Haloterrigena thermotolerans sp. nov., a halophilic archaeon from Puerto Rico

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An extremely halophilic Archaeon belonging to the order Halobacterales was isolated from the solar salterns of Cabo Rojo, Puerto Rico. The organism, designated strain PR5T, is rod-shaped, non-motile and requires at least 12 % (w/v) NaCl to grow. The strain is highly thermotolerant: its temperature optimum is 50 °C and growth is possible up to 60 °C. Polar lipid analysis revealed the presence of the bis-sulfated glycolipid S2-DGD-1 as sole glycolipid and the absence of the glycerol diether analogue of phosphatidyglycerosulfate. Both C20,C20 and C20,C25 core lipids are present. The G+C content of the DNA is 63.3 mol %. According to 16S rDNA sequence data, strain PR5T is closely related to the representatives of the genera Haloterrigena and Natrinema, but on the basis of its phenotypic properties, 16S rDNA sequence and DNA–DNA hybridization studies, strain PR5T cannot be assigned to any of the recognized species within these genera. On the basis of its polar lipid composition, the isolate has been assigned to the genus Haloterrigena. The creation of a new species, Haloterrigena thermotolerans, is therefore proposed to accommodate this isolate. The type strain is strain PR5T (= DSM 11552T = ATCC 700275T).

INTRODUCTION

Halophilic Archaea belonging to the order Halobacterales are found in large numbers in the crystallizer ponds of solar salterns worldwide. Their massive populations often impart a red colour to the brines (Oren, 1993, 1994). A variety of species of red halophiles have been isolated from saltern ponds, including Halobacterium salinarum, Halorubrum saccharovorum, Haloferax mediterranei, Haloferax gibbonsii, Haloferax denitrificans, Haloarcula hispanica, Haloarcula japonica, Halococcus morrhuae and Halococcus saccharolyticus. However, the dominant type of halophilic Archaea inhabiting such ponds still defies microbiologists’ attempts toward its isolation (Benloch et al., 1995; Oren et al., 1996).

In 1994, 20 halophilic archaeal strains were isolated from the solar salterns of Cabo Rojo, Puerto Rico. Some of these isolated strains could be classified in the genera Halobacterium and Haloferax on the basis of cell morphology and polar lipid composition (Montalvo-Rodriguez, 1996). Three isolates (designated PR3T, PR7 and PR19), while being similar to each other, did not resemble any of the hitherto described taxa within the Halobacteriales. A new genus and species were created to accommodate these strains, Halogeometricum borinquense, with strain PR3T (=

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Abbreviations: PG, phosphatidylglycerol; PGP-Me, phosphatidylglycerolphosphate methyl ester; PGS, phosphatidylglycerosulfate.
The GenBank accession number for the 16S rDNA gene sequence of Haloterrigena thermotolerans strain PR5T is AF115478.
ATCC 700274<sup>T</sup> as type strain (Montalvo-Rodríguez et al., 1998).

One isolate, designated PR<sub>5</sub><sup>T</sup>, consisted of long non-motile rods, and contained the bis-sulfated glycolipid S<sub>3</sub>-DGD-1 as sole glycolipid. In this paper, the characteristics of strain PR<sub>5</sub><sup>T</sup>, which appeared to be sufficiently different from the other recognized species of halophilic Archaea to be described as a new species, are presented.

**METHODS**

Collection of samples and isolation of halophilic Archaea. Water samples (400 ml portions) were collected in sterile plastic bags (Whirl-Pak) from the solar salterns in Cabo Rojo, Puerto Rico, in June 1994. Each sample was divided in 50 ml aliquots, which were then filtered through 0.45 µm sterile nitrocellulose membranes (Millipore). The membranes were transferred onto agar plates containing a modified Sehgal–Gibbons (SG) medium (Sehgal & Gibbons, 1960). This medium contained (g l<sup>−1</sup>): NaCl, 250; MgSO<sub>4</sub>.7H<sub>2</sub>O, 20; KCl, 2; sodium citrate (trisodium salt), 3; Casamino acids, 7.5; yeast extract, 1.0; and FeSO<sub>4</sub>.7H<sub>2</sub>O, 0.0023. For solid media, 20 g agar l<sup>−1</sup> was added. Plates were incubated in sealed plastic bags at 40°C. After 7 d incubation, representative colonies were transferred to SG broth medium and isolated in pure culture.

