-1), 5.8S rDNA and intergenic transcribed spacer 2 (ITS-2) were amplified using primers ITS1 and ITS4, as described previously (Varga et al., 2000) and by White et al. (1990). Amplification of part of the β-tubulin gene was performed using the primers Bt2a and Bt2b (Glass & Donaldson, 1995; Samson et al., 2004). Amplifications of the partial calmodulin gene were set up as described previously (Serra et al., 2006). Sequence analysis was performed with a Big Dye Terminator Cycle Sequencing Ready Reaction kit for both strands, and the sequences were aligned with the MT Navigator software (Applied Biosystems). All the sequencing reactions were purified by gel filtration through Sephadex G-50 (Amersham Pharmacia Biotech) equilibrated in double-distilled water and analysed on an ABI PRISM 310 Genetic Analyzer (Applied Biosystems). The resulting sequences of all the isolates were aligned by using the CLUSTAL method with the program DNAMAN (Lynnon Corporation). The unique ITS, β-tubulin and calmodulin sequences were deposited in the GenBank nucleotide sequence database (Table 1).

AFLP analysis. Twenty-three strains belonging to Aspergillus section Nigri were analysed by AFLP analysis (see Supplementary Table S1 available in IJSEM Online). Fungal strains were grown in shake cultures (150 r.p.m., 25 °C, 2 days) in Wickerham’s medium (40 g glucose, 5 g peptone, 3 g yeast extract, 3 g malt extract and distilled water to 1 l). Genomic DNA was extracted using an E.Z.N.A. Fungal DNA Miniprep kit (Omega Bio-tek), according to the manufacturer’s protocol. The DNA was dissolved in sterile water, diluted to 20 ng μl⁻¹ and stored at −20 °C.

We used an AFLP Microbial Fingerprinting kit (Applied Biosystems-Perkin-Elmer Corporation) according to the manufacturer’s instructions using primer combinations according to Perrone et al. (2006). Peak height thresholds were set at 200. Genotype software (Applied Biosystems) was set to medium smoothing. Bands of the same size in different individuals were assumed to be identical and to represent the same allele. Bands of different sizes were treated as independent loci with two alleles (present and absent). Data were analysed with an AFLP manager database developed by ACGT Bioinformatics S.r.l. and were exported in a binary format with ’1′ for the presence of a band/peak and ‘0’ for its absence. For clustering two different analyses were performed, fragments between 100 and 500 bp and between 200 and 500 bp were analysed with NTSYS software by using the Dice band/peak and ‘0’ for its absence. For clustering two different analyses with two alleles (present and absent). Data were analysed with an AFLP analysis.

Analysis of sequence data. Sequence alignments were performed by using CLUSTAL-X (Thompson et al., 1997) and improved manually. Evolutionary distances between the sequences were calculated using Kimura’s formula (Kimura, 1980) with the program DNADIST of the PHYLIP program package (Felsenstein, 1995). Phylogenetic trees were prepared by using the neighbour-joining method (Saitou & Nei, 1987) with the program NEIGHBOR of the PHYLIP package. Bootstrap values were calculated from 1000 replications of the bootstrap procedure using programs SEQBOOT, DNADIST, NEIGHBOR and CONSENSE of the package (Felsenstein, 1985, 1995). For parsimony analysis, the PAUP* version 4.0 software was used (Swofford, 2000). Aspergillus flavus CBS 100927T was used as an outgroup in these experiments.

