Lack of a surface layer in *Tannerella forsythia* mutants deficient in the type IX secretion system

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*Tannerella forsythia*, a Gram-negative anaerobic bacterium, is an important pathogen in periodontal disease. This bacterium possesses genes encoding all known components of the type IX secretion system (T9SS). *T. forsythia* mutants deficient in genes orthologous to the T9SS-encoding genes *porK*, *porT* and *sov* were constructed. All *porK*, *porT* and *sov* single mutants lacked the surface layer (S-layer) and expressed less-glycosylated versions of the S-layer glycoproteins TfsA and TfsB. In addition, these mutants exhibited decreased haemagglutination and increased biofilm formation. Comparison of the proteins secreted by the *porK* and WT strains revealed that the secretion of several proteins containing C-terminal domain (CTD)-like sequences is dependent on the *porK* gene. These results indicate that the T9SS is functional in *T. forsythia* and contributes to the translocation of CTD proteins to the cell surface or into the extracellular milieu.

INTRODUCTION

Oral biofilms comprise more than 700 bacterial species and matrix substances and contribute to the development of periodontal disease (Aas et al., 2005). Although chronic periodontitis is caused by a mixed infection, specific microorganisms including *Porphyromonas gingivalis*, *Tannerella forsythia* and *Treponema denticola* are considered important for the initiation and progression of chronic periodontitis (Holt & Ebersole, 2005).

*P. gingivalis* encodes a variety of virulence factors, such as the extracellular and cell-surface cysteine proteinases Arg-gingipain and Lys-gingipain (Potempa et al., 2003; O’Brien-Simpson et al., 2003). Recently, these proteinases were shown to be secreted by the Por secretion system (PorSS) (Sato et al., 2005; Sato et al., 2010). The proteins constituting the PorSS differ from those constituting other secretion systems. The *P. gingivalis* PorSS includes the PorK, PorL, PorM, PorN, PorP, PorQ, PorT, PorU, PorV (PG27, LptO), PorW and Sov proteins (Sato et al., 2010). Coding sequences (CDSs) encoding proteins homologous to the *P. gingivalis* PorSS proteins are present in the genomes of several bacteria in phylum *Bacteroidetes* (McBride & Zhu, 2013). Therefore, the PorSS has been called the type IX secretion system (T9SS) (Chagnot et al., 2013).

*T. forsythia*, an anaerobic Gram-negative bacterium, belongs to phylum *Bacteroidetes*. It appears to possess the T9SS because it has genes encoding all known components of the T9SS (Sato et al., 2010). *T. forsythia* is phylogenetically related to *P. gingivalis*, however, unlike *P. gingivalis*, *T. forsythia* does not form black-pigmented colonies on blood-agar plates. Mixed infection by *T. forsythia* and *P. gingivalis* enhanced abscess formation in a murine model (Takemoto et al., 1997; Yoneda et al., 2001). *T. forsythia* encodes multiple potential virulence factors, including the PrtH proteinase and surface components such as surface layer (S-layer) glycoproteins (TfsA and TfsB) and the leucine-rich-repeat protein BspA (Sharma, 2010). Some virulence-related proteins, including TfsA, TfsB and BspA, appear to have C-terminal domains (CTDs) that may function as a recognition signal for the T9SS (Veith et al., 2009; Shoji et al., 2011).

In this study, *T. forsythia* mutants deficient in *porK*, *porT* and *sov* orthologous genes that may be involved in the translocation of CTD proteins such as TfsA, TfsB and BspA to the cell surface were generated. The *porK*, *porT* and *sov* mutant cells exhibited morphological changes and expressed less-glycosylated versions of the S-layer proteins.

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Abbreviations: CDS, coding sequence; CTD, C-terminal domain; NdPA, Nine Secretion System-dependent protein A; PorSS, Por secretion system; S-layer, surface layer; T9SS, type IX secretion system.

A supplementary table and a supplementary figure are available with the online version of this paper.
TfsA and TfsB. In the porK mutant, several CTD proteins were not secreted into the extracellular milieu. These results indicate that the T9SS is functional in T. forsythia and is important for the virulence of this bacterium.

