Outer-membrane cytochrome-independent reduction of extracellular electron acceptors in \textit{Shewanella oneidensis}

Clemens Bücking, Annette Piepenbrock, Andreas Kappler and Johannes Gescher

**Correspondence**
Johannes Gescher
johannes.gescher@kit.edu

1Institut für angewandte Biowissenschaften, Angewandte Biologie, Karlsruher Institut für Technologie, Fritz-Haber-Weg 2, D-76131 Karlsruhe, Germany
2Geomicrobiology, Center for Applied Geosciences, University of Tübingen, Sigwartstrasse 10, D-72076 Tübingen, Germany

Dissimilatory metal reduction under pH-neutral conditions is dependent on an extended respiratory chain to the cell surface. The final reduction is catalysed by outer-membrane cytochromes that transfer respiratory electrons either directly to mineral surfaces and metal ions bound in larger organic complexes such as Fe(III) citrate, or indirectly using endogenous or exogenous electron shuttles such as humic acids and flavins. Consequently, a \textit{Shewanella oneidensis} deletion mutant devoid of outer-membrane cytochromes is unable to reduce Fe(III) citrate or manganese oxide minerals and reduces humic acids at lower rates. Surprisingly, the phenotype of this quintuple deletion mutant can be rescued by a suppressor mutation, which enables metal or humic acid reduction without any outer-membrane cytochrome. Furthermore, the type II secretion system, essential for metal reduction in wild-type \textit{S. oneidensis}, is not necessary for the suppressor strain. Using genome sequencing we identified two point mutations in key genes for metal reduction: \textit{mtrA} and \textit{mtrB}. These mutations are necessary and sufficient to account for the observed phenotype. This study is the first evidence for a catabolic, outer-membrane cytochrome-independent electron transport chain to ferric iron, manganese oxides and humic acid analogues operating in a mesophilic organism. Available bioinformatic data allow the hypothesis that outer-membrane cytochrome-independent electron transfer might resemble an evolutionary intermediate between ferrous iron-oxidizing and ferric iron-reducing micro-organisms.

**INTRODUCTION**

Bacterial dissimilatory metal reduction has been investigated intensely in the last few decades. This is due to its relevance as a respiratory process in a multitude of environments, and the possible applications in biotechnology such as remediation of contaminated soils and electricity production in microbial fuel cells. One of the best-studied model organisms is \textit{Shewanella oneidensis} MR-1, a $\gamma$-proteobacterium with 41 putative $c$-type cytochromes encoded in its genome (Meyer et al., 2004; Romine et al., 2008).

Several of these $c$-type cytochromes have been shown to form a respiratory network extending the respiratory chain from the cytoplasmic membrane through the periplasm to the outer membrane (Schuetz et al., 2009; Shi et al., 2007).

**Abbreviations:** AQDS, anthraquinone-2,6-disulfonate; NTA, nitrilotriacetic acid; PPHA, Pahokee peat humic acids.

Five supplementary figures and two supplementary tables are available with the online version of this paper.

From there, catabolic electrons are transferred directly or indirectly to a wide variety of terminal electron acceptors that are either poorly soluble or soluble but unable to pass the outer membrane. Examples of poorly soluble electron acceptors are iron(III) (oxyhydr)oxides such as haematite and ferrihydrite, and manganese oxides such as birnessite and pyrolusite. These minerals are ubiquitously distributed in soils and sediments. Examples of substances that are soluble but apparently unable to pass the outer membrane at least to an extent that supports growth are Fe(III) citrate and the humic acid analogue anthraquinone-2,6-disulfonate (Dobbin et al., 1996; Gescher et al., 2008; Pitts et al., 2003; Shyu et al., 2002).

In \textit{S. oneidensis}, the reduction of most extracellular electron acceptors is based on a protein complex formed by MtrA, MtrB and MtrC (mtr: metal reducing). This complex is most probably the extracellular terminal reductase (Hartshorne et al., 2009; Shi et al., 2006). The current model is that the decahaem $c$-type cytochromes MtrA, localized to the periplasmic side, and MtrC, localized to the outer surface of
the outer membrane, are held in close proximity via the outer membrane-spanning β-barrel protein MtrB. This close proximity is thought to enable electron transfer through the outer membrane (Hartshorne et al., 2009). OmcA (outer membrane cytochrome), another decahaem c-type cytochrome, is also localized to the cell surface and apparently involved in the reduction of manganese oxides (Bücking et al., 2010; Myers & Myers, 2003). Although in vitro evidence has been provided for a high-affinity complex between OmcA and MtrC, it seems that they are not colocalized in vivo (Lower et al., 2009; Shi et al., 2006; Zhang et al., 2008). Both proteins are lipoproteins that are exported through the outer membrane by the type II secretion system (Shi et al., 2008).

