**INTRODUCTION**

The genus *Pseudoalteromonas* includes both pigmented and non-pigmented, heterotrophic bacteria, cells of which are Gram-negative rods, motile by single polar flagella. The genus was proposed by Gauthier et al. (1995), who divided *Alteromonas* into two genera, *Alteromonas* and *Pseudoalteromonas*, based on 16S rDNA sequences. Species in the genus *Pseudoalteromonas*, typically associated with eukaryotic hosts (Holmström & Kjelleberg, 1999), are frequently isolated from marine animals such as sponges (Ivanova et al., 2002b), tunicates (Holmström et al., 1998) and mussels (Ivanova et al., 1996, 1998) and also from marine algae such as *Ulva lactuca* (Egan et al., 2001) and *Laminaria japonica* (Sawabe et al., 1998). Several species in the genus, such as *Pseudoalteromonas luteoviolacea* (Gauthier, 1982; Novick & Tyler, 1985), *Pseudoalteromonas rubra* (Gauthier, 1976), *Pseudoalteromonas citrae* (Gauthier, 1977), *Pseudoalteromonas aurantia* (Gauthier & Brittmayer, 1979) and *Pseudoalteromonas peptidolytica* (Venkateswaran & Dohmoto, 2000), were isolated from sea water. Members of *Pseudoalteromonas* are also known to be producers of bioactive substances (Holmström & Kjelleberg, 1999), *P. luteoviolacea* (Gauthier, 1982; Hanefeld et al., 1994), *P. rubra* (Gauthier, 1976), *P. aurantia* (Gauthier & Brittmayer, 1979), *Pseudoalteromonas rutherenia* (Ivanova et al., 2002a) and *Pseudoalteromonas marinalis* (Ivanova et al., 2002b) have been determined as antibiotic-producing bacteria. Recently, we isolated a novel species in the genus *Pseudoalteromonas* that produces anti-methicillin-resistant *Staphylococcus aureus* (MRSA) substances.

**Pseudoalteromonas phenolica** sp. nov., a novel marine bacterium that produces phenolic anti-methicillin-resistant *Staphylococcus aureus* substances

Alim Isnansetyo and Yuto Kamei

Marine and Highland Bioscience Center, Saga University, 152-1 Shonan-cho, Karatsu, Saga 847-0021, Japan

Four strains of aerobic, Gram-negative rods, motile by means of a single polar flagellum, that produced phenolic anti-methicillin-resistant *Staphylococcus aureus* (MRSA) substances and brown-pigmented colonies, were isolated from sea water. The G+C content of the DNA ranged from 39-9 to 40-6 mol%. The isolates grew at 18–37°C and pH 6.5–9.5 (optimal pH 7.5–9) and in medium containing 1–5 % (w/v) NaCl (optimal NaCl concentration 2–3.5 %). The isolates grew optimally in medium dissolved in 40–100 % artificial sea water. Based on 16S rDNA similarities, the novel strains were closely related to *Pseudoalteromonas luteoviolacea* and *Pseudoalteromonas piscicida*, with 96-3 and 95-7 % sequence similarity, respectively. However, the strains could be differentiated from *P. luteoviolacea* by seven traits and from *P. piscicida* by 10 traits. Analysis of DNA–DNA relatedness to these related species revealed low levels of DNA hybridization (19-6 % to *P. luteoviolacea* and 22.4 % to *P. piscicida*). However, the type strain, O-BC30T, and the other three bacterial isolates showed high DNA relatedness to each other, ranging from 84-8 to 93-7 %. Based on the results of phenotypic characterization, phylogenetic analysis based on 16S rDNA sequences and DNA–DNA hybridization, it is concluded that these isolates represent a novel species in the genus *Pseudoalteromonas*. Because the type strain, O-BC30T (=IAM 14989T =KCTC 12086T), produces phenolic anti-MRSA substances, the name proposed for this novel species is *Pseudoalteromonas phenolica* sp. nov.
et al. (Archer & Bosilevac, 2001; Hiramatsu et al., 2001; Isnansetyo et al., 2001; Kaatz et al., 2000; Witte, 1999). In an effort to discover alternative antibiotics against MRSA, we screened bacteria that originated from the marine environment for anti-MRSA activity. Four strains of brown-pigmented marine bacteria with potent anti-MRSA activity were isolated independently from sea water collected from the same location and at the same sampling time. These isolates were then identified by phenotypic and genotypic characterization to represent a novel species in the genus *Pseudoalteromonas*. This paper describes this novel bacterial species and proposes the name *Pseudoalteromonas phenolica* sp. nov.