RESULTS AND DISCUSSION

Isolates of the novel species described below have an unique echinulate conidium surface ornamentation, in contrast to all other members of the Aspergillus niger complex or the A. niger clade in Samson et al. (2004). The conidia of the novel species were more similar to those of A. aculeatus and A. carbonarius, but differed from those of the former species in being biseriate and from those of A. carbonarius in being much smaller. Physiological features such as very good growth and sporulation at 37 °C, poor growth but strong acid production on CREA agar, and very good growth and sporulation on CYA agar with 5 % NaCl were indicative of a close relationship with A. niger and A. tubingensis. The strong acid production on CREA agar indicates that the novel species can produce citric acid. Like A. niger, the novel species also produces xylanases and thermostable β-xyllosidases (Pedersen et al., 2007). Regarding extrolite profiles, the isolates of the novel species produced naphtho-γ-pyrones (including aurasperone B), pyrophen (Barnes et al., 1990) and tensidol A and B (Fukuda et al., 2006), in common with A. niger and A. tubingensis, but also produced several unique compounds (see Supplementary Table S2 in IJSEM Online), justifying the treatment of the isolates as representing a separate species. None of the isolates were found to produce ochratoxin A, kotanins, furalenone, antafumicins, asperazine or pyranonigrins, common to other species in the A. niger complex (Table 2). Two isolates produced large white sclerotia (ITEM 4544 and ITEM 6139). These strains contained the same sclerotal indol alkaloids as A. piperis and A. costaricaeensis (Samson et al., 2004). From a chemotaxonomic point of view, we regard isolates to be members of a novel species if they produce unique combinations of extrolites not seen in any other species. In this case some species-specific, but not yet structure-elucidated, extrolites were present in the novel species, whereas certain extrolites regarded as being species specific for other species in Aspergillus section Nigri were not present in the novel species.

In previous studies, a wide-ranging variation in mtDNA restriction fragment length polymorphism profiles was observed both among collection strains and in natural populations of the A. niger species complex (Varga et al., 1993, 1994). Most isolates were classified as A. niger or A. tubingensis according to their HaeIII–BglII digested mtDNA patterns. A. niger and A. tubingensis were grouped into five and six mtDNA types, respectively. However, six of the 73 Brazilian isolates examined exhibited unique mtDNA restriction fragment length polymorphism patterns. A. niger and A. tubingensis were grouped into five and six mtDNA types, respectively. However, six of the 73 Brazilian isolates examined exhibited unique mtDNA restriction fragment length polymorphism patterns. A. niger and A. tubingensis were grouped into five and six mtDNA types, respectively. However, six of the 73 Brazilian isolates examined exhibited unique mtDNA restriction fragment length polymorphism patterns. A. niger and A. tubingensis were grouped into five and six mtDNA types, respectively. However, six of the 73 Brazilian isolates examined exhibited unique mtDNA restriction fragment length polymorphism patterns. A. niger and A. tubingensis were grouped into five and six mtDNA types, respectively. However, six of the 73 Brazilian isolates examined exhibited unique mtDNA restriction fragment length polymorphism patterns. A. niger and A. tubingensis were grouped into five and six mtDNA types, respectively. However, six of the 73 Brazilian isolates examined exhibited unique mtDNA restriction fragment length polymorphism patterns. A. niger and A. tubingensis were grouped into five and six mtDNA types, respectively. However, six of the 73 Brazilian isolates examined exhibited unique mtDNA restriction fragment length polymorphism patterns.
Table 2. Morphological characteristics and extrolite production of species belonging to the *A. niger* species complex in *Aspergillus* section *Nigri*