Different models have been proposed of how the terminal metallic electron acceptor is finally reduced. MtrC and OmcA have been shown to directly reduce iron minerals in vitro (Realdon et al., 2010; Xiong et al., 2006). The transfer rates to ferric iron are rather low and cannot explain entirely the fast rates of iron reduction by whole cells. It has therefore been proposed that endogenous flavin redox shuttles are involved in this electron transfer process (Marsili et al., 2008; Ross et al., 2009; von Canstein et al., 2008). However, reduction of the flavin shuttle molecules depends on the whole electron transfer chain, since they apparently cannot pass the outer membrane and therefore also have to be reduced by outer-membrane cytochromes (Coursolle et al., 2010; Richter et al., 2010). In the aquatic sediments from which S. oneidensis MR-1 was originally isolated, humic acids are frequently present (Myers & Nealson, 1988a; Stevenson, 1994). They represent exogenous electron shuttles. Consequently, it was shown that Fe(III) reduction by S. oneidensis was stimulated by humic substances at concentrations as low as 5–10 mg carbon l−1. Humic substances can occur in dissolved or solid-phase form. Both have been studied extensively with respect to their effect on microbial Fe(III) reduction due to electron shuttling (Jiang & Kappler, 2008; Lovley et al., 1996; Roden et al., 2010).

Another possible mode of electron transfer is represented by so-called nanowires, which were first described by Reguera and co-workers for Geobacter sulfurreducens and shortly afterwards by Gorby and co-workers for S. oneidensis (Gorby et al., 2006; Reguera et al., 2005). It was hypothesized that these cell appendages can transfer electrons to an electron acceptor that is not in direct contact with the cell but rather located distantly from it. In S. oneidensis, electron transfer along the pilus is dependent on the presence of MtrC and OmcA (El-Naggar et al., 2010). Probably, all three modes of electron transfer (i.e. direct contact, electron shuttling and nanowires) operate simultaneously, but all of them fundamentally rely on the catalytic activity of outer-membrane c-type cytochromes.

In this study we report that a quintuple deletion mutant devoid of genes for outer-membrane cytochromes (Bücking et al., 2010) can regain the ability to reduce extracellular electron acceptors such as ferric iron, manganese dioxide, humic acids and anthraquinon-2,6-disulfonate (AQDS), a model compound for quinone moieties in humic acids. This mutant is not affected in outer-membrane stability, which excludes an increased reduction of extracellular electron acceptors by periplasmic c-type cytochromes. Resequencing of the mutant genome and subsequent mapping to the S. oneidensis reference genome revealed that one point mutation in MtrA and one in MtrB are the reasons for the evolution of an extended respiratory chain independent of outer-membrane cytochromes.

**METHODS**

**Reagents.** Chemicals and biochemicals were obtained from Sigma-Aldrich, Roth and Promega. Enzymes were purchased from New England Biolabs.

**Growth conditions and media.** All micro-organisms used in this study are listed in Table 1. *Escherichia coli* strains were grown aerobically in Luria–Bertani (LB) medium at 37 °C. *Saccharomyces cerevisiae* InvSc1 was grown on YPD medium and was selected for transformants on uracil-free medium purchased from Clontech. *S. oneidensis* strains were grown aerobically at 30 °C in LB medium or in minimal medium under anoxic conditions, supplemented with lactate (50 mM) as electron donor and carbon source, and 50 mM Fe(III) citrate, 5 mM Fe(III) nitritotriacetic acid (NTA) or 100 mM fumarate as electron acceptor, as described previously (Schuetz et al., 2009). Fe(III) citrate powder was dissolved in hot water and pH was adjusted to pH 7.4 slowly with 1 M NaOH. If necessary, kanamycin (50 μg ml−1), arabinose (1 mM) or 2,6-diaminopimelic acid (100 μg ml−1) was added to the medium. Fe(III) citrate and NTA-complexed ferric iron [Fe(III)-NTA] reduction was determined by measuring ferrous iron concentrations with the ferrozine reagent (Stokey, 1970), and was used as a surrogate for bacterial growth. For growth experiments with birnessite (manganese dioxide) as electron acceptor, 2.5 mM birnessite was added to minimal medium that was supplemented with 1 mM arabinose. Birnessite was prepared as described earlier (Burridge & Nealson, 1985). Manganese reduction was determined in three independent cultures using leucoberellin blue (Boogerd & de Vrind, 1987). The rate of abiotic Mn(IV) decrease in culture flasks was subtracted from the measured values.

