‘Candidatus Xenohaliotis californiensis’, a newly described pathogen of abalone, *Haliotis* spp., along the west coast of North America

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**INTRODUCTION**

A relatively new disease, withering syndrome (WS), has resulted in catastrophic declines in populations of black abalone, *Haliotis cracherodii* Leach, in California, USA (Haaker *et al.*, 1992; Steinbeck *et al.*, 1992; VanBlaricom *et al.*, 1993; Friedman *et al.*, 1997). This chronic disease is also responsible for heavy losses of red abalone, *Haliotis rufescens* Swainson, in California, USA, and Baja California, Mexico (Haaker *et al.*, 1995; Moore *et al.*, 2000). Withering syndrome is characterized by degeneration of the digestive gland of the host (atrophy and/or metaplasia) and depletion of glycogen reserves (Kismohandaka *et al.*, 1993; Gardner *et al.*, 1995). Animals cease feeding and catabolize foot muscle protein as an energy source that results in atrophy of the pedal muscle (Kismohandaka *et al.*, 1993), and, ultimately, death. The incubation period of WS is approximately 5–7 months in black abalone (Friedman *et al.*, 1997; Friedman & Fan, 1998) and approximately 7–8 months in red abalone at 18 °C (J. D. Moore & C. S. Friedman, unpublished results).

Until recently, an aetiological agent had not been identified. Gardner *et al.* (1995) suggested that a *Rickettsiales*-like prokaryote (RLP) that infects the mucosal epithelium of the gastrointestinal tract might be the causative agent of WS. Friedman & Fan (1998) who transmitted the disease between black abalone observed a significant difference in intensity of RLP

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infection between abalone with and without WS, but no correlation between intensity of infection with RLPs and condition of the abalone in the infected treatments. In contrast, significant correlations were observed between intensity of RLP infection and degree of WS in wild and farmed red abalone (Haaker et al., 1995; Moore et al., 2000). The differences in responses to RLP infections between the two species may account for this discrepancy. Black abalone respond predominantly with degenerative changes to the digestive gland, while red abalone display a metaplasia of the mucosal epithelium in this gland. The metaplasia results in more target tissue for the bacterium compared to a loss of the same tissues in black abalone. In another study, Friedman et al. (unpublished results) found that both the RLPs and WS were responsive to treatments with oxytetracycline, providing strong evidence that the RLP is the aetiological agent of WS. Collectively, these studies indicate that the RLP in abalone is the aetiological agent of WS and that the pathogenesis of WS is complex and differs between host species. 

Riskettsia-like bacteria are commonly observed in many marine invertebrates (Sparks, 1985; Friedman et al., 1989; LeGall et al., 1988). Until recently, these intracellular bacteria were thought to be non-pathogenic commensal organisms, unlike their terrestrial relatives (Anderson et al., 1992; Brenner et al., 1993). Due to the current inability to culture RLPs that infect marine invertebrates, differentiation of these bacteria has relied upon morphological characteristics as observed by light and electron microscopy methods (Friedman et al., 1989; VanBlaricom et al., 1993; Gardner et al., 1995; Bower et al., 1996; Wu & Pan, 1999). Based on light and electron microscopy, the bacterium observed within membrane-bound cytoplasmic vacuoles of gastrointestinal epithelial cells of abalone resembled members of the tribe Ehrlichiaeae in the family Riskettsiaceae. In order to determine the relationship of the RLP in abalone to known members of the family Riskettsiaceae, we examined antigenic, physiological and life history characteristics and sequenced the 16S rDNA gene of the bacterium associated with WS in black abalone from California. The 16S rDNA has been exploited for developing primers for PCR (Andree et al., 2000) and in situ hybridization tests (Antonio et al., 2000) to detect the agent in abalone tissues.