**METHODS**

**Bacterial strains and growth conditions.** The bacterial strains were isolated on 30 July 1999. The temperature and salinity of the sea water at the sampling site were 27 °C and 38%. The samples were obtained with sterile polypropylene bottles and transported to our laboratory in a cool-box. The sample was smeared on ZoBell 2216E agar medium containing (l-1) 5 g polypepton (Nihon Seiyaku) and 1 g yeast extract (Nihon Seiyaku) dissolved in 75% Jamarine S artificial sea water (ASW) (Jamarine Laboratory) (Sako et al., 1996a, b). The plates were incubated at 25 °C for 24 h. Bacterial colonies on the plate were then overlaid with MRSA suspension and then observed for 24 h. When antagonistic activity of colonies was observed against MRSA, these active colonies were isolated on ZoBell 2216E agar medium. Active isolates designated O-BC20, O-BC30(a), O-BC40(b) and O-BCX3 were isolated and are described in this paper. The reference strains *P. luteoviolacea IAM 14710T*, *Pseudoalteromonas piscicida IAM 12932T*, *Pseudoalteromonas haloplanktis* subsp. *haloplanktis IAM 12915T* and *Alteromonas macleodii IAM 12920T* were provided by the IAM Culture Collection (University of Tokyo, Japan). All of these strains were routinely cultivated aerobically at 25 °C on ZoBell 2216E agar medium.

**Physiological and biochemical characterization.** Morphological observation and phenotypic tests including catalase, cytochrome oxidase, chitinase, aagarase, arginine dihydrolase, cellulase, gelatinase, lecithanase, DNase and lipase activities were carried out based on the methods described by Smibert & Krieg (1994). Oxidative or fermentative metabolism of glucose was determined by the method of Hugh & Leifson (1953). The requirement for NaCl was tested in a medium containing (l-1) 5 g MgCl2, 2 g MgSO4, 0.5 g CaCl2, 1 g KCl, 5 g peptone (Sigma-Aldrich) and various concentrations of NaCl at pH 7-5, adjusted with KOH (Smibert & Krieg, 1994). Growth in various concentrations of ASW was examined in ASW formula 2 (Smibert & Krieg, 1994) supplemented with 5 g peptone l-1, pH 7-5. Determination of substrate utilization as single carbon and energy sources was performed in marine minimal medium (Östling et al., 1991; Holmström et al., 1998) containing 1% (w/v) substrate of interest. Growth under anaerobic conditions was tested in an anaerobic jar system (GasPak system; BBL).

**Electron microscopy.** For observation of cell morphology and flagellation, bacterial cells grown overnight on ZoBell 2216E agar were suspended in physiological saline solution. One drop of the suspension was negatively stained with one drop of 1% phosphotungstic acid after the suspension was placed on a carbon-coated copper grid. Observation was carried out under a transmission electron microscope (model H-800; Hitachi).

**Molecular characterization**

**DNA base composition.** DNA preparation was carried out based on the procedure of Marmur (1961), with minor modifications. To extract and deproteinize the DNA, phenol/chloroform/isoamyl alcohol (25:24:1) was used instead of chloroform/isoamyl alcohol (24:1) used in the standard procedure. The G+C content of DNA was determined by the HPLC method, as described previously (Tamaoka & Komagata, 1984). The G+C content of DNA from *Escherichia coli* was used as a reference.