The strains *Haloterrigena turkmenica* VKM B-1734<sup>T</sup>, *Halobacterium salinarum* NRC 817, *Halofexar mediterranei* ATCC 33500<sup>T</sup>, *Halorubrum saccharovorum* ATCC 29252<sup>T</sup>, *Halococcus hispanica* ATCC 33960<sup>T</sup>, *Halococcus marismortui* ATCC 43049<sup>T</sup>, *Halomicromonas vallismortis* ATCC 29715<sup>T</sup> and *Natrialba asiatica* JCM 9576<sup>T</sup> were used as reference strains. Cultures of *Natrienema pellirubrum* NCIMB 786<sup>T</sup> and *Natrienema pallidum* NCIMB 7777<sup>T</sup> were obtained as a gift from W. D. Grant (Leicester, UK).

**Morphological, cultural and physiological characteristics.** In most experiments described below, SG medium was used for growth of strain PR<sub>5</sub><sup>T</sup>, modified by lowering the NaCl concentration to 200 g l<sup>−1</sup> and decreasing the yeast extract concentration to 0.5 g l<sup>−1</sup>. Cultures were examined for motility and morphological features in wet mounts. Optimal conditions for growth were determined by growing cultures on a rotary shaker (120 r.p.m.) in 40 ml portions of medium in 100 ml Erlenmeyer flasks, and changing concentrations of medium components and temperature as indicated. Unless otherwise indicated, growth temperature was 35°C. The extent of growth was assessed by measuring the turbidity of the cultures at 600 nm. The pH tolerance was tested in modified SG medium buffered with 20 mM MES (pH 5.0, 5.5), PIPES (pH 6.0, 6.5), Tricine (pH 7.0, 7.5, 8.0) or TAPS (pH 8.0, 8.5). For growth experiments in buffered medium above pH 7.5, the Mg<sup>2+</sup> concentration of the medium was reduced to 20 mM to avoid the formation of massive precipitates. Gram stains were prepared as described by Dussault (1955).

Antibiotic sensitivity was tested by spreading bacterial suspensions on plates of SG medium and applying antibiotic discs (penicillin, 10 µg; bacitracin, 10 µg; novobiocin, 30 µg; kanamycin, 30 µg; ampicillin, 30 µg; vancomycin, 30 µg; sulfamethazole and trimethoprim, 25 µg each; and chloramphenicol, 30 µg) (Bonelo et al., 1984; Colwell et al., 1979; Torreblanca et al., 1986). The results were recorded as sensitive or resistant after 14 d incubation at 40°C, with sensitive being defined as the appearance of a zone of inhibition extending at least 2 mm beyond the antibiotic disc. Sensitivity to neomycin and anisomycin was tested in liquid cultures, to which the antibiotics were added to a concentration of 25 µg ml<sup>−1</sup>.

**Lipid analyses.** Polar lipid extraction and characterization by TLC on silica gel plates (20 × 20 cm; Sigma) was performed as described by Torreblanca et al. (1986), using single development in chloroform/methanol/acetic acid/water (85:22:5:10:4, v/v). Plates were stained for glycolipids and other lipids using the x-naphthol stain, the CeSO<sub>4</sub>/H<sub>2</sub>SO<sub>4</sub> stain, the orcinol/FeCl<sub>3</sub> stain and the molybdate stain for detection of phospholipids (Kates, 1972; Oren et al., 1996; Torreblanca et al., 1986). The presence of C<sub>30</sub>C<sub>20</sub> and C<sub>23</sub>C<sub>6</sub> core lipids was assessed by hydrolysing lyophilized cell material overnight at 30°C in methanol/toluene/sulfuric acid (5:5:0.2, v/v), extracting the core lipids with hexane, followed by chromatography on silica gel plates, using hexane/diethylther (4:1, v/v) as eluent (Tindall, 1985). Spots were visualized with iodine vapours. Carotenoid pigments were extracted with methanol/aceton (1:1, v/v) and absorption spectra were recorded in a Hewlett Packard model 8452A diode array spectrophotometer.