**METHODS**

**Animals.** Adult black abalone that measured 76–127 mm were collected in July of 1996 from Vandenberg Airforce Base (VAFB), California, a location at which WS is found. Abalone free of WS that measured 51–76 mm were collected from Sobranes Point (SP), near Carmel, California in May of 1996.

**Histology.** In order to observe the morphology of RLP inclusions, selected tissues were placed in Invertebrate Davidson’s solution (Shaw & Battle, 1957) for 24 h and processed for routine paraffin histology. Deparaffinized 5 μm sections were stained with haematoxylin and eosin or Brown and Brenn’s Gram stain (Luna, 1968) and viewed by light microscopy.

**Transmission electron microscopy.** The post-oesophageal was excised from several black abalone and 1–2 mm² pieces were fixed in 4% glutaraldehyde, 0.15 M sodium cacodylate and 0.15 M sodium chloride, pH 7.3 for 2–18 h. The specimens were post-fixed in 1% OsO₄ for 1 h at 4 °C (Rosen et al., 1978). Tissues were rinsed twice in buffer, dehydrated through a graded series of acetone, infiltrated and embedded in epoxy resin. Ultrathin sections (10–20 nm) were stained with uranyl acetate and lead citrate and examined with a Zeiss 10C transmission electron microscope at 80 kV.

**Antigenic reactivity with Chlamydiaceae-specific antibodies.** The heteromorphic appearance of individual bacteria within some inclusions is suggestive of the distinct developmental stages (e.g. elemental and reticulate bodies) of *Chlamydia* and *Chlamydophila* spp., the two genera that now comprise the family Chlamydiaceae (Everett et al., 1999). Although other evidence suggested that the RLP is more closely related to members of the order Riskettsiales than to those in the order Chlamydiales, we tested the ability of two monoclonal antibodies that possess broad reactivity with species of Chlamydiaceae to recognize the RLP. The samples consisted of Invertebrate Davidson’s-preserved deparaffinized tissue sections from two black abalone and one red abalone. Avian lung tissue infected with *Chlamydia psittaci* fixed in 10% neutral buffered formalin served as a positive control. The antibody (Mab AC-1; American Research Products) recognized a genus-specific epitope of the chlamydial lipopolysaccharide antigen. The second antibody we used (BIO29-65, *Chlamydia Indirect IP KH*; Bartels) recognizes all serovars of *Chlamydia trachomatis*, *Chlamydophila (Chlamydia) psittaci* and *Chlamydophila (Chlamydia) pneumoniae*. Both antibodies were used according to the manufacturer’s protocols with indirect immunoperoxidase detection using 3-amino-9-ethylcarbazole as the chromagen. A section from each sample was incubated with normal mouse IgG in lieu of specific antibody as a negative control.

**Antimicrobial susceptibility.** Due to the inability to culture the RLP, antimicrobial susceptibility was evaluated in vivo using infected abalone. Both cultur ed red and wild black abalone infected with the RLP in the field were used as experimental animals. Abalone were administered the following antimicrobials in two trials: (1) daily intramuscular (IM) injections for 14 d: chloramphenicol (12 mg kg⁻¹, Sigma, *n* = 8–10), clarithromycin (1.96 mg kg⁻¹, Abbott Laboratories, *n* = 6–8), sarafloxacin (12.98 mg kg⁻¹, Abbott Laboratories, *n* = 6–8) and tetracycline (50 mg kg⁻¹, Sigma, *n* = 8–10). (2) In a separate trial, abalone were administered nine IM doses of oxytetracycline (21 mg kg⁻¹, Liquimycin-LA 200, Pfizer, *n* = 9–12; Table 1). Animals were sampled for histology as described above 1 d after administration of the final dose in the first trial and two weeks after the final dose in the second trial. The non-parametric Mann–Whitney Rank Sum test (SigmaStat version 2.03 1992–1997) was used to examine differences in prevalences of the bacterium between treated and untreated control groups.

