Some Factors Affecting Production and Assay of *Escherichia coli* Haemolysins

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**SUMMARY**

The amount of haemolysin produced by *Escherichia coli* grown under various gas phases was determined by the amount of growth obtained under these conditions. Calcium or strontium was required for activation of haemolysin. The haemolytic reaction was stopped by sodium citrate. The loss of haemolytic activity after incubation with trypsin and chymotrypsin indicated that at least the active group of the haemolytic molecule is protein or a peptide.

**INTRODUCTION**

Several studies have shown discordant results about the production and assay of *Escherichia coli* haemolysins. This paper attempts to clarify the effects of various gas phases on the production of haemolysin and to demonstrate a cation requirement for activation of the haemolysin.

**METHODS**

*Organism.* A haemolysin-producing strain of *Escherichia coli* type 06 (Iowa Stock Culture, ISC, no. 447), originally obtained from a patient with pyelonephritis, was used. A standard inoculum was obtained as previously described (Snyder & Koch, 1966).

*Measurement of haemolytic activity.* Total haemolysin content of the cultures was obtained by two-fold dilution of whole cultures. Filterable haemolysin was determined in the same manner with culture filtrates passed through a Millipore filter (0.45 μm porosity). The amount of haemolysin/ml. culture or filtrate was determined as previously described (Snyder & Koch, 1966).

*Measurement of growth and pH value.* Turbidometric measurements of growth were obtained with a Spectronic 20 spectrophotometer at a wavelength of 625 μm. The pH value of the culture was determined with a Beckman Zeromatic II pH meter.

*Preparation of media.* Beef heart for infusion medium (BHI) was prepared from Difco Beef Heart for infusion as described by the manufacturer. When glucose was added to the media, 1 ml. of a Millipore-filtered solution 20% (w/v) glucose was added to 100 ml. autoclaved media. In media buffered with phosphate, 0.23 g. K₃HPO₄ and 0.078 g. KH₂PO₄ were added to each 100 ml. of medium before sterilization. Alkaline meat-extract broth was prepared as described by Smith (1963).

*Incubation of cultures.* Anaerobic growth was obtained by using Brewer jars. The jars were flushed with nitrogen three times and filled with hydrogen, a palladium catalyst being used for removal of residual oxygen. Incubation in a CO₂ atmosphere
RESULTS

Role of cations in haemolysis

Filtrates of *Escherichia coli* cultures grown in BHI medium did not cause haemolysis of sheep erythrocytes unless Ca was added to the filtrate or to the diluent used for titration of haemolytic activity. This indicated that Ca was required for activation but not for production of haemolysin. Addition of various amounts of the Ca showed that a final concentration of 0.005 M-CaCl₂ caused maximal activation of the haemolysin (Table 1).

To determine whether other cations could substitute for calcium, various concentrations of other cations were added to the 0.15 M-NaCl used as for assay. Neither barium nor magnesium salts were effective; however, strontium replaced calcium (Table 1).

Table 1. Effect of various salts on haemolytic activation

<table>
<thead>
<tr>
<th>Salt</th>
<th>0.01 M</th>
<th>0.005 M</th>
<th>0.0025 M</th>
</tr>
</thead>
<tbody>
<tr>
<td>CaCl₂</td>
<td>32</td>
<td>32</td>
<td>16</td>
</tr>
<tr>
<td>MgSO₄</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>MgCl₂</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>BaCl₂</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>SrCl₂</td>
<td>32</td>
<td>16</td>
<td>16</td>
</tr>
<tr>
<td>Sr(NO₃)₂</td>
<td>8</td>
<td>16</td>
<td>16</td>
</tr>
</tbody>
</table>

Table 2. Effect of medium preparation on the calcium requirement for haemolytic activity

<table>
<thead>
<tr>
<th>Type of infusion</th>
<th>Without calcium</th>
<th>With calcium</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fresh veal (121°)*</td>
<td>64</td>
<td>128</td>
</tr>
<tr>
<td>Fresh beef heart (121°)</td>
<td>32</td>
<td>128</td>
</tr>
<tr>
<td>BHI (121°)</td>
<td>0</td>
<td>64</td>
</tr>
<tr>
<td>Fresh veal (56°)</td>
<td>0</td>
<td>128</td>
</tr>
<tr>
<td>Fresh beef heart (56°)</td>
<td>0</td>
<td>64</td>
</tr>
<tr>
<td>BHI (56°)</td>
<td>0</td>
<td>64</td>
</tr>
</tbody>
</table>

* Temperature of infusion.

Addition of CaCl₂ to culture filtrates caused a precipitate. To determine whether the role of Ca was in removing an inhibitor, uninoculated medium was precipitated with CaCl₂, the precipitate removed by filtration and the medium was dialyzed to remove excess Ca. This medium was then sterilized and inoculated with *Escherichia coli*. Haemolytic activity was not shown by the filtrates from cultures grown in the dialyzed medium unless calcium was added.

Smith (1963) reported production of haemolysin by *Escherichia coli* grown in
alkaline broth prepared by autoclaving 1 part fresh veal in 2 parts water, 1 % proteose-peptone and 0.5 % NaCl at a pH 7.6. A calcium requirement for activation of haemolysin was not reported. To determine whether the method of media preparation or type of medium affected the requirement for calcium, the experiment shown in Table 2 was done. Filtrates of cultures grown in fresh meat infusions prepared by the method of Smith did not require added Ca for haemolytic activity; however, dehydrated beef heart for infusion (Difco, Detroit, Michigan) prepared in the same manner required Ca. Haemolytic activity could not be obtained with filtrates from alkaline fresh meat infusions prepared at 56°C unless calcium was added. Addition of Ca to the filtrates from cultures grown in autoclaved infusions increased the haemolytic activity. Autoclaved fresh veal infusion dialyzed before inoculation with *E. coli* did not permit production of activated haemolysin. However, addition of calcium to the filtrate resulted in haemolytic activity.