**Amplification of 16S rDNA and sequencing.** Bacterial DNA was extracted from cells by using the EZNA Bacterial DNA kit (Omega Biotek) as recommended by the manufacturer. PCR was carried out to amplify the 16S rDNA with a Perkin-Elmer model GeneAmp PCR System 2400 thermal cycler (Applied Biosystems) with universal primers 27f and 1492r corresponding to positions 27 in the forward direction and 1492 in the reverse direction of the *E. coli* 16S rDNA sequence (Brosius et al., 1978). The amplification was performed using the PCR conditions described by Sawabe et al. (1998). The thermal profile consisted of 30 cycles of denaturation at 94 °C for 180 s, annealing at 55 °C for 60 s and extension at 72 °C for 90 s. The PCR product was purified by polyethylene glycol (PEG 6000) precipitation and used as the template for sequencing.

The ABI PRISM Dye Terminator cycle sequencing kit (Applied Biosystems) was used following the protocols described by the manufacturer. The PCR cycle-sequencing product was purified by using CENTRI-SEP columns (Princeton Separations) according to the procedure described by the manufacturer. 16S rDNA sequences were determined with a Perkin-Elmer model ABI PRISM 310 Genetic Analyzer (Applied Biosystems).

**Phylogenetic analysis and alignment.** A similarity search with the 16S rDNA sequence was performed with 16S rDNA sequences available in the GenBank/EMBL/DDBJ databases using the BLAST algorithm (Altschul et al., 1990). Multiple sequence alignment of representative sequences was carried out by using the CLUSTAL W program, version 1.8 (Thompson et al., 1994). A neighbour-joining analysis (Saitou & Nei, 1987) and bootstrap analysis of 1000 data resamplings were performed to determine the robustness of each topology. A phylogenetic tree was made by using the program TREEVIEW (Page, 1996). 16S rDNA sequence similarity values to representative 16S rDNA sequences retrieved from GenBank were calculated by the computer program GENEFYX (Software Development).

**DNA–DNA hybridization.** DNA–DNA hybridization was carried out in microplate wells (Black Maxisorp; Nunc) using a fluorimetric method (Ezaki et al., 1989; Willems et al., 2001). The fluorescence intensity was detected by a fluorescence multi-well plate reader (Cytolfluor II; PerSeptive Biosystems). The two most closely related strains, *P. luteoviolacea IAM 14710T* and *P. piscicida IAM 12932T*, a non-pigmented *Pseudoalteromonas* strain, *P. haloplanktis* subsp. *haloplanktis IAM 12915T*, *A. macleodii IAM 12920T* and *E. coli* XL-1 Blue (Stratagene) were used as reference strains.

**RESULTS AND DISCUSSION**

This paper proposes an anti-MRSA substance-producing bacterium isolated from the marine environment as a novel species in the genus *Pseudoalteromonas*. Strain O-BC30(a) and the other three strains were Gram-negative rods, motile by a single polar flagellum. These bacteria were aerobic and non-fermentative, utilized glucose oxidatively, hydrolysed starch and did not produce H2S from thiosulfate. DNase,
The 16S rDNA sequence (1450 bases) of strain O-BC30T was closest to P. luteoviolacea, P. piscicida and P. rubra (formerly Alteromonas luteoviolacea, Pseudomonas piscicida and Alteromonas rubra; Gauthier et al., 1995) with 96·3, 95·7 and 95·7 % sequence similarity, respectively. These sequence similarity data are consistent with the phylogenetic analysis (Fig. 1), which grouped these species in the same clade. Strain O-BC30T exhibited 95·1, 95·0 and 94·2 % 16S rDNA sequence similarity, respectively, to the other three members of this clade, Pseudoalteromonas flaviultra, P. maricaloris and P. peptidolytica.

The 16S rDNA sequence similarity between P. luteoviolacea and P. piscicida and strain O-BC30T is less than 97·0 %, implying that these strains will not reassociate with more than 60 % DNA–DNA relatedness (Rosselló-Mora & Amann, 2001; Stackebrandt & Goebel, 1994). However, the 97 % threshold level of 16S rDNA similarity can not differentiate the genus Pseudoalteromonas to the species level (Venkateswaran & Dohmoto, 2000).