**Tissue selection and DNA extraction.** A small piece of posterior oesophageus (post-oesophageus) was examined by phase-contrast microscopy to assess the degree of RLP infection in the VAFB animals and to assure that the SP abalone were RLP-free. The post-oesophageus from each abalone was individually rinsed 3 × in sterile 3% (w/v) saline/0.1% Tween 80 solution, followed by a rinse in sterile 3% saline to remove exogenous bacteria present in the
lumen of the post-oesophagus. DNA was extracted according to the methods of Sambrook et al. (1989). Briefly, rinsed tissues were homogenized in lysis/protease K buffer. After 1 h at 55 °C the DNA was extracted in a phenol/chloroform solution. Isoamyl alcohol was added, mixed for 10 min and centrifuged. The top aqueous phase was removed and 0·1 vol. 3 M sodium acetate was added. Cold 100% ethanol was added to precipitate the DNA. The pellet was washed once in 70% ethanol, air-dried and resuspended in Tris-EDTA (TE) buffer. The DNA concentration was determined by spectrophotometry.

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**PCR amplification and sequencing.** A sample containing 150 ng host and parasite DNA was used to amplify the RLP 16S rDNA sequence using EUB A and EUB B universal, eubacterial 16S rDNA primers (Gibco-BRL; Giovannoni, 1991). All the amplifications were performed in standard 50 µl reactions containing 10 mM Tris/HCl pH 8·3 (at 25 °C), 50 mM KCl, 1·5 mM MgCl2, 0·001% (w/v) gelatin, 400 µM dNTPs, 5 µM tautomethyl ammonium chloride, 40 pmol each primer, and 2 U Taq polymerase. The PCR thermal cycler used was a model PTC-100 (MJ Research). The amplification entailed 37 cycles of 1 min at 94 °C, followed by 1 min at 56 °C, followed by 1 min at 72 °C. This was preceded by a denaturation step where samples were held at 95 °C for 5 min. The thermal cycler program finished with an extended elongation step where samples were held at 72 °C for 7 min. An aliquot of the amplified DNA was separated on a 1·5% (w/v) agarose gel, stained with ethidium bromide and examined using UV illumination. The entire RLP 16S rDNA gene (1525 bases) was cloned into pCR2.1 using a TOPO Cloning kit (Invitrogen) following the manufacturer’s protocols. Clones were screened using PCR and primers that flanked the multi-cloning site of the vector. Clones were determined to be positive if they had the appropriate size insert in the plasmid (~1600 bp). The 16S rDNA gene from the clone designated p16RK3 was sequenced according to Andree et al. (1997) and aligned manually using Mac DNAseq software (Hitachi Software Engineering America).

**Phylogenetic analysis.** The sequence from clone p16RK3 was compared with data in GenBank using the BLAST algorithm (Altschul et al., 1990). The organisms with the highest ranking were used in our initial taxonomic characterization together with extant *Rickettsiales*-like bacteria from various taxa. We applied the methods of Jukes & Cantor (1969) in performing evolutionary distance analysis with bootstrap support (500 replications) using MEGA software ( Molecular Evolutionary Genetics Analysis version 1.01). This initial phylogenetic analysis used 1340 characters (or ~86%) of the 16S rDNA gene (data not shown). Positions in which no homology was observed between species were omitted from the analysis. A second evaluation included only those organisms that were most proximal to the abalone RLP in our first evaluation. For this second analysis we included most of the 16S rDNA sequence (1496 bases or 98% of the abalone RLP gene) from the 15 most proximal species used in our first analysis. We eliminated only the most distal ends of the 16S rDNA sequence such that the entire sequence of the shortest 16S rRNA gene deposited in GenBank served as the template for comparison. In addition, we deleted a large insertion of ~150 bases from the 16S rDNA gene of *Caedibacter caryophila* as this sequence was not shared with any of the other species in our study.