**Biochemical tests.** Unless specified otherwise, tests for phenotypic properties were carried out as indicated in the proposed minimal standards for the description of new taxa in the order *Halobacteria* (Oren et al., 1997). Nitrate reduction was tested in liquid SG medium supplemented with 0.5% NaNO<sub>3</sub>. Formation of gaseous products from nitrate was detected by the presence of gas bubbles in Durham tubes. Anaerobic growth in the presence of either nitrate or arginine was tested as described previously (Hartmann et al., 1980; Mancinelli & Hochstein, 1986). Production of acids from sugars was examined in SG medium supplemented with 0.5% of the sugars tested. After 2 d, the pH was measured with a pH electrode. To determine the ability to use sugars as energy sources, the yeast extract and Casamino acids concentration in the SG medium were reduced to 0.025% each, and the medium was amended with 0.05% NH<sub>4</sub>Cl and 0.5% of the sugar to be tested. After 3 d incubation at 35°C, the OD of the cultures was compared with that of the control culture without added sugar.

Indole production was detected with Kovac’s reagent after having grown the cells in modified SG medium amended with 0.25% tryptone. To determine starch hydrolysis, the strains were streaked onto modified SG plates with 0.2% soluble starch solidified with 2% (w/v) agar. Starch hydrolysis was detected by flooding the plates with iodine solution after the colonies were fully grown. Presence of catalase was determined by adding 1% (v/v) H<sub>2</sub>O<sub>2</sub> solution to colonies on modified SG agar medium. Gelatin hydrolysis was determined by growing colonies on agar plates amended with 0.4% gelatin and flooding the plates with a solution of 15% (w/v) HgCl<sub>2</sub> in 20% (w/v) HCl after growth was obtained (Gutiérrez & González, 1972). Tween 80 hydrolysis was tested as outlined by Gutiérrez & González (1972). The presence of oxidase was determined with tetramethyl-p-phenylenediamine. 2HCl (Holding & Collee, 1971). Appropriate positive and negative controls were run for all the above tests.

**Phase-contrast and electron microscopy.** Cells were examined for motility using a Zeiss standard microscope equipped with phase-contrast optics. For photography, drops of culture were mixed on a microscope slide with an equal volume of melted 2% (w/v) agar containing 20% (w/v) NaCl and covered with a cover-slip. For EM examination, 4 µl drops of culture were applied to carbon-coated...
collodion-stabilized copper grid for 30 s. The drops were washed away with 4–5 drops of 1% (w/v) aqueous uranyl acetate and the grids were allowed to air-dry. Specimens were examined in a JEOL-JEM model 100CX TEM operating at 80 kV.

**DNA base composition.** The G+C content of the DNA was determined by the Deutsche Sammlung von Mikroorganismen und Zellkulturen (DSMZ), Braunschweig, Germany. The DNA for the G+C content was isolated and purified by chromatography on hydroxyapatite. The G+C content was determined by HPLC.

**16S rDNA characterization.** PCR-mediated amplification of the 16S rDNA and purification of the PCR products was carried out as described previously (Barns et al., 1994; Rainey et al., 1996). Purified PCR products were sequenced at the DSMZ, using the ABI PRISM Dye Terminator Cycle Sequencing Ready Reaction kit (Applied Biosystems) as directed in the manufacturer’s protocol. Sequence reactions were electrophoresed using the Applied Biosystems 373A DNA Sequencer. A total of 1451 bases were determined by EM after negative staining with uranyl acetate.

**Preparation of labelled DNA and DNA–DNA hybridization experiments.** DNA was labelled by the multiprime system with a commercial kit (RPN 1601 Y; Amersham) with deoxy[1,2,5-3H]cytidine 5’-triphosphate (Amersham). The mean specific activity obtained with this procedure was 8.4 x 10^6 c.p.m. µg^-1 DNA. The labelled DNA was denatured prior to hybridization by heating at 100 °C for 5 min and then placed on ice.