**Standard procedures.** Membrane fractions were prepared as described previously (Schuetz et al., 2009). Protein concentrations were determined by the method of Bradford (1976) with BSA as standard. Proteins were separated on polyacrylamide gels according to Laemmli (1970). Haem proteins were visualized by peroxidase staining (Thomas et al., 1976). Proteins containing a C-terminal strep-tag were detected on a Western blot using a strep-tag antibody (Qiagen).

**Construction of markerless S. oneidensis mutants.** Strains containing ΔgspD, ΔdmsA-1 dmsB-1 and ΔdmsA-2 dmsB-2 deletions were constructed according to Schuetz et al. (2009), with minor modifications. All primers used are listed in Table S1. Primers 1–4 were used to amplify 500 bp regions up- and downstream of gspD. The resulting fragments were cloned into a linearized pMQ150 plasmid using a *Saccharomyces cerevisiae*-based recombination system (Shanks et al., 2006). The resulting plasmid pMQ150ΔgspD was transformed into *E. coli* WM3064, which was then used as donor strain for conjugation-based mating with *S. oneidensis*. Primers 5–8 were used to construct pMQ150ΔdmsA-1 dmsB-1, and primers 9–12 were used to assemble pMQ150ΔdmsA-2 dmsB-2.

**Genome sequencing.** Genomic DNA was isolated using the Illustra *Saccharomyces cerevisiae* InvSc1 was grown on YPD medium and was selected for transformants on uracil-free medium purchased from Clontech. *S. cerevisiae* InvSc1 was grown on YPD medium and was selected for transformants on uracil-free medium purchased from Clontech. *S. oneidensis* MR-1 was originally isolated, humic acids are frequently present (Myers & Nealson, 1988a; Stevenson, 1994). They represent exogenous electron shuttles. Consequently, it was shown that Fe(III) reduction by S. oneidensis was stimulated by humic substances at concentrations as low as 5–10 mg carbon l−1. Humic substances can occur in dissolved or solid-phase form. Both have been studied extensively with respect to their effect on microbial Fe(III) reduction due to electron shuttling (Jiang & Kappler, 2008; Lovley et al., 1996; Roden et al., 2010).

Another possible mode of electron transfer is represented by so-called nanowires, which were first described by Reguera and co-workers for Geobacter sulfurreducens and shortly afterwards by Gorby and co-workers for S. oneidensis (Gorby et al., 2006; Reguera et al., 2005). It was hypothesized that these cell appendages can transfer electrons to an electron acceptor that is not in direct contact with the cell but rather located distantly from it. In S. oneidensis, electron transfer along the pilus is dependent on the presence of MtrC and OmcA (El-Naggar et al., 2010). Probably, all three modes of electron transfer (i.e. direct contact, electron shuttling and nanowires) operate simultaneously, but all of them fundamentally rely on the catalytic activity of outer-membrane c-type cytochromes.

In this study we report that a quintuple deletion mutant devoid of genes for outer-membrane cytochromes (Bücking et al., 2010) can regain the ability to reduce extracellular electron acceptors such as ferric iron, manganese dioxide, humic acids and anthraquinon-2,6-disulfonate (AQDS), a model compound for quinone moieties in humic acids. This mutant is not affected in outer-membrane stability, which excludes an increased reduction of extracellular electron acceptors by periplasmic c-type cytochromes. Resequencing of the mutant genome and subsequent mapping to the S. oneidensis reference genome revealed that one point mutation in MtrA and one in MtrB are the reasons for the evolution of an extended respiratory chain independent of outer-membrane cytochromes.
instructions. Sequencing was performed on an Illumina HiSeq 2000 with a read length of 46 bp by GATC Biotech. The reads were assembled using the software BWA (Li & Durbin, 2009), and identification of single nucleotide polymorphisms as well as insertions and deletions was done with SAMtools (Li et al., 2009) and the assembly viewer Tablet (Milne et al., 2010).

