Isolation and Characterization of *Desulfitobacterium dehalogenans* gen. nov., sp. nov., an Anaerobic Bacterium Which Reductively Dechlorinates Chlorophenolic Compounds

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An organism that is able to reductively ortho-dechlorinate 2,4-dichlorophenol and 3-chloro-4-hydroxyphenylacetate (3-Cl-4-OHPA) was isolated from a methanogenic lake sediment. This organism, an anaerobic, motile, Gram-type-positive, rod-shaped bacterium, grew in the presence of 0.1% yeast extract when pyruvate, lactate, formate, or hydrogen was used as the electron donor for reductive dehalogenation of 3-Cl-4-OHPA. Sulfite, thiosulfate, and sulfur were reduced to sulfide, nitrate was reduced to nitrite, and fumarate was reduced to succinate. Dissimilatory reduction of sulfate could not be demonstrated, and no adenyllysulfate reductase was detected with an immunoassay. The organism fermented two pyruvate molecules to one lactate molecule, one acetate molecule, and one carbon dioxide molecule. The pH and temperature optima for both growth and dechlorination of 3-Cl-4-OHPA were 7.5 and 38°C, respectively. The doubling time under these conditions was approximately 3.5 h. On the basis of the results of a 16S rRNA analysis and the inability of the organism to use sulfate as an electron acceptor, strain JW/IU-DC1 is described as the type strain of the new taxon *Desulfitobacterium dehalogenans* gen. nov., sp. nov.

Although there have been many reports of reductive dehalogenation of chloroaromatic compounds in sediments and enrichment cultures (25), only two obligate anaerobes have been isolated in pure culture, the Gram-type-negative sulfate-reducing bacterium *Desulfomonile tiedjei* DCCB-1 (10, 28) and a clostridium-like strain, DCCB-2 (22).

In this paper we describe the isolation and characterization of an anaerobic Gram-type-positive organism, *Desulfitobacterium dehalogenans* gen. nov., sp. nov., that is capable of reductive ortho-dechlorination of chloroaromatic compounds such as 2,4-dichlorophenol (2,4-DCP) and 3-chloro-4-hydroxyphenylacetate (3-Cl-4-OHPA).

**MATERIALS AND METHODS**

**Origin of enrichment culture and organism.** The enrichment culture used in this study was obtained from a freshwater sediment collected from a pond located in a wooded area of the Sandy Creek Nature Park, Athens, Ga. (7, 19). *Desulfobacterium dehalogenans* JW/IU-DC1 (T = type strain) was isolated from the enrichment culture as described below.

**Enrichment, isolation, and cultivation of the dechlorinating organism.** Using oxygen-free nitrogen gas, we prepared media by the Hungate technique (21) and sterilized media by autoclaving for 40 min at 121°C. Unless indicated otherwise, cultures were grown anaerobically at 37°C in Hungate tubes containing 5 to 10 ml of medium.

The enrichment culture was grown in a mineral medium described previously (39), except that our medium contained 40 mM KH₂PO₄ and no TAPS. The enrichment medium was supplemented with yeast extract (0.2 to 1%) as a carbon source. The pH was kept within the range from 8.0 to 8.4 by adjusting it with anaerobic sterile 2 N NaOH. The medium used for further enrichment and isolation was prepared by using the spent medium from a coculture of *Clostridium* sp. strain JW/IU-YU1 and a facultative anaerobic coccus (JW/IU-DC1), both of which were isolated previously from the primary stable enrichment culture. The spent medium was obtained by growing the coculture in the mineral medium described above supplemented with 0.5 or 0.1% yeast extract and 0.1% (wt/vol) glucose. After centrifugation of the cultures (which were kept in Hungate tubes), transfer of the supernatant into new sterile, anaerobic tubes, autoclaving for 40 min at 121°C, and adjustment of the pH to 8.2 with anaerobic, sterile 2 N NaOH, the supernatant was supplemented with 15 mM 3-Cl-4-OHPA. Final isolation was accomplished by repeatedly isolating single colonies growing in agar (0.6 and 0.9%, wt/vol) shake cultures, using the enrichment medium described above supplemented with 2 mM pyruvate, 2 mM formate, and 1 mM 3-Cl-4-OHPA. The mineral medium (base medium) used for cultivation of the purified culture was the same as the enrichment medium (see above) except that it contained 40 mM KH₂PO₄ and no TAPS. The pH was adjusted to 7.5 with anaerobic sterile 2 N NaOH after the medium was autoclaved at 121°C for 40 min. Unless indicated otherwise, the medium was supplemented with 0.1% yeast extract, 20 mM pyruvate, and 10 mM 3-Cl-4-OHPA (purified-culture medium).