DNA–DNA hybridization studies were performed by the competition procedure of the membrane method described by Johnson (1981). Competitor DNAs were sonicated in a Braun sonifier at 50 W for two 15 s time intervals. Membrane filters (HAHY; Millipore) containing reference DNA were washed in 2 M NaCl, 2 M NaOH at the optimal renaturation temperature (56 °C). The radioactivity bound to the filters was measured in a liquid scintillation counter (Beckman) and the percentage of homology was calculated according to Johnson (1981). Two independent determinations were carried out for each experiment.

**RESULTS**

Strain PR5^T^ was isolated from a saltern crystallizer pond in Cabo Rojo, Puerto Rico, using the same set of samples from which Halogloeocapsa borainguense was obtained (Montalvo-Rodriguez et al., 1998). The bacterium is a non-motile, slender rod, measuring 4–13 x 0.7–1.0 µm (Fig. 1). No flagella were observed by EM after negative staining with uranyl acetate. After fixation with acetic acid (Dussault, 1955), the cells stained Gram-negative. Colonies on agar are pale red, 0.5–1.0 mm in diameter, opaque, circular and convex with an entire margin. Absorption spectra of cell extracts in methanol/acetone (1:1, v/v) showed maxima at 496 nm and 528 nm, with a shoulder at 474 nm, typical of α-bacteriourfberin and derivatives characteristically found in most members of the Halobacteria.

Strain PR5^T^ required at least 2 M NaCl for growth. Optimum growth was obtained between 3.0 and 3.5 M NaCl. At 4.3 M NaCl, growth was significantly slower than at 3.5 M. Cells lysed when suspended in media with NaCl concentrations below 1 M. In contrast to most neutrophilic members of the Halobacteria, strain PR5^T^ showed only a low requirement for Mg^{2+} ions. Magnesium concentrations as low as 5 mM were sufficient for optimal growth and the rod-shaped morphology was maintained at suboptimal Mg^{2+} concentrations. The pH range for growth was 6.5–8.2, with an optimum between 7.0 and 7.5. In the standard medium used (modified SG broth), the isolate grew optimally at 50 °C with a doubling time of 6 h. Growth was obtained at temperatures as high as 60 °C, but not at 62 °C. At 60 °C, most cells were highly elongated, with lengths of 20–25 µm and higher, suggesting that cell division was inhibited at this high temperature.

Isolate PR5^T^ is chemo-organotrophic and strictly aerobic. Oxidase and catalase reactions were positive. Casamino acids were an excellent source of carbon, nitrogen and energy. Tests with different sugars and related compounds (glucose, sucrose, fructose, mannose, maltose, galactose, lactose, raffinose, cellobiose, trehalose, arabinose, ribose, xylose, mannitol, dulcitol, glycerol) showed that none of these stimulated growth, either when added to the standard growth medium, or when used at a concentration of 5 g l^-1 in medium in which the concentrations of Casamino acids and yeast extract were reduced to 0.25 g l^-1 each. Production of acids from sugars was never observed. Amylase was not produced. No anaerobic growth was obtained in the presence of L-arginine or nitrate. Some nitrite was produced from nitrate in aerobic cultures. Gas formation on nitrate was never observed. Tween 80 was hydrolysed. Weak gelatinase activity was detected only after prolonged (2–3 weeks) incubation; casein was not hydrolysed. Indole was not produced.

The isolate proved sensitive to anisomycin, bacitracin, novobiocin, vibriostatic agent (O/129) and sulfamethazine plus trimethoprim. No inhibition was observed by ampicillin, penicillin, chloramphenicol, erythromycin, neomycin, streptomycin, tetracycline, kanamycin or vancomycin.

Strain PR5^T^ possessed both C_{20}, C_{20} and C_{20}, C_{20} diether core lipids. Polar lipids detected were phosphatidylglycerol (PG), phosphatidylglycerolphosphate methyl ester (PGP-Me) and a single glycolipid, chromatographically identical to S2-DGD-1 of Natrialba asiatica (Kamekura & Dyall-Smith, 1995; Matsubara et al., 1994). Phosphatidylglycerosulfate (PGS) was absent.
or below the detection limit. A minor, as yet unidentified lipid, running somewhat slower than PGS, was sometimes found.