**METHODS**

**Bacterial strains and culture conditions.** All bacterial strains and plasmids used in this study are listed in Table 1. T. forsythia cells were grown anaerobically (10% CO₂, 10% H₂, and 80% N₂) in enriched brain heart infusion broth (BHI) medium (Sato et al., 2010) supplemented with 10 μg ml⁻¹ N-acetylmuramic acid (MurNAc) (Sigma-Aldrich) and 5% (v/v) heat-inactivated calf serum (CS) and on enriched tryptic soy agar (Sato et al., 2010) supplemented with 10 μg ml⁻¹ MurNAc and 5% (v/v) defibrinated laked sheep blood. For the selection and maintenance of erythromycin (Em)-resistant T. forsythia strains, Em was added to the medium at a concentration of 5 μg ml⁻¹.

**Construction of bacterial strains.** Genomic nucleotide sequence data of T. forsythia ATCC 43037 was obtained from the GenBank database (accession number: CP003191). The T. forsythia porK insertion mutant was constructed as follows. A 0.6 kb 5'-terminal region of porK was amplified from the chromosomal DNA of T. forsythia ATCC 43037 using the Pyrobest DNA polymerase (TaKaRa) and PCR with the primers TFporKDF and TFporKDR. The amplified DNA was cloned into pCR4 Blunt TOPO and digested with BamHI sites of pBluescript II SK(−) to generate pKD1030. A 0.8 kb 3'-terminal region of porK was amplified from the chromosomal DNA of ATCC 43037 with the primers TFPorKDF and TFPorKDR. The amplified DNA was cloned into pCR4 Blunt TOPO and digested with BamHI and NotI. The resulting fragment was then inserted into the BamHI and NotI sites of pKD1030 to generate pKD1031. The 1.1 kb BamHI ermF DNA cassette was inserted into the BamHI site of pKD1031, resulting in pKD1032 (porK::ermF). The pKD1032 was linearized with NotI and introduced into ATCC 43037 by electroporation to generate the NTF1 strain.

The T. forsythia porT deletion mutant (NTF2) was constructed as described above except that the DNA regions upstream and downstream of porT were amplified by PCR from the chromosomal DNA of the strain ATCC 43037 with the primers TFPorTU and TFPorTD and the primers TFPorTF and TFPorTD, respectively. The T. forsythia sov insertion mutant was constructed as follows. A 2.0 kb internal region of the sov gene was amplified from the chromosomal DNA of the strain ATCC 43037 by PCR with the primers TFsovF and TFsovR. The amplified DNA was cloned into pCR4 Blunt TOPO to generate pKD1036. The 1.1 kb BamHI ermF DNA cassette was inserted into the BamHI site in the sov region of pKD1036, resulting in pKD1037 (sov::ermF). The pKD1037 was linearized with NotI and introduced into T. forsythia ATCC 43037 by electroporation to generate the NTF3 strain.

**Electron microscopy.** To examine bacterial cell shape, the cells were washed and negatively stained on carbon-coated grids with 1% ammonium molybdate. To prepare ultrathin sections, the cells were fixed with 2% paraformaldehyde and 5% glutaraldehyde in 30 mM HEPES buffer (pH 7.4) overnight at 4°C. The samples were post-fixed with 1% osmium tetroxide for 2 h and then with 0.5% uranyl acetate for 30 min. The fixed cells were dehydrated in a series of

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### Table 1. Bacterial strains and plasmids used in this study

<table>
<thead>
<tr>
<th>Strain or plasmid</th>
<th>Description</th>
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<tr>
<td><strong>Bacterial strains</strong></td>
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<td>43037</td>
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<td><strong>Plasmids</strong></td>
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Em’, erythromycin resistance; Ap’, ampicillin resistance; Cp’, chloramphenicol resistance; Km’, kanamycin resistance.
25–100% ethanol and embedded in Quetol-651 resin (Nishin EM). The ultrathin sections were stained with 1% uranyl acetate and 1% lead citrate. The stained samples (bacterial cells and ultrathin sections) were observed using a JEM-1210 transmission electron microscope (JEOL).

**Gel electrophoresis and immunoblot analysis.** SDS-PAGE and immunoblot analyses were performed as previously described (Shoji et al., 2011). The blotted membranes were treated with anti-TfA and anti-TfB antisera (Sakakibara et al., 2007).

The glycoproteins in SDS-PAGE gels were stained using the Pro-Q Emerald 300 fluorescent stain (Invitrogen). After staining with Pro-Q Emerald 300, total protein staining was performed with SYPRO Ruby (Invitrogen).