**Accession numbers for nucleotide sequences.** The 16S rDNA gene sequences of selected bacteria were electronically retrieved from GenBank. These included: *Anaplasma marginale*, M60313; *Caedibacter caryophila*, X71837; *Cowdria ruminantium*, X61659; *Coxiella burnetii*, M21291; *Ehrlichia sennetsu*, M73225; *Ehrlichia risticii*, M21290; *Ehrlichia phagocytophila*, M73224; *Ehrlichia bovis*, U03775; *Escherichia coli*, J01695; *NHP bacterium*, U65509; *Piscirickettsia salmonis*, X60783; *Rickettsia prowazekii*, M21789; *Rickettsia rickettsii*, M21293; and *Wolbachia pipientis*, X61768.

**RESULTS**

**Histology**

The RLP stained Gram-negative and was observed within cytoplasmic vacuoles of abalone gastrointestinal epithelial cells. The bacterial inclusions were usually situated apical to the nucleus of the host cells (Fig. 1). The bacterium was found in cells of the post-oesophagus, transport ducts of the digestive gland.
Fig. 2. (a) Transmission electron micrograph showing pleomorphic rod-shaped RLPs (arrows) within membrane-bound phagosomes of abalone gastrointestinal cells. Bar, 2 µm. (b) The bacterium divides by binary fission (arrows) and is emarginated by a trilaminar cell wall and lacks a visible peptidoglycan layer (inset: arrowhead). Ribosomes are located peripherally and fibrils of nucleoid material are centrally located. Bar, 500 nm, inset: Bar, 100 nm. (c) Inclusion containing heteromorphic bacteria. Arrows illustrate the larger spherical life stage that is predominantly observed in smaller phagosomes. The rod-shaped stage is denoted by an arrowhead. Bar, 2 µm.
No morulae were observed. Deeply purple with haematoxylin and eosin (Fig. 1). An electron-dense inner plasma membrane and an emarginated by a trilaminar cell wall that consists of the outer membrane of equal electron density that are separated by an electron-lucent layer without a visible peptidoglycan layer (Fig. 2b). The outer cell wall, although wavy, lacks the intense rippling observed in members of the genus *Ehrlichia*. The bacterium contains many ribosomes, located around the periphery of the cell, and centrally located fibrils of nucleoid material (Fig. 2b insert). The bacterium lacks a slime layer that is observed surrounding rickettsiae in the typhus and spotted fever groups (Weiss & Moulder, 1984a).

**Antigenic reactivity with Chlamydiaceae-specific antibodies**

No specific staining of the RLP inclusions was observed using either *Chlamydiaceae*-specific antibody or the preimmune mouse IgG. In contrast, the sections of avian lung infected with *Chlamydophila* (*Chlamydia psittaci*) had scattered positive (red) staining throughout the tissue (not shown).

**Antimicrobial susceptibility**

Microscopic examination of stained tissue sections from infected abalone treated with antimicrobials indicated that the RLP was only sensitive to the tetracyclines: oxytetracycline (*p* < 0.001, *n* = 9–12) and tetracycline (*p* < 0.05, *n* = 8–10, Table 1). Administration of nine IM doses of 21 mg oxytetracycline kg⁻¹ effectively cured RLP infections in both red and black abalone (Table 1).

### 16S rDNA sequence analysis

A sequence alignment was made using the 16S rRNA sequences from p16RK3, and the 29 representative α- and γ-Proteobacteria listed above that were used in a similar analysis of the shrimp NHP bacterium (Loy *et al.*, 1996). The bacterial agent from abalone shares sequence motifs with the shrimp NHP bacterium but only 70-9% sequence similarity overall. A BLAST search analysis determined that the 16S rDNA sequence of the abalone bacterium appears to be most closely related to *Anaplasma marginale* (77.3% similarity), *Ehrlichia bovis* (75.8%) and *Wolbachia pipientis* (74.1%) when the entire 16S sequence was used in the analysis (data not shown).