**Cloning of mtrA and mtrB and their mutated versions.** Genomic DNA of either *S. oneidensis* wild-type or ΔOMCΔ was used as a template to amplify *mtrA* (primers 13, 14), *mtrB* (primers 15, 16) or *mtrA* and *mtrB* (primers 13, 16). Primer 16 contains furthermore the sequence for a strep-tag, which was added to the C terminus of MtrB.

**Riboflavin and AQDS reduction rates.** Cells for these assays were grown under anoxic conditions with fumarate as electron acceptor and, if necessary, in the presence of arabinose as inducer for gene expression. Cells were harvested and washed twice before the start of the experiments. Riboflavin reduction was measured in cuvettes with butyl rubber stoppers at 30 °C and 450 nm. The assay contained 60 μM riboflavin, 100 mM HEPES, pH 7.4 and 10 mM lactate, and was initiated by the addition of riboflavin. Reduction rates were calculated as a function of protein concentration in the assay. AQDS reduction was measured in a similar assay at a wavelength of 436 nm using 1 mM AQDS (molar absorption coefficient 3500 M⁻¹ cm⁻¹). Both assays were typically finished within 20 min.

**Humic acid reduction rates.** Humic acid reduction was measured continuously over 18 days. Pahokee peat humic acids (PPHA), purchased from the International Humic Substances Society (IHSS), were added at a concentration of 1 g l⁻¹ with 2 mM lactate as electron donor. At selected time points, samples were withdrawn and the oxidation state of humic acids was analysed by an electron shuttling assay similar to that described earlier (Lovley et al., 1996). Briefly, under anoxic conditions, samples were filtered (0.2 μm) and reacted with 5 mM Fe(III) citrate for 1 h. Subsequently, the sample/Fe(III) citrate mixture was diluted 1:2 with 1 M HCl. Humic acids were precipitated and removed by centrifugation, and the Fe(II) content of the cell suspension. Fifty microlitres of cell suspension was pipetted into a well of a microtitre plate. The assay was started by the addition of 150 μl of a solution containing 10 mM lactate and 10 mM Fe(III) citrate. At different time points (0–30 min) the reaction was stopped by the addition of 100 μl 3 M HCl. The Fe[^{2+}] concentration of the samples was determined using the ferrozine reagent (Stokey, 1970).

**Ferric iron reduction cell suspension assays.** Cells were grown under anoxic conditions overnight in minimal medium with fumarate as electron acceptor, and with 1 mM arabinose to induce *mtrA* and *mtrB* expression (*mtrA* and *mtrB* are under arabinose promoter control in ΔOMC cells) (Bücking et al., 2010). Cells were harvested and washed twice using minimal medium without fumarate and lactate under anoxic conditions, and resuspended to a final OD₆₀₀ between 3 and 5. Specific reduction rates of independent triplicate cultures were obtained by normalization to the protein content of the cell suspension. Fifty microlitres of cell suspension was pipetted into a well of a microtitre plate. The assay was started by the addition of 150 μl of a solution containing 10 mM lactate and 10 mM Fe(III) citrate. At different time points (0–30 min) the reaction was stopped by the addition of 100 μl 3 M HCl. The Fe[^{2+}] concentration of the samples was determined using the ferrozine reagent (Stokey, 1970).

**Riboflavin and AQDS reduction rates.** Cells for these assays were grown under anoxic conditions with fumarate as electron acceptor and, if necessary, in the presence of arabinose as inducer for gene expression. Cells were harvested and washed twice before the start of the experiments. Riboflavin reduction was measured in cuvettes with butyl rubber stoppers at 30 °C and 450 nm. The assay contained 60 μM riboflavin, 100 mM HEPES, pH 7.4 and 10 mM lactate, and was initiated by the addition of riboflavin. Reduction rates were calculated as a function of protein concentration in the assay. AQDS reduction was measured in a similar assay at a wavelength of 436 nm using 1 mM AQDS (molar absorption coefficient 3500 M⁻¹ cm⁻¹). Both assays were typically finished within 20 min.