<table>
<thead>
<tr>
<th>Species</th>
<th>Conidial size (μm)</th>
<th>Vesicle size (μm)</th>
<th>Colour and size of sclerotia (mm)</th>
<th>Extrolites produced</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>A. brasiliensis</em> sp. nov</td>
<td>3.5–4.5</td>
<td>30–45</td>
<td>Found only in some strains, white, 1–1.5</td>
<td>Naphtho-γ-pyrones (including aurasperone B), pyrophen, tensidol A and B, (dihydrocarolic acid, 14-epi-14-hydroxy-10,23-dihydro-24,25-dehydro-aflavinine, 10,23-dihydro-24,25-dehydro-aflavinine)*</td>
</tr>
<tr>
<td><em>A. costaricaensis</em></td>
<td>3.1–4.5</td>
<td>40–80</td>
<td>Pink to yellow, 1.2–1.8</td>
<td>Aflavinines (see above), corymbiferan lactones, funalenone, naphtho-γ-pyrones</td>
</tr>
<tr>
<td><em>A. foetidus</em></td>
<td>3.5–4.5</td>
<td>50–80</td>
<td>–</td>
<td>Antafumicins, asperazine, funalenone, naphtho-γ-pyrones, pyranonigrin A</td>
</tr>
<tr>
<td><em>A. lacticoffeatus</em></td>
<td>3.4–4.1</td>
<td>40–65</td>
<td>–</td>
<td>Kotanins, ochratoxin A, pyranonigrin A, tensidol B</td>
</tr>
<tr>
<td><em>A. niger</em></td>
<td>3.5–5</td>
<td>45–80</td>
<td>–</td>
<td>Funalenone, (kotanins), naphtho-γ-pyrones, (ochratoxin A), pyranonigrin A, pyrophen, tensidol A and B</td>
</tr>
<tr>
<td><em>A. piperis</em></td>
<td>2.8–3.6</td>
<td>40–55</td>
<td>Yellow to pink–brown, 1.0–1.7</td>
<td>Aflavinines (see above), naphtho-γ-pyrones, pyranonigrin A</td>
</tr>
<tr>
<td><em>A. tubingensis</em></td>
<td>3–5</td>
<td>40–80</td>
<td>White to pink, 0.5–0.8</td>
<td>Asperazine, funalenone, naphtho-γ-pyrones, pyranonigrin A, tensidol A and B</td>
</tr>
<tr>
<td><em>A. vadensis</em></td>
<td>3–4</td>
<td>25–35</td>
<td>–</td>
<td>Asperazine, funalenone, naphtho-γ-pyrones, nigragillin, polar kotanin-like compound</td>
</tr>
</tbody>
</table>

*These extrolites were only produced by isolates producing sclerotia.

The Netherlands, and from grape berries from Portugal (Table 1).

We examined the genetic relatedness of these isolates to other black aspergilli using sequence analysis of the ITS region of the rRNA gene cluster and parts of the calmodulin and β-tubulin genes. The isolates were found to form a monophyletic clade supported by high bootstrap values on phylogenetic trees based on β-tubulin, ITS and calmodulin sequence data (Fig. 1 and Supplementary Figs S1 and S2 in IJSEM Online). During analysis of part of the β-tubulin gene, 392 characters were analysed. Among the 193 polymorphic sites, 160 were found to be phylogenetically informative. The neighbour-joining tree based on partial β-tubulin gene sequences is shown in Fig. 1. The topology of the tree is the same as that of a parsimony tree constructed by using the program PAUP (length, 414 steps; consistency index, 0.6836; retention index, 0.8039). The calmodulin dataset included 671 characters, with 283 parsimony informative characters. The topologies of the neighbour-joining tree (Supplementary Fig. S1) and the parsimony tree were the same (tree length, 718; consistency index, 0.6964; retention index, 0.8546). The ITS dataset included 479 characters with 70 parsimony informative characters. The neighbour-joining tree shown in Supplementary Fig. S2 has the same topology as the parsimony tree (tree length, 139; consistency index, 0.8849; retention index, 0.9342).

These isolates also formed a well-defined cluster on an UPGMA tree based on AFLP data (see Supplementary Fig. S3 in IJSEM Online). AFLP data have the special advantage of containing both individual (fingerprinting) and species-specific information. Our data indicate that these isolates are well separated from other black aspergilli based on all molecular approaches used. During AFLP analysis, clear polymorphisms both within and between species were obtained for each of the four primer pairs. Each primer combination consistently distinguished the nine different species of black aspergilli analysed by AFLP with similarity among the different species of less than 20 % (Supplementary Fig. S3). The 11 strains that were grouped by sequence analysis as *A. brasiliensis* formed a main AFLP cluster that was clearly differentiated from all the other species of section *Nigri*, but they showed a great degree of genetic variability among each other with a similarity of 25 % (Dice similarity index). In particular, two main clusters were formed within the *A. brasiliensis* group; the first grouped the four Portuguese strains from grapes at a similarity of 52 %, and the second one grouped at a similarity of 42 % all other strains isolated from soil from Brazil, Australia, USA and The Netherlands.