### Transmission electron microscopy

The bacterium was pleomorphic and was found dividing within cytoplasmic vacuoles of host cells that appear to be phagosomes or phagolysosomes (Fig. 2). The bacterium is predominantly a slender, rod-shaped organism with a pleomorphic spherical to coccobacillus form observed primarily in smaller and, presumably, younger phagosomes (Fig. 2a). The rod-shaped form replicates by binary fission and measures a mean of 332 nm × 1550 nm, with a range of 122–598 nm × 570–4460 nm (Fig. 2a, b). The spherical form measures a mean of 1405 nm in maximum dimension, with a range of 783–2190 nm (Fig. 2c). In addition, there appears to be an intermediate, coccobacillus size variant with a mean diameter of 620 nm (559–658 nm) and length of 1270 nm (Fig. 2c). The bacterium is emarginated by a trilaminar cell wall that consists of an electron-dense inner plasma membrane and an

### Table 1. In vivo testing of the susceptibility of red and black abalone to several antimicrobials administered by 9–12 intramuscular injections

<table>
<thead>
<tr>
<th>Antimicrobial Dose</th>
<th>Percentage of red abalone with RLP foci</th>
<th>Percentage of black abalone with RLP foci</th>
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<tr>
<td>Chloramphenicol</td>
<td>90 (n = 10)</td>
<td>100 (n = 8)</td>
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<tr>
<td>12 mg kg⁻¹</td>
<td>80 (n = 10)</td>
<td>87.5 (n = 8)</td>
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<tr>
<td>Clarithromycin</td>
<td>100 (n = 8)</td>
<td>100 (n = 6)</td>
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<tr>
<td>1.96 mg kg⁻¹</td>
<td>100 (n = 8)</td>
<td>100 (n = 6)</td>
</tr>
<tr>
<td>Oxytetracycline (Liquimycin LA)</td>
<td>100 (n = 10)</td>
<td>100 (n = 8)</td>
</tr>
<tr>
<td>Sarafloxacin</td>
<td>0† (n = 10)</td>
<td>0† (n = 9)</td>
</tr>
<tr>
<td>21 mg kg⁻¹</td>
<td>100 (n = 8)</td>
<td>100 (n = 6)</td>
</tr>
<tr>
<td>Tetracycline</td>
<td>100 (n = 8)</td>
<td>100 (n = 6)</td>
</tr>
<tr>
<td>None*</td>
<td>90 (n = 10)</td>
<td>100 (n = 8)</td>
</tr>
<tr>
<td>50 mg kg⁻¹</td>
<td>100† (n = 10)</td>
<td>0† (n = 8)</td>
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</table>

* Control abalone were administered diluent (2% sterile saline) only.
† Treatments with rickettsial infection prevalences significantly less than that of controls (*p* < 0.001), Mann–Whitney Rank Sum test.
‡ Treatments with rickettsial infection prevalences significantly less than that of controls (*p* < 0.05), Mann–Whitney Rank Sum test.

and, less frequently, the intestine. The predominantly observed inclusion was spherical to oblong in shape, measured 14.1–56.4 μm with a mean maximum dimension of 35.8 ± 13.0 μm, and contained homogeneous and densely packed bacteria that stained deeply purple with haematoxylin and eosin (Fig. 1). No morulae were observed.

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most closely with Ehrlichia risticii and Ehrlichia senmetsu (data not shown). Collectively, these data illustrate the uniqueness of the 16S rDNA sequence of the abalone RLP.