**Humic acid reduction rates.** Humic acid reduction was measured continuously over 18 days. Pahokee peat humic acids (PPHA), purchased from the International Humic Substances Society (IHSS), were added at a concentration of 1 g l⁻¹ with 2 mM lactate as electron donor. At selected time points, samples were withdrawn and the oxidation state of humic acids was analysed by an electron shuttling assay similar to that described earlier (Lovley et al., 1996). Briefly, under anoxic conditions, samples were filtered (0.2 μm) and reacted with 5 mM Fe(III) citrate for 1 h. Subsequently, the sample/Fe(III) citrate mixture was diluted 1:2 with 1 M HCl. Humic acids were precipitated and removed by centrifugation, and the Fe(II)

### Table 1. Yeast and bacterial strains used in this study

<table>
<thead>
<tr>
<th>Strain no.</th>
<th>Strain</th>
<th>Relevant genotype</th>
<th>Reference or source</th>
</tr>
</thead>
<tbody>
<tr>
<td>JG26</td>
<td><em>Saccharomyces cerevisiae</em> InvSc1</td>
<td>MATa/MATα leu2-3,12,1 ura3-52-2 his3-Δ1/1 his3-Δ1</td>
<td>Invitrogen</td>
</tr>
<tr>
<td>JG7</td>
<td><em>S. oneidensis</em> MR-1</td>
<td>Wild-type</td>
<td>Venkateswaran et al. (1999)</td>
</tr>
<tr>
<td>JG132</td>
<td><em>S. oneidensis</em> MR-1 ΔOMC (outer-membrane cytochrome deletion mutant)</td>
<td>Δ(mtrD-mtrC) ΔSO_2931 ΔSO_1659, 3100633:: (araC, PBAD)</td>
<td>Bücking et al. (2010)</td>
</tr>
<tr>
<td>JG12</td>
<td><em>S. oneidensis</em> MR-1 ΔOMCΔ (suppressor of outer-membrane cytochrome deletion mutant)</td>
<td>ΔOMCΔ mtrA: position 869 C to A, mtrB: position 656 C to A</td>
<td>This study</td>
</tr>
<tr>
<td>JG202</td>
<td><em>E. coli</em> WM3064 pMQ150ΔgspD</td>
<td>pMQ150ΔgspD</td>
<td>This study</td>
</tr>
<tr>
<td>JG411</td>
<td><em>E. coli</em> WM3064 pMQ150ΔdmsA-1 dmsB-1</td>
<td>pMQ150ΔdmsA-1 dmsB-1</td>
<td>This study</td>
</tr>
<tr>
<td>JG305</td>
<td><em>E. coli</em> WM3064 pMQ150ΔdmsA-2 dmsB-2</td>
<td>pMQ150ΔdmsA-2 dmsB-2</td>
<td>This study</td>
</tr>
<tr>
<td>JG450</td>
<td><em>S. oneidensis</em> MR-1 ΔOMC ΔmtrA mtrB</td>
<td>pmtrA mtrB</td>
<td>This study</td>
</tr>
<tr>
<td>JG462</td>
<td><em>S. oneidensis</em> MR-1 ΔOMC ΔmtrAΔ</td>
<td>pmtrAΔ</td>
<td>This study</td>
</tr>
<tr>
<td>JG463</td>
<td><em>S. oneidensis</em> MR-1 ΔOMC ΔmtrBΔ</td>
<td>pmtrBΔ</td>
<td>This study</td>
</tr>
<tr>
<td>JG449</td>
<td><em>S. oneidensis</em> MR-1 ΔOMCΔmtrBΔ</td>
<td>pmtrAΔ mtrBΔ</td>
<td>This study</td>
</tr>
<tr>
<td>JG434</td>
<td><em>S. oneidensis</em> MR-1 ΔOMCΔΔmtrA-1 ΔdmsA-1 ΔdmsB-1 ΔdmsA-2</td>
<td>ΔdmsA-2</td>
<td>This study</td>
</tr>
<tr>
<td>JG444</td>
<td><em>S. oneidensis</em> MR-1 ΔOMCΔΔgspD</td>
<td>ΔOMCΔΔgspD</td>
<td>This study</td>
</tr>
<tr>
<td>JG443</td>
<td><em>S. oneidensis</em> MR-1 ΔgspD</td>
<td>ΔgspD</td>
<td>This study</td>
</tr>
<tr>
<td>JG52</td>
<td><em>S. oneidensis</em> MR-1 ΔmtrA</td>
<td>ΔmtrA</td>
<td>Schuetz et al. (2009)</td>
</tr>
<tr>
<td>JG55</td>
<td><em>S. oneidensis</em> MR-1 ΔmtrB</td>
<td>ΔmtrB</td>
<td>Schuetz et al. (2009)</td>
</tr>
<tr>
<td>JG468</td>
<td><em>S. oneidensis</em> MR-1 ΔmtrAΔΔmtrBΔ</td>
<td>ΔmtrAΔ mtrBΔ</td>
<td>This study</td>
</tr>
<tr>
<td>JG469</td>
<td><em>S. oneidensis</em> MR-1 ΔmtrBΔΔmtrBΔ</td>
<td>ΔmtrBΔ mtrBΔ</td>
<td>This study</td>
</tr>
<tr>
<td>JG474</td>
<td><em>S. oneidensis</em> MR-1 ΔOMCΔΔmtrAΔΔmtrBΔ</td>
<td>ΔOMCΔΔmtrAΔΔmtrBΔ</td>
<td>This study</td>
</tr>
</tbody>
</table>
concentration was determined by the ferrozine assay (Stokey, 1970). The reducing capacity in micro electron equivalents (μeq l⁻¹) was calculated as the amount of electrons transferred to Fe(III), i.e. the amount of Fe(II) formed. The amount of electrons transferred by the bacteria to humic acids was calculated by subtracting the abiotic reducing capacity of the sterile control from the reducing capacity of the humic acids at the respective time points. Experiments were performed in independent duplicates.