Our *Aspergillus* species concept is based on a polyphasic approach (Frisvad & Samson, 2004): A novel species is different from any other species in a diagnostic sense in both phenotypic and genotypic features. Here we have used morphological, physiological and chemotaxonomical features to characterize the phenotype and sequencing of three genes combined with AFLP results to characterize the isolates genotypically. Because the isolates were unique with regard to morphology, extrolite profiles and genotypic features, we propose the name *Aspergillus brasiliensis* sp. nov. for these isolates.

The name *brasilienis* refers to the locality where the culture was isolated. Since its discovery the species has also been
found in other localities, but we propose to maintain the name to avoid confusion because the epithet has been used and cited in various publications.

**Latin diagnosis of Aspergillus brasiliensis Varga, Frisvad & Samson sp. nov.**

*Coloniae post 7 dies 71–76 mm diam in agaro CYA et CYAS dicto, in MEA 52–70 mm, in YES 75–80 mm, in agaro farina avenacea confecto 32–36 mm, in CREA 32–44 mm. Coloniae primum albae, deinde obscure brunneae vel atrae, reverseum cremeum vel dilute brunneum. Conidiorum capitula primum globosa, deinde radiantia, nonnumquam in nonnullas columnas divisa; stipes 700–1700 μm, walls thick, smooth, pale brown; vesiculae 30–45 μm wide, nearly globose; metulae covering virtually the entire surface of the vesicle, measuring 22–30 × 3–6 μm; phialides lageniformes, 7–9 × 3–4 μm; conidia subglobose, 3.5–4.8 μm diam, echinulate. No sclerotia observed in the culture ex-type. All isolates produced several naphthopyrones (including auraspereone B), tensidol A and B and pyrophen. The type strain, CBS 101740T (=IMI 381727T=IBT 21946T), was isolated from soil, Pedreira, São Paulo, Brasilia.**

**Description of Aspergillus brasiliensis Varga, Frisvad & Samson sp. nov.**

*Aspergillus brasiliensis* (bra.si.li.en’sis. N.L. masc. adj. brasiliensis from Brazil, the place of isolation).

Colony diameters at 7 days: CYA at 25 and 37 °C, and CYAS at 25 °C: 71–76 mm; MEA 52–70 mm; YES 75–80 mm; OA 32–36 mm; CREA 32–44 mm, poor growth, strong acid production. Colony first white then dark brown to black (Fig. 2). Exudates absent, reverse cream-coloured to light brown. Conidial heads globose at first and later radiate occasionally developing into several conidial columns; stipes 700–1700 × 8–13 μm, walls thick, smooth, pale brown; vesicles 30–45 μm wide, nearly globose; biseriate; metulae covering virtually the entire surface of the vesicle, measuring 22–30 × 3–6 μm; phialides flask-shaped, 7–9 × 3–4 μm; conidia subglobose, 3.5–4.8 μm in diameter, echinulate. No sclerotia observed in the culture ex type. All isolates produced several naphthopyrones (including auraspereone B), tensidol A and B and pyrophen. The type strain, CBS 101740T (=IMI 381727T=IBT 21946T), was isolated from soil, Pedreira, São Paulo, Brasilia.
Fig. 2. Aspergillus brasiliensis sp. nov. CBS 101740T. (a) Colonies on CYA; (b) colonies on OA; (c) colonies on MEA; (d–g) conidiophores; (h) conidia under light microscope; (i) conidia as seen using SEM. Bars, 10 μm (d–h) and 5 μm (i).
Brasil. Isolates ITEM 4544 and ITEM 6139 produce large white sclerotia.

ACKNOWLEDGEMENTS

We thank Gaetano Stea from the Institute of Sciences of Food Production, CNR, Bari, Italy for his valuable technical assistance and Armando Venancio from Centro de Engenharia Biologica, Universidade do Minho, Braga, Portugal for providing the Portugal strains. This research was supported in part by a grant from the Italian Ministry of Education, University and Research (MIUR) Project 12818 – SIVINA 'Individuazione di metodologie innovative trasferibili per migliorare la sicurezza dei vini rossi di qualità del Salento'.

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