The DNA relatedness between O-BC30T and the type strains

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pigmentation</td>
<td>Brown</td>
<td>Purple</td>
<td>–</td>
<td>–</td>
<td>Red</td>
<td>Yellow</td>
<td>Orange</td>
<td>Yellow</td>
</tr>
<tr>
<td>Growth at 37 °C</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>+</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Catalase</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Utilization of:</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>+</td>
<td>–</td>
</tr>
<tr>
<td>Maltose</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>+</td>
<td>–</td>
</tr>
<tr>
<td>D-Mannose</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Sacrose</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>+</td>
</tr>
<tr>
<td>L-Galactose</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>ND</td>
<td>ND</td>
<td>–</td>
</tr>
<tr>
<td>D-Arabinoose</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>ND</td>
<td>ND</td>
<td>–</td>
<td>+</td>
</tr>
<tr>
<td>Melibiose</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>ND</td>
<td>ND</td>
<td>–</td>
<td>+</td>
</tr>
<tr>
<td>D-Mannitol</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>+</td>
</tr>
<tr>
<td>D-Sorbitol</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>ND</td>
<td>–</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Glycerol</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>+</td>
</tr>
<tr>
<td>N-Acetylglucosamine</td>
<td>+</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>ND</td>
</tr>
<tr>
<td>Citrate</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>+</td>
</tr>
<tr>
<td>Trehalose</td>
<td>+</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>ND</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Glutamate</td>
<td>d</td>
<td>–</td>
<td>+</td>
<td>+</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>DL-Malate</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>+</td>
<td>ND</td>
<td>–</td>
<td>–</td>
<td>+</td>
</tr>
<tr>
<td>Fumarate</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Acetate</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>ND</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Pyruvate</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>ND</td>
<td>+</td>
</tr>
<tr>
<td>L-Tyrosine</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>ND</td>
<td>ND</td>
<td>+</td>
<td>ND</td>
</tr>
<tr>
<td>Tween 20</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>–</td>
<td>ND</td>
<td>ND</td>
<td>–</td>
</tr>
<tr>
<td>G+C content of DNA (mol%)</td>
<td>39·9–40·6</td>
<td>40–43</td>
<td>43–46</td>
<td>41–45</td>
<td>46–48</td>
<td>41–42</td>
<td>41·7 ± 0·4</td>
<td>38·9 ± 0·4</td>
</tr>
</tbody>
</table>

Table 1. Differential characteristics between Pseudoalteromonas phenolica sp. nov. and closely related species

Strain/species: 1, P. phenolica sp. nov.; 2, P. luteoviolacea IAM 14710T; 3, P. piscicida IAM 12932T; 4, P. haloplanktis subsp. haloplanktis IAM 12915T; 5, P. rubra; 6, P. peptidolytica MBICC F1250A1T; 7, P. flavipulchra NCIMB 2033T; 8, P. maricaloris KMM 636T. All strains studied utilized D-glucose and Tween 80 but not L-arabinose, adonitol, lactose, L-rhamnose, D-ribose, D-xylene, myo-inositol, erythritol, salicin, z-D-galacturonate, gluconate, D-glucuronate, 2-oxoglutarate, sarcosine, L-ascorbate, urea, ethanol, propanol, histidine, L-leucine, L-tyrosine, L-threonine, propionate, DL-lactate, aconitate or glycine. Data were taken from this study and from Gauthier (1976, 1982), Baumann et al. (1984), Ivanova et al. (2002b), Novick & Tyler (1985) and Venkateswaran & Dohmoto (2000). +, Positive; −, negative; d, 11–89 % of strains positive, ND, no data available.

cytochrome oxidase, lipase, lecithinase and gelatinase were positive, but catalase, chitinase, agarase, arginine dihydro-
lase and cellulase were negative. Growth factors were not
required for the growth. The G+C content of the DNA
ranged from 39·8 to 40·6 mol% (Table 1).

All of the strains grew well at 25–37 °C and slowly at 18 °C, but did not grow at 5 or 45 °C. The strains grew at pHs
ranging from 6·5 to 9·5, with optimum growth at pH 7·5–9. Growth was not seen on TCBS agar medium (Holt & Krieg,
1994), but the bacteria grew well on Marine agar 2216
(Difco) and ZoBell 2216E agar. The strains grew optimally
in ZoBell 2216E broth dissolved in 40–100 % ASW and
slowly in 20–30 % ASW, but did not grow in medium
containing 12·5 % ASW. Growth occurred in medium
containing NaCl at concentrations of 1 to 5 % (optimal
concentration 2–3·5 %).