Attempts to measure quantitatively Ca in the autoclaved infusions by several methods (Ferro & Ham, 1957; Clark & Collip, 1925; Diehl & Ellingbee, 1956) were unsuccessful. However, small amounts of calcium were detected in the infusions prepared by autoclaving. Calcium was not found in the other media.

**Inhibition of haemolytic activity.** Since the haemolytic reaction required calcium, the effects of sodium citrate and EDTA on inhibition of the reaction were investigated. Addition of 0.1 ml of 1.0 M-sodium citrate to 2 ml of an incubating haemolysin-erythrocyte mixture stopped the haemolytic reaction. EDTA partially inhibited the haemolysin.

**Effect of enzymes on the haemolysin.** Samples of haemolysin were incubated with trypsin, chymotrypsin, deoxyribonuclease and ribonuclease. Inactivation of the haemolysin was obtained only with trypsin and chymotrypsin, suggesting that the haemolysin or active portion of the molecule is a protein or peptide.

**Effect of gaseous environment on haemolysin production**

Figures 1, 2 show the relationships between growth in BHI media, haemolysin production and pH value in cultures of *Escherichia coli* type 06, ISC 447, grown aerobically in a shake culture (Fig. 1) and a stationary culture (Fig. 2). Calcium was added to the culture and filtrates for the haemolysin assay. The same measurements were made with cultures grown in 20% CO₂ (v/v) and air (Fig. 3) and anaerobically (Fig. 4). Filterable and non-filterable haemolysin was produced in all media, however, more haemolysin was produced by organisms grown aerobically than by those grown under the other environmental conditions tested. The production of filterable haemolysin during the logarithmic phase of growth confirms a previous report (Snyder & Koch, 1966). Both the stationary aerobic culture (Fig. 2) and the cultures grown in CO₂ (Fig. 3) showed a diauxic type of growth with an increase in both total and filterable haemolysin during each accelerated growth phase. In all cultures an increase in haemolysin production was associated with a decrease in culture pH value.

To determine whether better growth of the organisms would result in increased production of haemolysin, glucose and a phosphate buffer was added to BHI medium and the previous experiments were repeated. Increased growth, as indicated by O.D. about 0.7, was obtained in all cultures except the aerobic shaken culture. Diauxic growth was not obtained in the stationary aerobic culture or the cultures grown in CO₂ and haemolysin was produced only during the single logarithmic growth phase.
When compared with cultures grown without glucose there was an increase in total haemolysin produced by cultures grown in stationary aerobic culture as well as in CO₂ and anaerobically. However, there was no significant increase in the yield of filterable haemolysin.

Fig. 1. Effect of agitation on haemolysin production in medium without added carbohydrate. •—• Growth, ○—○ total haemolysin in culture, •—• filterable haemolysin.

Fig. 2. Effect of stationary growth on haemolysin production in medium without added carbohydrate. •—• Growth, ○—○ total haemolysin in culture, •—• filterable haemolysin.

Fig. 3. Effect of CO₂ on haemolysin production in medium without added carbohydrate. •—• Growth, ○—○ total haemolysin in culture, •—• filterable haemolysin.

Fig. 4. Effect of anaerobiosis on haemolysin production in medium without added carbohydrate. •—• Growth, ○—○ total haemolysin in culture, •—• filterable haemolysin.
DISCUSSION

The data presented show that production of *Escherichia coli* haemolysis was obtained during growth in various gas phases. More haemolysin was produced aerobically either in shaken or stationary cultures than in cultures grown in CO₂ or anaerobically. However, since better bacterial growth was obtained in cultures growing aerobically, glucose and phosphate were added to the cultures to improve growth. These additions increased bacterial growth especially in the cultures grown in CO₂ and anaerobically and increased the production of haemolysin. No significant increase in filterable haemolysin was obtained. Tomic-Karovic (1955) found more haemolysin was produced when *E. coli* was grown on blood agar in an atmosphere of CO₂ than in air and that the least haemolysin was produced in an anaerobic atmosphere. Ishii (1960) found greater production of haemolysin under CO₂ than in air. Lovell & Rees (1960) reported less haemolytic activity in cultures grown in CO₂ or hydrogen than those grown aerobically. Widholm (1953) found that various mixtures of oxygen and nitrogen did not affect haemolysin production. The differences in the reports most likely can be attributed to differences in time of sampling the culture filtrates. Cultures growing under more ideal environmental conditions will produce haemolysin at a faster rate than other cultures. In all cases, production of haemolysin is associated with a decrease in pH value and with growth of the organisms.

Our experiments on the role of cations in haemolysis show that cations are responsible for activation of the haemolysin and are not responsible for production. Bamforth & Dudgeon (1952) reported that calcium was required for haemolytic activity and that barium and strontium could be substituted for calcium; we were not able to show haemolytic activity with barium. Other workers (Lovell & Rees, 1960; Smith, 1963) did not report a requirement for calcium. However, these workers used an alkaline meat extract broth. Our studies with the alkaline meat extract show (1) that some calcium is present in the alkaline meat extract, and (2) that haemolysin cannot be demonstrated in cultures of *Escherichia coli* grown in dialyzed broth unless calcium is added.

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REFERENCES