**DISCUSSION**

Wayne et al. (1987) recommended that bacterial nomenclature should reflect genomic relationships. Confirmatory phenotypic data should also be sought prior to the introduction of a new name (Nelson et al., 1984). Recent bacterial taxonomy has increasingly relied on both phenotypic and genomic information including a new species in the genus Rickettsia, Rickettsia japonica (Uchida et al., 1992) and in the genus Nocardia, Nocardia crassostreae (Friedman et al., 1998). Those bacteria that cannot be cultured provide fewer phenotypic characters and therefore phylogenetic analyses become central to taxonomic placement. Based on proposals by Murray & Schleifer (1994) and Murray & Stackebrandt (1995; the provisional status of *Candidatus* was created for unculturable organisms and poorly characterized organisms, e.g. *Candidatus Arsenophonus tristominarum*, Hyspa & Dale, 1997). Despite this, several species of *Ehrlichia* have been described based principally upon genomic sequences such as *Ehrlichia chaffeensis* (Anderson et al., 1991), *Ehrlichia ewingii* (Anderson et al., 1992) and *Ehrlichia muris* (Wen et al., 1995). *Ehrlichia chaffeensis* was differentiated largely on its unique 16S rRNA sequence. Phylogenetic analyses using 16S sequence analysis have been used to unify two genera: Bartonella and *Rochalimaea* in the family *Bartonellaceae*, and to remove the family *Bartonellaceae* from the order Rickettsiales (Brenner et al., 1993). We used a combination of morphological, tinctorial, physiological, pathological and genomic traits to identify the bacterium associated with WS in

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**Table 2.** Genetic distance table illustrating the relationship of the WS bacterium and other Proteobacteria using MEGA

A total of 1496 nucleotides were included in the data set after missing information data and gaps were removed.

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**Fig. 3.** Dendrogram showing evolutionary relationship of the abalone RLP (WS-RLP) to members of the families Rickettsiaceae and *Piscirickettsiaceae* with *Escherichia coli* as a representative genetically distant bacterium. The number at branch points represents the percentage occurrence of a given branch during 500 replicates of the bootstrap analysis. Each bar is approximately equal to a distance of 0.002668. Superscript numbers denote the following sequence accession numbers: 1, AF133090; 2, M21291; 3, X60783; 4, J01695.
abalone as a previously undescribed bacterium in the family Rickettsiaceae and propose inclusion in the provisional status, Candidatus with the name of ‘Candidatus Xenohaliotis californiensis’.

The RLP that infects abalone is a small, Gram-negative, pleomorphic bacterium that forms large intracytoplasmic inclusions within gastrointestinal epithelial cells. These inclusions lack the morphology of morulae (Fig. 1), characteristic of the genus Ehrlichia. The bacterium is an obligate intracellular parasite as evidenced by unsuccessful culture on artificial media (C. S. Friedman & R. P. Hedrick, unpublished results). Collectively, these traits confirm placement of the abalone RLP in the order Rickettsiales. Tissue-specificity of the abalone RLP resembles that of Rickettsia spp. or Anaplasma marginale which often infect the midgut of arthropod vectors (Ribeiro et al., 1996). Anaplasma marginale also infects blood cells of its mammalian host, as do members of the genus Ehrlichia (Anderson et al., 1991; Ribeiro et al., 1996; Ristic & Huxsoll, 1984). Wolbachia pipiens infects germinal cells and epithelia of the insect vector (mosquito), while Wolbachia melophagi is observed in the lumina of the host (sheep) gastrointestinal tract (Weiss et al., 1984). Host-specificity and transmission of the abalone RLP is unique as this type of bacterium has not previously been described in a marine gastropod and transmission is direct (Friedman et al., 1997) as suggested for Piscirickettsia salmonis, a marine agent that infects salmon (Fryer et al., 1992).