The G+C content of the DNA of strain PR5T was 63.3 mol% (HPLC). Comparison of the 16S rDNA sequence (1451 bases determined, with 16 bases missing at the 5′ side and 6 bases at the 3′ end; GenBank accession number AF115478) with published sequences of other members of the Halobacteriales shows strain PR5T to be related to the genera Haloterrigena (Ventosa et al., 1999) and Natrinema (McGenity et al., 1998) (Fig. 2).

The phylogenetic distance between strain PR5T and the type strains of its closest relatives (Fig. 2) and the differences in phenotypic properties (Table 1) all suggest that PR5T is sufficiently different from the recognized species within the Haloterrigena–Natrinema group to warrant its classification as a new species. On the basis of its polar lipid composition it is proposed that strain PR5T be assigned to the genus Haloterrigena. Its DNA relatedness with Haloterrigena turkmenica, the only species of the genus described thus far, is sufficiently low (48% hybridization of Haloterrigena turkmenica DNA with 3H-labelled DNA from strain PR5T) and phenotypic properties are sufficiently different to justify the creation of a new species.

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**Fig. 1.** Strain PR5T, as viewed by phase-contrast microscopy. Bar, 10 µm.

**Fig. 2.** Phylogenetic tree showing the relationship between isolate PR5T, species allocated to the genera Haloterrigena and Natrinema, and close relatives, based on 16S rRNA sequences. The tree was constructed using the neighbour-joining method. Bootstrap values (1000 replicates) are shown at the nodes. Bar, 0.014 expected changes per site. The outgroup is Natronobacterium gregoryi.
Table 1. Some properties differentiating *Haloterrigena thermotolerans* from the validly described species within the genera *Haloterrigena* and *Natrinema*

<table>
<thead>
<tr>
<th>Property</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Morphology</td>
<td>Rods</td>
<td>Coccoid</td>
<td>Rods*</td>
<td>Rods</td>
</tr>
<tr>
<td>Cell size (μm)</td>
<td>4–13</td>
<td>1·5–2·0</td>
<td>1–4</td>
<td>1·5–6·0</td>
</tr>
<tr>
<td>Lysis in distilled water</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Temperature range (°C)</td>
<td>25–60</td>
<td>NR</td>
<td>20–45</td>
<td>NR</td>
</tr>
<tr>
<td>Temperature optimum (°C)</td>
<td>50</td>
<td>45</td>
<td>NR</td>
<td>37–40</td>
</tr>
<tr>
<td>Use of sugars as carbon source</td>
<td>–</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Acid formation from sugars</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Gelatin liquefaction</td>
<td>Weak</td>
<td>–</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Glycolipid content</td>
<td><em>S₂</em>-DGD-1</td>
<td><em>S₂</em>-DGD-1</td>
<td>One major glycolipid, distinct from <em>S₂</em>-DGD-1, running very slowly on TLC plates</td>
<td>Several, unidentified, distinct from <em>S₂</em>-DGD-1</td>
</tr>
<tr>
<td>Presence of PGS</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>DNA G+C content (mol%)</td>
<td>63·3</td>
<td>59·2–60·2</td>
<td>69·9 (major)†, 60·0 (minor)</td>
<td>NR</td>
</tr>
</tbody>
</table>

*Natrinema pellirubrum* was reported to be rod-shaped (McGenity *et al*., 1998), but in our cultures cells are extremely pleomorphic.

† These data were presented by McGenity *et al*., 1998) as representative for the genus. However, they relate to *Natrinema pellirubrum* only.

**DISCUSSION**

The 16S rDNA sequence of strain PR5*T* clearly demonstrates its affiliation with representatives of the genera *Haloterrigena* (Ventosa *et al*., 1999) and *Natrinema* (McGenity *et al*., 1998). These genera were created independently to accommodate a number of strains formerly classified in other genera but phylogenetically unrelated to them and a few new isolates. Some strains have been assigned to both genera and they may thus overlap at least in part. Additional studies will be required to reassess the taxonomic status of the two genera. Similarity percentages between the PR5*T* sequence and the type strains of *Haloterrigena turkmenica*, *Natrinema pellirubrum* and *Natrinema pallidum* were 96·2, 96·6 and 96·2%, respectively, sufficiently high to enable classification within either of these genera, but sufficiently low to warrant the creation of a separate taxon at the species level (Devereux *et al*., 1990).