**Light microscopy and electron microscopy.** Phase-contrast light microscopy (model PM-10AD; Olympus Optical Co., Ltd., Tokyo, Japan) was used for routine examinations and for taking photomicrographs of agar-coated slides. Electron microscopy and preparation of cells for ultrathin sectioning were carried out as described previously (13, 17, 29) by using uranyl acetate and lead citrate for poststaining. Cells were negatively stained as described by Valentine et al. (30) and Beuscher et al. (3) by using 2% uranyl acetate.

**Gram stain reaction and Gram type.** Gram staining was performed by a modified Hucker method (12), using an Enhanced Gram Stain Kit (Carr Scarborough Microbiologicals, Inc., Decatur, Ga.). The Gram type (34) was determined...
by the polymixin B-lipopolysaccharide assay as described previously (36).

**Substrate utilization and electron donors for dehalogenation.** Tubes containing the base medium described above were supplemented with 0.1% yeast extract and sterilized substrates at final concentrations of 0.2% (wt/vol) for the sugars and 20 mM for all of the other organic substrates. Control tubes contained no added carbon source. Cultures grown on 20 mM pyruvate at 37°C and pH 7.5 were used as inocula (2% vol/vol). The use of potential electron donors for reductive dehalogenation was assayed in cultures containing 10 mM 3-Cl-4-OHPA. Tubes containing hydrogen as the electron donor were incubated with shaking at 150 rpm (New Brunswick Scientific Co., Inc., Edison, N.J.). A culture grown on medium containing 20 mM pyruvate plus 10 mM 3-Cl-4-OHPA was used as the inoculum. Growth was monitored by determining the increase in optical density at 600 nm at 37°C and pH 7.5. Assays for reductive dechlorination (see below) were performed in duplicate. The maximum incubation time was 11 days.

**Electron acceptors.** The use of different electron acceptors was studied with cultures grown in base medium supplemented with 0.1% yeast extract and various test compounds, using 20 mM pyruvate as the carbon source at 37°C and pH 7.5. All assays were carried out in duplicate. Two uninoculated tubes were used as controls. Chromophoric compounds and products of the reduction of these compounds were quantified by high-performance liquid chromatography (HPLC) (Rabbit HP System; Rainin Instrument Co.,) using a C_{18} reversed-phase column as described by Kohring et al. (19). The test for dissimilatory reduction of nitrate (5 mM) and fumarate (10 mM) was performed in Hungate tubes containing purified-culture medium. Concentrations of nitrate and nitrite were measured spectrophotometrically as described by Hanson and Philips (14). Fumarate and succinate were quantified by gas chromatography, using a model 5880A instrument (Hewlett-Packard, Avondale, Pa.) and a Chromosorb WAW 10% SP-1000 glass column (Supelco, Inc., Bellefonte, Pa.). They were measured with a flame ionization detector as described previously (35). Purified-culture medium lacking Na_{2}S and supplemented with 0.1% FeSO_{4} was used to assay for dissimilatory reduction of sulfate, sulfite, thiosulfate, and sulfur. Tubes were supplemented with 5 and 10 mM Na_{2}SO_{4}, 2 mM Na_{2}S_{2}O_{3}, 5 and 20 mM Na_{2}S_{3}O_{8}, and sulfur powder (about 0.2% added with a spatula in an anaerobic chamber). All tubes were pressurized with hydrogen (20 lb/in^{2}). Adenyllylsulfate reductase was detected with an immunoassay test kit for the detection of sulfate-reducing bacteria (Conoco Specialty Products, Houston, Tex.).