**Two-dimensional gel electrophoresis (2D-PAGE).** 2D-PAGE was performed as described previously (Sato et al., 2013). T. forsythia strains were grown in serum-free medium. Particle-free culture supernatants were obtained as previously described (Sato et al., 2013). The proteins in the particle-free culture supernatant fraction were precipitated with 10% (w/v) trichloroacetic acid at 4°C. The precipitated proteins were harvested by centrifugation at 4°C for 20 min, washed three times with cold diethyl ether, dried at room temperature for 30 min and then resuspended in a cell lysis solution (7 M urea, 2 M thiourea, 4% CHAPS, 1 mM EDTA and 5 mM tributylphosphine). The samples were applied to 13 cm immobilized pH gradient strips (GE Healthcare Bio-Sciences) with a pH range from 4 to 7 (first dimension) swollen with a rehydration solution (7 M urea, 2 M thiourea, 4%, w/v, CHAPS, 0.5%, v/v, IPG buffer pH 4 to 7 (GE Healthcare), 1 mM EDTA, 12 μl ml⁻¹ destreak reagent (GE Healthcare)) and bromophenol blue. The 2D electrophoresis (SDS-PAGE) was performed in polyacrylamide gels, and the proteins were stained with Coomassie brilliant blue (CBB) R250.

**MS analysis and database search for protein identification.** Proteins were identified by peptide-mass fingerprinting after in-gel trypsic digestion as previously described (Sato et al., 2010). Gel plugs containing proteins were subjected to washing with 50% (v/v) acetonitrile, washing with 100% acetonitrile, reduction with 10 mM DTT (Wako), alkylation with 55 mM iodoacetamide, washing/washout with 50% (v/v) acetonitrile and digestion for 10 h with 10 mM DTT, iodoacetamide (Wako), SDS (Wako), CaCl₂ (Sigma-Aldrich), MgCl₂ (Wako) and ZnCl₂ (Wako). The resulting peptides were extracted from the gel plugs with 0.1% (v/v) trifluoroacetic acid and 50% (v/v) acetonitrile, with 10 μl of 150 mM Tris/HCl buffer (pH 7.8), 50 μl of 4 mM BAPNA and 125 μl distilled water, and the assay mixtures were incubated at 37°C for 2 h. The release of p-nitroaniline was determined by measuring the OD₄₅₀ nm using a microplate reader (Bio-Rad).

**Biofilm formation.** Biofilm formation was measured by a microtitre plate biofilm assay using a previously reported protocol with slight modification (O’Toole & Kolter, 1998; Honma et al., 2007). Briefly, an overnight culture adjusted to an OD₅₉₅ nm of 1.0 was diluted 1:10 with fresh medium. The cells were aliquoted into the wells of a 96-well microtitre plate (250 μl per well) and incubated anaerobically for 1–4 days. After removal of the planktonic cells by washing twice with PBS, the biofilm was stained by incubation with 100 μl of 0.1% (w/v) crystal violet solution for 5 min. The plate was washed twice with distilled water and destained with 200 μl of 95% (v/v) ethanol for 5 min. Biofilm mass was evaluated at OD₅₉₅ nm at 1, 2, 3, and 4 days using a microplate reader. Total biofilm formation was evaluated as the absorbance of crystal violet stained biofilms at OD₅₉₅ nm divided by the absorbance of total growth (including biofilm and planktonic cells) at OD₅₉₅ nm.

For visualization by microscopy, T. forsythia biofilms were formed in 4-well Lab Tek II chamber slides (Nunc) as described previously (Honma et al., 2007). The diluted cells described above were dispensed in each chamber and incubated anaerobically for 3 days. Following incubation, chambers were washed with PBS and biofilms were stained with the SYTO9 LIVE/DEAD BacLight Bacterial Viability kit (Life Technologies). The stain was prepared according the manufacturer’s instruction and coverslips mounted using Vector Shield (Vector Laboratories). The edges of the coverslips were coated with nail polish and the slide was kept in the dark until analysed. Microscope images of fluorescence were examined using ZEISS Axioskop plus microscope mounted with an AxioCam HRc camera with a Plan-Neofluar 20× 0.5 NA objective (ZEISS). The Z-stack images of the biofilm were acquired with confocal laser scanning microscope (LSM510 META, Axiovert200M) with a LD-Achroplan 40× 0.6 NA objective (ZEISS). For three-dimensional reconstruction, the image analysis was done using Z-series image stacks of each biofilm with the LSM image browser and ZEN2009 software.