The 16S rDNA sequence (1450 bases) of strain O-BC30T
was aligned by comparison with available sequences from
GenBank/EMBL/DDBJ. The result revealed that strain

http://ij.sgmjournals.org

P. phenolica sp. nov. produces anti-MRSA substances

585
of the most closely related species, P. luteoviolacea IAM 14710^T and P. piscicida IAM 12932^T, was respectively 19.9 and 22.4%. DNA–DNA hybridization was also performed between O-BC30^T and non-pigmented P. luteoviolacea IAM 12915^T and E. coli XL-1 Blue, resulting in DNA–DNA relatedness of 14.1, 11.9 and 14.1%. In addition, the DNA relatedness values between the other three bacterial isolates [O-BC20, O-BC40(B) and O-BCX3] and the reference strains were also very low, ranging from 3.7 to 27.3%. All of the DNA relatedness values were less than 70%, suggesting that all strains studied and reference strains could not be placed within the same species. In contrast, the DNA relatedness between strain O-BC30^T and the other three novel strains ranged from 84.8 to 93.7%, suggesting that these four strains should be included in the same species. Phylogenetic analysis based on 16S rDNA sequences and DNA–DNA hybridization therefore reveals that these four strains of the anti-MRSA substance-producing marine bacteria represent a novel species in the genus Pseudoalteromonas.

The most closely related species, P. luteoviolacea, is reported to be an antibiotic-producing bacterium (Baumann et al., 1984; McCarthy et al., 1985; Hanefeld et al., 1994). However, the nature and antibacterial spectra of the antibiotics produced by strain O-BC30^T are different from those produced by P. luteoviolacea. Strain O-BC30^T produces phenolic antibiotics, which are only active against Gram-positive bacteria. However, P. luteoviolacea produces antibiotics against both Gram-positive and -negative bacteria. Because strain O-BC30^T produces phenolic anti-MRSA substances, the name proposed for this novel species is Pseudoalteromonas phenolica sp. nov.

**Description of Pseudoalteromonas phenolica sp. nov.**

Pseudoalteromonas phenolica (phe.no’li.ca. N.L. n. phenolum phenol; N.L. fem. adj.phenolica related to phenol).

Cells are Gram-negative rods (0.6–0.9×1.5–2.8 μm), motile by means of a single polar flagellum; colonies are convex, circular and brown. Catalase-, chitinase-, agase-, arginine dihydrolase- and cellulase-negative. DNase-, cytochrome oxidase-, lipase-, lecithinase- and gelatinase-positive. The DNA G+C content is 39.9–40.6 mol%. Growth is observed at 18–37°C, at pH 6.5–9.5 (optimum pH 7.5–9) and in medium containing 1–5% (w/v) NaCl (optimum concentration 2–3.5%). Decomposes starch. Does not produce H₂S from thiosulfate. Utilizes D-glucose, Tween 80, Tween 20, maltose, D-mannose, sucrose, laevulose, N-acetylglucosamine, trehalose, glutamate, fumarate and acetate. Does not utilize L-arabinose, D-arabinose, adonitol, D-galactose, lactose, D-mannitol, D-galacturonate, D-glucuronate, 2-oxoglutarate, DL-malate, sarcoine, D-ribose, D-xylose, D-sorbitol, myo-inositol, erythritol, glycerol, salicin, z-D-galacturonate, gluconate, D-glucuronate, 2-oxoglutarate, DL-malate, sarcosine, L-ascorbate, urea, ethanol, propanol, histidine, L-leucine, L-tyrosine, L-threonine, propionate, DL-lactate, aconitate, pyruvate, glycine, citrate or DL-malate. Produces phenolic antibiotics.

The type strain, strain O-BC30^T (= IAM 14989^T = KCTC...
was isolated from sea water collected off Ogasawara Island, Tokyo, Japan.

REFERENCES