**pH and temperature ranges.** The pH and temperature ranges for growth and dechlorination were determined by growing the dechlorinating organism in Hungate tubes containing 10 ml of purified-culture medium. The medium used to determine the pH range contained 25 mM potassium phosphate. The pH was kept constant by frequently adjusting it with sterile anaerobic 2 N NaOH. The pH range was determined at 37°C. The temperature range was determined at pH 7.5 by using a temperature gradient incubator (Scientific Industries, Inc., Bohemia, N.Y.) set at temperatures ranging from 1.6 to 53°C with a shaking speed of 15spm.

**Sporeulation.** To induce spore formation, Desulfotobacterium dehalogenans was grown under various conditions, including in liquid or agar (0.6%, wt/vol)-solidified purified-culture medium. In a coculture with Clostridium sp. strain JW/1U-YU1 and facultatively anaerobic coccus strain JW/1U-YU2, Desulfotobacterium dehalogenans was grown in 0.5% yeast extract medium supplemented with 0.4 mM 2,4-DCP at 37°C and pH 8.0. Cultures were analyzed for spore formation by phase-contrast light microscopy and by testing for heat resistance (i.e., growth after treatment at 87°C for 30 min).

**Fermentation products.** H_{2} and CO_{2} were determined by gas chromatography (Varian, Walnut Creek, Calif., instrument equipped with a Porapak Q 80/100-mesh column [Supelco] and a thermal conductivity detector). The temperatures of the injection port and the column were 160 and 120°C, respectively. The carrier gas was nitrogen, and the flow rate was 30 ml/min. Volatile fatty acids and alcohols, as well as nonvolatile fatty acids, were analyzed as described previously (35) by using a model 5880A gas chromatograph (Hewlett-Packard) equipped with a Chromosorb WAW 10% SP-1000 glass column (Supelco) and a flame ionization detector. Formic acid contents were determined by monitoring NAD reduction at 340 nm in the presence of NAD-dependent formate dehydrogenase obtained from yeast cells (Boehringer Mannheim GmbH, Mannheim, Germany). Acetic acid contents were determined enzymatically by using an acetic acid reagent kit (catalog no. 148-261; Boehringer Mannheim GmbH).

**Determination of G+C content.** DNA was isolated as described by Ausubel et al. (2) by using GCA gradient ultracentrifugation. The guanine-plus-cytosine (G+C) content was determined after enzymatic digestion and HPLC separation of nucleosides as described by Whitman et al. (32) and Mesbah et al. (24).

**16S rRNA isolation, sequencing method, and data analysis.** 16S rRNA was isolated from strain JW/1U-DCIT\textsuperscript{1} by previously described procedures (23, 38). The dioxygenase chain termination method (4, 27), adapted for direct 16S rRNA sequencing using reverse transcriptase (20), was employed. Synthesized strands were labeled by including [α-\textsuperscript{35}S]thio-dATP (4). A standard set of primers (usually eight) specific for (eu)bacterial 16S rRNAs (4, 20, 31) was routinely used. All sequences were >90% complete. The sequences were aligned by using sequence editor aeq2 (C. R. Woese) with the sequences of a representative collection of (eu)bacterial 16S rRNAs (26, 37). Corrected pairwise distances (expressed as estimated numbers of changes per 100 nucleotides) were computed from percentages of similarity by using the Jukes-Cantor correction (16) as modified by G. J. Olsen (described in reference 31) to accommodate the actual nucleotide ratios. Dendograms were constructed from evolutionary distance matrices by using the algorithm of De Soete (9).