**Statistical analysis.** The data are presented as means ± SD. The biofilm formation data were analysed using one-way ANOVA with the Tukey–Kramer multiple-comparison test. P-values less than 0.05 were considered significant.

**RESULTS**

**Construction of T. forsythia mutants deficient in T9SS proteins.**

The erythromycin-resistance DNA cassette was inserted into the T. forsythia CDSs bfor_c_1_3635, bfor_c_1_6468 and bfor_c_1_12435 (locus tags by the Human Oral Microbiome Database), which were orthologous to P. gingivalis porK, porT and sov, to generate T. forsythia porK (NTF1), porT (NTF2) and sov (NTF3) mutants, respectively (Fig. S1).

**Cell morphology of the porK, porT and sov mutants.**

The bacterial cells were negatively stained with ammonium molybdate and analysed by electron microscopy. WT T. forsythia cells were prolate, ellipsoid-like with sharp ends,
whereas the porK, porT and sov mutants were rod-like with round ends (Fig. 1a). The WT cells showed a lattice structure on the cell surface, whereas the porK, porT and sov mutants exhibited amorphous and fragile surfaces. The lattice structure of the WT T. forsythia cell surface is the S-layer, which contains the TfsA and TfsB proteins (Sakakibara et al., 2007; Sekot et al., 2012); therefore, the porK, porT and sov mutants appeared to lack the S-layer. Analysis of the ultrathin cross-sections revealed that the WT cells had an S-layer with a thickness of approximately 20 nm on the outer membrane, whereas the porK mutant cells did not possess this structure on the outer membrane (Fig. 1b).

SDS-PAGE and immunoblot analyses of whole-cell lysates

The SDS-PAGE profiles of whole-cell lysates revealed that major proteins with molecular masses of 230 and 270 kDa in the WT cells were not present in the porK, porT or sov mutant cells; furthermore, the porK, porT and sov mutant cells contained major proteins with molecular masses of 165 and 205 kDa that were absent in the WT cells (Fig. 2). Immunoblotting analysis using antisera against the TfsA and TfsB proteins revealed that the 230 and 270 kDa proteins in the WT cells were the TfsA and TfsB proteins, respectively, and that the 165 and 205 kDa proteins in the porK, porT and sov mutants were derived from the TfsA and TfsB proteins, respectively. The molecular masses of the TfsA and TfsB proteins without signal peptides are 133.3 kDa and 150.8 kDa, respectively (Lee et al., 2006); therefore, it was examined whether these proteins were glycosylated (Fig. 3). ProQ-Emerald carbohydrate staining analysis suggested that the 230 kDa TfsA protein and the 270 kDa TfsB protein were strongly glycosylated in WT cells and that the 165 kDa TfsA protein and 205 kDa TfsB protein were also glycosylated in the porK, porT and sov mutants, albeit to a lesser extent.

2D-PAGE analysis of particle-free culture supernatants

2D-PAGE was performed to analyse the particle-free (membrane-free) culture supernatants from the WT and porK strains (Fig. 4). Time of flight mass spectrometry (TOFMS) analysis revealed the presence of the T. forsythia proteins bfor_c_1_1931 (tentatively named type Nine Secretion System-dependent protein A, NdpA), bfor_c_1_8519 (NdpB), bfor_c_1_10593 (NdpC), bfor_c_1_10600 (NdpD) and bfor_c_1_14540 (NdpE) in the particle-free culture supernatant of the WT cells but not the porK mutant cells (Table 2).

Haemagglutination

Purified S-layers cause erythrocyte agglutination, and S-layer-deficient mutant strains exhibit decreased haemagglutination activity (Sabet et al., 2003; Sakakibara et al., 2007). The haemagglutination activities of the porK, porT and sov mutants on sheep erythrocytes were measured (Fig. 5). The haemagglutination activities of the porK, porT and sov mutants were 12.5 % lower compared with the WT strain. The haemagglutination activities of the tfsA and tfsB single mutants were 50 % of the WT level, whereas the haemagglutination activity of the tfsA tfsB double mutant cells was 25 % of the WT level.
Biofilm formation

The T. forsythia wecC mutant strain, which lacks UDP-N-acetylmannosaminuronic acid dehydrogenase and has truncated S-layer glycans, shows increased biofilm formation (Honma et al., 2007; Posch et al., 2011). An examination of the biofilm-forming ability of the porK, porT and sov mutants (Figs 6 and 7) revealed that these mutants showed increased biofilm formation.