The abalone bacterium is pleomorphic with multiple size variants as observed in some genera of rickettsia. For example, the life cycle of Coxiella burnetti and Cowdria ruminantium includes a vegetative and relatively electron-lucent large cell variant and a small cell variant that is quite electron-dense (Weiss & Moulder, 1984b; Jongejan et al., 1991). Bonami & Pappalardo (1980) observed a dimorphic, membrane-bound, intracytoplasmic rickettsia which is pathogenic to the crab, Carcinus mediterraneus. The life cycle of this crab pathogen also included an electron-dense small-cell variant and a larger and more electron-lucent cell variant. The progression of size variants suggests that this is a single species with a dimorphic life history as observed in other members of the family Rickettsiaceae. In addition, the rickettsia found in abalone lacks a visible peptidoglycan layer that is often lacking in rickettsial bacteria (e.g. Ehrlichia spp.; Kawahara et al., 1993). However, the outer cell wall of the abalone bacterium lacks the intense rippling observed in members of the genus Ehrlichia. The RLP from abalone lacked a slime layer found in Rickettsia spp. in the typhus and spotted fever groups (Weiss & Moulder, 1984a). In addition, pan-Chlamydia antibodies did not recognize the abalone RLP. These traits illustrate the unique morphology and life history of the RLP that infects abalone relative to described members of the family Rickettsiaceae.

Using rRNA sequence analyses, Weisburg et al. (1989) determined that the family Rickettsiaceae is poly-phyletic and is comprised three tribes: Rickettsiiaea, Ehrlichieae and Wolbachieae. Our study indicates that the abalone RLP shares similarities with members of all three tribes but is phylogenetically distinct from each tribe. Evolutionary relationships to other rickettsiae shown in the distance table (Table 2) and dendrogram (Fig. 3) illustrate that the 16S rDNA of the abalone RLP is a member of the z-subclass of the class Proteobacteria. Like other members of the family Rickettsiaceae in the z-subclass, the bacterium does not belong to the four recognized subtaxa of the z-Proteobacteria (α-1, α-2, α-3 and α-4; Fig. 3). This information clearly differentiates the abalone RLP from the γ-subclass of the class Proteobacteria that includes genera such as Coxiiella and Piscirickettsia. In addition, members of the genus Rickettsia form a monophyletic group within the z-subclass that is distinct from the abalone RLP and four genera (Ehrlichia, Wolbachia, Cowdria and Anaplasma) with which it forms a polyphyletic group. As illustrated in Fig. 3, the 16S rDNA sequence of the abalone RLP represents a lineage that is distinct from the other genera within this group. As shown in our study, van Vliet et al. (1992) also found that despite differences in morphology and host and target cell preferences, Cowdria ruminantium and Ehrlichia canis and Ehrlichia chaffeensis form a closely related cluster upon analysis of 16S gene sequences. Collectively, these traits in conjunction with a lack of the full complement of phenotypic data required for validation of a new genus and species, we propose the name, ‘Candidatus Xenohaliotis californiensis’ (Xe.no.ha.li.o.tis’ L. fem. n. xeno foreign; Haliotis generic name of its abalone host; N.L. fem. n. Xenohaliotis a foreign organism in abalone; cal.i.for.ni.en.sis. N.L. fem. adj. californiensis regarding California, USA, the area where the organism was initially identified).

Description of ‘Candidatus Xenohaliotis californiensis’

Gram-negative, periodic acid–Schiff base stain-negative, pleomorphic coccobacillus, ~332 × 1550 nm with an irregular spherical cell stage ~1450 nm in maximum dimensions. Stains basophilic with haematoxylin and eosin. Non-motile. Gram-negative cell wall that lacks visible peptidoglycan and slime layers, ribosome rich with fibrilar nucleoid. Replicates within spherical to oblong, intracytoplasmic membrane-bound vacuoles of abalone gastrointestinal transport or absorptive epithelium. Bacteria-filled vacuoles are generally located distal to host nucleus. Not cultivable on synthetic media or in fish cell lines (e.g. CHSE-214, American Type Culture Collection). Sensitive to tetracyclines but not to chloramphenicol, clarithromycin or sarafloxacin. Based on 16S rDNA sequence analysis, the bacterium is a member of an undefined z-subclass of the class Proteobacteria. The bacterium can be diagnosed with tissue squashes stained with propidium iodide, microscopic examination of stain tissue sections and can be differentiated from other closely
related \textit{\alpha-Proteobacteria} by its unique 16S rDNA sequence.

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\section*{REFERENCES}