**Sensitivity to oxygen.** Sensitivity of bacterial growth and dechlorination of 3-Cl-4-OHPA to oxygen was determined in 160-ml serum bottles containing 10 ml of purified-culture medium (which contained no reducing solution) under a nitrogen atmosphere in the presence of 0, 0.5, 1, 2, 5, and 9% air. The medium in each tube was inoculated with 0.25 ml of a culture of strain JW/1U-DCIT\textsuperscript{1} in the exponential growth phase. The bottles were incubated in a shaking incubator (New Brunswick Scientific Co.) at 37°C and 150 rpm. The maximum incubation time was 15 days.

**Effect of sodium chloride on growth and dechlorination.** Sensitivity of growth and dechlorination to NaCl was tested by transferring portions (2%, vol/vol) of an exponentially growing culture into fresh purified-culture medium containing 0, 0.5, 1, 1.5, 2, and 2.5 M NaCl.

**Nucleotide sequence accession number.** The GenBank accession number for the 16S rRNA sequence of strain JW/1U-DCIT\textsuperscript{1} is L28946.
TABLE 1. Evolutionary distances between Desulfitobacterium dehalogenans and related organisms

<table>
<thead>
<tr>
<th>Organism</th>
<th>Evolutionary distance</th>
</tr>
</thead>
<tbody>
<tr>
<td>Desulfitobacterium dehalogenans</td>
<td>9.2</td>
</tr>
<tr>
<td>Desulfitomaculum orientis</td>
<td>14.6</td>
</tr>
<tr>
<td>Desulfitomaculum nigrificans</td>
<td>15.0</td>
</tr>
<tr>
<td>Heliosoma chlorum</td>
<td>14.7</td>
</tr>
<tr>
<td>Sporomusa paucivorans</td>
<td>19.0</td>
</tr>
<tr>
<td>Megaplasta eldigenii</td>
<td>18.6</td>
</tr>
<tr>
<td>Desulfitomaculum australicum</td>
<td>15.4</td>
</tr>
<tr>
<td>Desulfitomaculum thermobenzoicum</td>
<td>14.8</td>
</tr>
<tr>
<td>Acetogenium kivui</td>
<td>17.7</td>
</tr>
<tr>
<td>Clostridium thermoacetiticum</td>
<td>20.8</td>
</tr>
<tr>
<td>Clostridium thermosacccharo-lyticum</td>
<td>18.2</td>
</tr>
<tr>
<td>Clostridium fervidus</td>
<td>20.1</td>
</tr>
<tr>
<td>Clostridium butyricum</td>
<td>21.2</td>
</tr>
<tr>
<td>Clostridium pasteurianum</td>
<td>18.9</td>
</tr>
<tr>
<td>Clostridium perfringens</td>
<td>21.5</td>
</tr>
<tr>
<td>Clostridium kluyveri</td>
<td>21.8</td>
</tr>
<tr>
<td>Clostridium barkeri</td>
<td>18.2</td>
</tr>
<tr>
<td>Clostridium acidilentic</td>
<td>18.6</td>
</tr>
<tr>
<td>Clostridium litusebacrense</td>
<td>18.4</td>
</tr>
<tr>
<td>Clostridium aminovalericum</td>
<td>20.8</td>
</tr>
<tr>
<td>Clostridium symbiosum</td>
<td>17.0</td>
</tr>
<tr>
<td>Clostridium thermocellum</td>
<td>18.0</td>
</tr>
<tr>
<td>Bacillus subtilis</td>
<td>16.1</td>
</tr>
</tbody>
</table>

*Only those positions having known bases in all sequences were used in the analysis. Bacillus subtilis was used as an unweighted outgroup.