Trypsin-like activity

T. forsythia produces an enzymic endopeptidase that degrades BAPNA; this endopeptidase activity was attributed to a trypsin-like proteinase (Grenier, 1995). To examine whether the porK, porT and sov mutations affect the trypsin-like endopeptidase activity of T. forsythia, BAPNA hydrolysis using porK, porT and sov mutant cells in the presence and absence of various chemicals including protease inhibitors was monitored (Table 3). The hydrolysis activities of the porK, porT and sov mutant cells were similar to that of the WT. The BAPNA-hydrolysis activities of the mutant and WT cells were completely or partially suppressed by TLCK, ZnCl₂ and leupeptin, indicating that the WT and mutant cells contained similar BAPNA-hydrolysis properties.

DISCUSSION

The T9SS (PorSS) was discovered in the periodontal pathogen P. gingivalis (Sato et al., 2010). Subsequently, homologous
genes encoding putative T9SS components were observed in several bacterial species of phylum Bacteroidetes, suggesting that the T9SS is conserved in at least a subset of this phylum (McBride & Zhu, 2013; Chagnot et al., 2013). The T9SS is related to gliding motility of bacteria of phylum Bacteroidetes (Sato et al., 2010; Nakane et al., 2013).

T. forsythia, which belongs to phylum Bacteroidetes, is a member of the ‘red complex’ together with P. gingivalis and T. denticola and is considered a major pathogen underlying periodontal disease. T. forsythia possesses several putative virulence factors such as trypsin-like protease, PrtH protease, sialidases, BspA leucine-rich repeat protein and the S-layer. In this study, we generated three T9SS-deficient T. forsythia mutants in which the porK, porT and sov genes were mutated. All mutations caused identical phenotypes such as absence of the S-layer, decreased haemagglutination activity and increased biofilm formation, suggesting that these properties are related to the T9SS.

The S-layer is a paracrystalline surface-protein array expressed in several bacteria and is thought to function as a protective coat against external sieves and ion traps (Sleytr & Beveridge, 1999; Sabet et al., 2003; Messner et al., 2010). The T. forsythia S-layer mediates adhesion to human gingival epithelial cells and subsequent invasion (Sakakibara et al., 2007) and delays recognition of the bacterium by the host innate immune

### Table 2. Identification of protein spots in 2D-gels

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<th>HOMD name</th>
<th>Description</th>
<th>Mascot score</th>
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<td>a</td>
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<td>Hypothetical protein</td>
<td>WT 261, NTF1 616</td>
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HOMD, Human Oral Microbiome Database.

*CTD family protein (Veith et al., 2009).

†Probable CTD family protein.

§Antigenic protein (non-CTD) (Veith et al., 2009).

### Fig. 5. Haemaggulutination. Suspensions of T. forsythia cells and serial twofold dilutions in PBS were applied to the wells of a microtitre plate from left to right and mixed with a sheep erythrocyte suspension.
The S-layer contributes to *T. forsythia* serum resistance and oral bacterial coaggregation (Shimotahira et al., 2013) and consists of the TfsA and TfsB proteins. S-layer proteins generally contain N-terminal signal peptides, with a few exceptions (Boot & Pouwels, 1996). Because the primary products of the *tfsA* and *tfsB* CDSs contain N-terminal signal peptides, these proteins are probably translocated across the inner membrane by a Sec-dependent mechanism. The TfsA and TfsB proteins contain CTD-like sequences at the C terminus, and CTD sequences are a signal for T9SS-mediated translocation across the outer membrane (Shoji et al., 2011), suggesting that the S-layer proteins are translocated across the outer membrane by the T9SS. Posch et al. (2013) observed that the molecular masses of His-tagged TfsA and TfsB proteins expressed in *Bacteroides fragilis*, which are probably located in the periplasm because of lack of T9SS in *B. fragilis* (Sato et al., 2010; McBride & Zhu, 2013), are ~170 kDa and ~200 kDa, respectively, which are consistent with the molecular masses of the TfsA and TfsB proteins in the *T. forsythia* T9SS-deficient mutants. *T. forsythia* as well as *B. fragilis* has an O-glycosylation system (Fletcher et al., 2009; Coyne et al., 2013; Posch et al., 2011, 2013). These findings suggest that in *T. forsythia* cells, the S-layer proteins are primarily O-glycosylated at the inner membrane and/or in the periplasm, and after translocation across the outer membrane by the T9SS, the proteins are further glycosylated on the cell surface.