A unique feature of strain PR5*T* is its ability to grow at elevated temperatures: optimum growth was recorded at 50 °C and growth was possible up to 60 °C. The appearance of greatly elongated cells at 60 °C suggests that the first function to be damaged by the supraoptimal temperatures may be related to cell division. None of the other *Haloterrigena* or *Natrinema* strains described shows such thermophilic or thermotolerant behaviour. Within some of the other genera of the *Halobacteriaceae*, growth was recorded at quite high temperatures, and optima of up to 50 °C for growth have been reported for *Halobacterium salinarum*, *Halorubrum saccharovorum*, *Haloarcula marismortui* and *Haloferax denitrificans*. The most thermotolerant species described thus far seems to be *Haloarcula quadrata* (Oren *et al*., 1999), which grows optimally between 50 and 53 °C but does not grow above 55 °C, and *Natralba asiatica* strain B1T, which has its optimum between 35 and 40 °C but can grow up to 52 °C (Kamekura & Dyall-Smith, 1995). However, individual enzymes from halophilic Archaea may display a highly thermophilic character (Keradjopoulos & Holldorf, 1977).

Strain PR5*T* shares the presence of *S₂*-DGD-1 with...
Haloterrigena turkmenica VKM B-1734T. This glycolipid is clearly distinct from the (yet unidentified) glycolipids of Natrinema pellirubrum and Natrinema pallidum. The genus Natrinema is heterogeneous with respect to the glycolipids encountered in the different species. Such a phenomenon may occur in other genera within the Halobacteriaceae as well (Natralba, Halorubrum) (Kamekura, 1998, 1999).

Because of the affiliation of strain PR5T with the genera Haloterrigena and Natrinema and the profound differences with the validly described species within these genera, strain PR5T should be classified as a new species. On the basis of its polar lipid composition, its assignment to the genus Haloterrigena appears to be warranted and the name Haloterrigena thermotolerans is proposed in view of its ability to grow at high temperatures. For the time being, the description of the species is necessarily based on a single isolate. It is desirable to obtain additional isolates that belong to the same taxon. The property of growth at high temperatures may probably be used in the designation of specific enrichment procedures toward the isolation of other representatives of the species.

**Description of Haloterrigena thermotolerans** sp. nov.

Haloterrigena thermotolerans (ther.mo.to.le.rans. Gr. n. thermo heat; tolerans L. pres. part. of tolero tolerate; M.L. adj. thermotolerans heat-tolerant).

Cells are rod-shaped, 4–13 µm by 0.7–1.0 µm. Non-motile. Require at least 2 M NaCl for growth. Cells lyse in less than 1 M NaCl. Optimum NaCl concentration for growth is 3.0–3.5 M. Magnesium requirement low, with optimal growth and normal morphology being obtained in 5 mM Mg²⁺. Colonies are circular, entire, convex, opaque, pale red, 0.5–1.0 mm in diameter. Chemo-organotrophic, strictly aerobic. Nitrogen and carbon source: Casamino acids. Sugars (glucose, fructose, sucrose, ribose and others) and glycerol are not used as carbon sources and do not give rise to acid production. Does not produce amylase. Reduces nitrate to nitrite. Does not grow anaerobically in the presence of nitrate or l-arginine. Liquefaction of gelatin weak. Tween 80 hydrolysed. Does not produce indole. Oxidase- and catalase-positive. Possesses C₂₀, C₂₈, and C₃₁ C₈ diether core lipids. Polar lipids are PG, PG-P-Me and S₂-DGD-1 as sole or main glycolipid. PGS absent or below detection limit. Growth in the pH range 6.5–8.2; optimum 7.0–7.5. Temperature optimum 50 °C; weak growth is possible up to 60 °C. Sensitive to anisomycin, bacitracin, novobiocin, vibriostatic agent (O/129) and sulfamethazone plus trimethoprim. Insensitive to ampicillin, penicillin, chloramphenicol, erythromycin, neomycin, streptomycin, tetracycline, kanamycin and vancomycin. The G+C content of the DNA is 63.3 mol% (HPLC). Type strain: PR5T, isolated from a saltern crystallizer pond in Puerto Rico, was deposited as DSM 11552T (= ATCC 700275T).

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