RESULTS AND DISCUSSION

Isolation of dechlorinating strain JW/1U-DC1T. The starting enrichment culture (7) grown in the presence of 1% (wt/vol) yeast extract contained approximately 10⁶ to 10⁷ dehalogenating cells per ml of culture, as determined by serial dilution (1:5). Changing the chlorinating compound from 1 mM 2,4-DCP (higher concentrations were inhibitory) to 15 mM 3-Cl-4-OHPA and using spent medium from a coculture of the yeast extract-utilizing organism Clostridium sp. strain JW/1U-YU1...
and a facultatively anaerobic coccus (JW/IU-YU2), both of which were isolated from the enrichment culture, increased the concentration of the dechlorinating organisms approximately 100-fold. The culture consisted mainly of two different organisms. A pure culture of the dechlorinating organism was finally obtained by isolating single colonies in soft agar shake cultures containing 0.6 and 0.9% (wt/vol) agar (21). Higher agar concentrations were inhibitory for the dechlorinating culture but not for the contaminating organism, an unidentified clostridium-like organism. A convenient routine check for the purity of this culture was the lack of growth on and in 1.2% of 3-C1-4-OHPA and sulfate, respectively.

Phylogeny. The 16S rRNA analysis of strain JW/IU-DCT and related organisms based on 16S rRNA sequences (see Materials and Methods and Table 1). Bar = 10 inferred changes per 100 nucleotides. Abbreviations: A., Acetogenium; B., Bacillus; C., Clostridium; D., Desulfotomaculum; E. Desulfobacterium; H., Helibacterium; N., Megasphaera; S., Sporomusa. Bacillus subtilis was used as an arbitrary outgroup organism.

![FIG. 1. Unrooted phylogenetic tree for Desulfotobacterium dehalogenans JW/IU-DCT and related organisms based on 16S rRNA sequences (see Materials and Methods and Table 1). Bar = 10 inferred changes per 100 nucleotides. Abbreviations: A., Acetogenium; B., Bacillus; C., Clostridium; D., Desulfotomaculum; D. Desulfobacterium; H., Helibacterium; M., Megasphaera; S., Sporomusa. Bacillus subtilis was used as an arbitrary outgroup organism.](image)

**TABLE 2. Selected properties of Desulfotobacterium dehalogenans and Desulfotomaculum species**

<table>
<thead>
<tr>
<th>Species</th>
<th>Cell morphology</th>
<th>Morphology and location of spores</th>
<th>Growth with pyruvate without electron acceptors</th>
<th>Autotrophic growth</th>
<th>Electron acceptors</th>
<th>G+C content of DNA (mol%)</th>
<th>Oxidation of organic substrates</th>
</tr>
</thead>
<tbody>
<tr>
<td>Desulfotobacterium dehalogenans</td>
<td>Straight or curved rods</td>
<td>Usually not present</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>45</td>
</tr>
<tr>
<td>Desulfotomaculum orientis</td>
<td>Straight or curved rods</td>
<td>Slightly oval, terminal to subterminal</td>
<td>-d</td>
<td>-e</td>
<td>-d</td>
<td>-d</td>
<td>45</td>
</tr>
<tr>
<td>Desulfotomaculum nigricans</td>
<td>Rods</td>
<td>Oval, central</td>
<td>+d</td>
<td>+e</td>
<td>+d</td>
<td>-d</td>
<td>-f</td>
</tr>
<tr>
<td>Desulfotomaculum auranticus</td>
<td>Rods</td>
<td>Oval, central</td>
<td>+d</td>
<td>+e</td>
<td>+d</td>
<td>-d</td>
<td>-f</td>
</tr>
</tbody>
</table>

* Desulfotobacterium dehalogenans and the Desulfotomaculum species (5, 18) use lactate, pyruvate, H2, and formate but not acetate as electron donors for reduction of 3-Chloro-2,4-dihydroxyphenylacetate and sulfate, respectively.
* The values for Desulfotomaculum species are different from the values in references 5 and 6.
* Data from reference 33.
* Data from reference 18.
* Data from reference 15.
* Determined in the presence of 1 mmol of acetate per liter.
of the organism to dehalogenate various chlorophenolic compounds).