Using 2D-gel analysis, we observed that the NdpA, NdpB, NdpC, NdpD and NdpE proteins were released into the WT but not the *porK* culture supernatant. The NdpA and NdpE proteins are thermolysin metallopeptidase homologues; the NdpB protein is a PorU homologue (Sato et al., 2010; Glew et al., 2012); the NdpC protein is a karilysin (Karim et al., 2010) and the NdpD protein is a putative lysyl endopeptidase homologous to *P. gingivalis* PepK (Nonaka et al., 2014). These proteins are putative peptidases and possess CTD-like sequences at their C termini. These results indicate that the T9SS is functional in *T. forsythia* and is involved in the secretion of CTD proteins.

Sabet et al. (2003) purified the S-layer from *T. forsythia* and observed that the S-layer was sufficient to mediate the haemagglutination of sheep erythrocytes. Sakakibara et al. (2007) generated *tfsA* and *tfsB* single mutants and a *tfsA* mutant.*
Enzymic activity of the WT without any additive in the reaction mixture was taken as 100%.

*Trypsin-like activities of various S-layer glycans (Posch et al., 2011). In this study, it was observed that the porK, porT and sov mutants caused decreased haemagglutination of chicken erythrocytes. The haemagglutination activity of the T9SS-deficient mutants was weaker than that of the S-layer-deficient mutants, suggesting that cell-surface proteins other than TfsA and TfsB, which are secreted by the T9SS, are also involved in haemagglutination.

Honma et al. (2007) isolated a wecC mutant that showed increased biofilm formation and observed that the molecular masses of both S-layer proteins were decreased in the wecC mutant. Subsequently, the decreased molecular mass of the S-layer proteins was correlated with truncated S-layer glycans (Posch et al., 2011). In this study, it was observed that the T9SS-deficient mutants lacked S-layers and contained TfsA and TfsB proteins with decreased molecular masses and reduced glycosylation compared with WT cells; these mutants also showed increased biofilm formation. These results indicate that S-layers or S-layer glycans suppress T. forsythia biofilm formation.

Trypsin-like endopeptidase activity was observed in the T. forsythia cell envelope (Grenier, 1995). The T9SS-deficient mutants showed the same trypsin-like activity as the WT, suggesting that the T9SS is not required for translocation of the trypsin-like enzyme(s) to the cell envelope.

In this study, T. forsythia porK, porT and sov mutant strains were generated, and these mutants were found to lack the S-layer. Several CTD proteins such as thermolysin were not observed in the culture supernatant of the porK mutant cells. These results indicate that the T9SS is functional in T. forsythia and contributes to translocation of the CTD proteins to the cell surface or into the extracellular milieu.

Table 3. Trypsin-like activity

<table>
<thead>
<tr>
<th>Inhibitor</th>
<th>Conc. (final)</th>
<th>Residual activity (%)*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>WT</td>
</tr>
<tr>
<td>None</td>
<td>–</td>
<td>100</td>
</tr>
<tr>
<td>EDTA</td>
<td>10 mM</td>
<td>107.6</td>
</tr>
<tr>
<td>Iodoacetamide</td>
<td>10 mM</td>
<td>113.7</td>
</tr>
<tr>
<td>TLCK</td>
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<td>1.3</td>
</tr>
<tr>
<td>Leupeptin</td>
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<td>57.5</td>
</tr>
<tr>
<td>CaCl₂</td>
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<td>87.3</td>
</tr>
<tr>
<td>ZnCl₂</td>
<td>10 mM</td>
<td>28.6</td>
</tr>
<tr>
<td>DTT</td>
<td>10 mM</td>
<td>112.3</td>
</tr>
</tbody>
</table>

*Trypsin-like activities of various T. forsythia strains with various protease inhibitors and metal ions were determined using BAPNA as a substrate. Enzymic activity of the WT without any additive in the reaction mixture was taken as 100%.

REFERENCES


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