**Colony morphology.** The organism does not grow in or on the surfaces of solid 1 to 1.5% agar media containing 0.1% yeast extract, 20 mM pyruvate, and 1 to 10 mM 3-Cl-4-OHPA. Colonies in 0.7 to 0.8% agar shake tubes are translucent, white, and spherical to slightly irregular and have diameters of 2 to 3 mm.

**Morphology of vegetative cells.** Exponential-phase cells that are grown in liquid medium are slightly curved rods that are 0.5 to 0.7 μm in diameter and 2.5 to 4 μm long (Fig. 2A and B). The cells become more pleiomorphic (i.e., thicker and shorter cells appear) in the early stationary growth phase.

**Gram stain reaction and Gram type.** The Gram stain reaction is positive, regardless of the growth phase. Furthermore, tests for formation of the lipopolysaccharide-polymyxin B complex give no indications for the presence of lipopolysaccharide (36). Thus, the organism is a Gram-type-positive (34)

Growth properties. (i) Effects of pH and temperature on growth. The pH range for growth and dechlorination of 3-Cl-4-OHPA is pH 6 to 9 (as determined at 37°C), and the optimum pH is 7.5 (Fig. 3). The shortest doubling time is about 3.5 h at pH 7.5 and 37°C. A similar pH profile with an optimum at about pH 7.5 has been observed for dechlorination of 2,4-DCP (data not shown). The temperature range for growth and dechlorination of 3-Cl-4-OHPA is 13 to 45°C, and the optimum temperature is around 38°C (Fig. 4).

(ii) Effect of oxygen. We determined the sensitivity of strain JW/IU-DC1T to oxygen by measuring growth and dechlorination of 3-Cl-4-OHPA in 160-ml serum bottles containing 10 ml of medium under a nitrogen gas phase containing different concentrations of air. Growth and dechlorination do not occur under aerobic conditions and when the concentration of air in the gas phase is 5% or more. However, dechlorination does occur under a nitrogen atmosphere containing 2% air, indicating that the organism is resistant to microaerophilic conditions. Determining whether the organism can use O2 as an electron acceptor will require further research. It has been shown recently that under special conditions some sulfate reducers, which have been assumed to be strictly anaerobic bacteria, not only can survive exposure to air (1) but also can utilize O2 (8, 11).

(iii) Effect of sodium chloride. Growth and complete dechlorination of 10 mM 3-Cl-4-OHPA occur in the presence of 0.5 M NaCl, but growth and dechlorination do not occur in the presence of an concentration of 1 M or more. This observation is consistent with the previous observation that the enrichment culture is not active when it is transferred to marine sediments unless the sediment is diluted at 2:1 with water, thus reducing the salt concentration (7).

(iv) Substrates and fermentation products. The only defined carbon source found which is utilized in the presence of 0.1% yeast extract under anaerobic conditions is pyruvate. Fermentation of 22.2 mol of pyruvate results in the formation of 10 mol of lactate, 9.2 mol of acetate, and 11.2 mol of carbon dioxide. We conclude that pyruvate is fermented according to the following equation: $2\text{CH}_3\text{COO}^- + \text{H}_2\text{O} \rightarrow \text{CH}_3\text{CH(OH)COO}^- + \text{CH}_3\text{COOH} + \text{CO}_2$.

Motility and flagellation. Most exponentially growing cells in purified-culture medium are motile, whereas cells in the later growth stages exhibit mainly tumbling motion. Each cell has one to four flagella (Fig. 2B).

FIG. 3. Effects of pH on the doubling time (tD) of JW/IU-DC1T (A) and the observed time for 50% dehalogenation (tso) of 10 mM 3-Cl-4-OHPA (B). Desulfitobacterium dehalogenans was grown at 37°C in modified purified-culture medium (See Materials and Methods).

FIG. 4. Effects of temperature on the doubling time (tD) of JW/IU-DC1T (A) and the observed time for 50% dehalogenation (tso) of 10 mM 3-Cl-4-OHPA (B). Desulfitobacterium dehalogenans was grown in purified-culture medium at pH 7.5.
Desulfitobacterium dehalogenans
reductive dechlorination and, consequently, growth: the dou-
bling times are 3.5, 6.2, 11.4, and 15.5 h, respectively. No
growth occurs in the absence of 3-C1-4-OHPA with lactate,
propionate, i-valerate, acetate, and formate.

(v) Electron donors. In the presence of 0.1% yeast extract
and 10 mM 3-C1-4-OHPA as an electron acceptor, pyruvate,
lactate, formate, and hydrogen are used as electron donors for
reductive dechlorination and, consequently, growth: the dou-
bling times are 3.5, 6.2, 11.4, and 15.5 h, respectively. No
growth occurs in the absence of 3-C1-4-OHPA with lactate,
propionate, i-valerate, acetate, and formate.

(vi) Electron acceptors. In addition to chlorophenolic com-
pounds, such as 3-C1-4-OHPA and 2,4-DCP, the following
inorganic and organic compounds are used as electron accep-
tors in the presence of 0.1% yeast extract and 20 mM pyruvate:
sulfite (2 mM), thiosulfate (20 mM), and sulfur (5%, wt/vol),
which are reduced to sulfide; nitrate (5 mM) (reduced to
nitrite); and fumarate (10 mM) (reduced to succinate). The
test for dissimilatory sulfate reduction, however, is negative.
Moreover, no adenylylsulfate reductase is detected by an
immunoassay (see Materials and Methods), thus excluding the
possibility that the organism is a sulfate-reducing bacterium.
Besides 3-C1-4-OHPA, cultures of JW/IU-DC1T grown in
purified-culture medium reductively ortho-dechlorinate a vari-
ety of chlorophenols, including the dichlorophenols 2,3-dichlo-
rophenol, 2,4-dichlorophenol, and 2,6-dichlorophenol, the tri-
chlorophenols 2,3,4-trichlorophenol, 2,3,6-trichlorophenol,
and 2,4,6-trichlorophenol, the tetrachlorophenols 2,3,4,5-tetra-
chlorophenol, 2,3,4,5-tetrachlorophenol, and 2,3,5,6-tetrachlo-
rophenol, and pentachlorophenol. The following chlorophe-
nots are not dehalogenated: 2-chlorophenol, 3-chlorophenol,
4-chlorophenol, 2,5-dichlorophenol, 3,4-dichlorophenol, 3,5-
dichlorophenol, and 3,4,5-trichlorophenol.

4-OHPA (mM)

FIG. 5. Correlation between dechlorination of 3-C1-4-OHPA
(measured by the formation of the product 4-hydroxyphenylacetate
[4-OHPA]) and growth of JW/IU-DC1T (optical density at 600 nm
[OD600]). Desulfitobacterium dehalogenans was grown in purified-
culture medium at 38°C and pH 7.5 by using pyruvate as the major
carbon source and 3-C1-4-OHPA as the major electron acceptor.

G+C content. The G+C content of the genomic DNA is
45% mol% (as determined by the chemical method). Although
this value is similar to values found for Desulfotomaculum
species (Table 2), other properties justify placing this organism
in a separate genus.

Type strain. Strain JW/IU-DC1 is the type strain of
Desulfotobacterium dehalogenans; this strain has been deposited
in the Deutsche Sammlung von Mikroorganismen as strain
DSM 9161. The properties of this strain are the same as the
properties described above for the species since it is the only
strain that has been isolated so far.

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