**Outer-membrane integrity testing.** For outer-membrane integrity testing, cells were grown under anaerobic conditions with fumarate as electron acceptor and in the presence of 1 mM arabinose. After washing the cells twice with 100 mM HEPES, pH 7.4, first an ethidium bromide influx assay was performed to measure outer-membrane permeability. The assay was performed according to Murata et al. (2007) with minor modifications. The assay was initiated by the addition of ethidium bromide (final concentration 6 μM). Fluorescence was measured continuously over 5 min (excitation and emission wavelengths of 545 and 600 nm, respectively) using a microtitre plate reader (Varioscan, ThermoFisher). Ethidium bromide influx rates are displayed as relative fluorescence units (RFU) per minute and milligram protein. As a positive control, 1 mg polymyxin B ml⁻¹ was added to the cells. Polymyxin B destabilizes the membrane and increases permeability (Vaara, 1992).

Second, iron diffusion into the cells was measured after 30 min incubation with 10 mM Fe(III) citrate but without the electron donor at room temperature. Afterwards, the cells were washed twice and acidified to a final concentration of 1 M HCl. All Fe(III) was reduced at room temperature. Afterwards, the cells were washed twice and incubation with 10 mM Fe(III) citrate but without the electron donor. Polymyxin B destabilizes the membrane and increases permeability (Vaara, 1992).

As a third test, antibiotic susceptibility was tested in a disc diffusion test. One hundred microlitres of cell suspension (OD₆₀₀ 0.132, corresponding to McFarland standard 0.5) was spread on LB plates supplemented with 1 mM arabinose, and subsequently paper discs containing 25 μg colistin, 30 μg kanamycin or 30 μg chloramphenicol (Oxoid, Thermo Fisher) were applied to the surface. After overnight incubation at 30 °C, plates were photographed and zones of inhibition were measured with ImageJ (http://imagej.nih.gov/ij/).

**Bioinformatic analysis.** The integrated microbial genomes tool of the Joint Genome Institute was used to search in available microbial genomes for homologues of mtrAB gene cassettes (http://img.jgi.doe.gov). The analysis was performed using the BLASTP algorithm and a concatenator of the protein sequences of MtrA and MtrB from *S. oneidensis*. Detected mtrAB-like gene clusters were analysed for genes encoded upstream and downstream of mtrA. Only gene clusters that did contain mtrAB gene clusters without genes homologous to mtrC or mtrF were further analysed.

### RESULTS

#### Evolution of a ΔOMC suppressor mutant

Previously we deleted in *S. oneidensis* all five genes that encode outer-membrane cytochromes. Additionally, a PBAD promoter was inserted in the genome of this strain upstream of mtrA and mtrB, two genes essential for metal reduction. The deletion of the outer-membrane cytochromes resulted in an almost complete loss of the ability to reduce poorly soluble electron acceptors (Bücking et al., 2010). In Fe(III) citrate medium, the Fe(II) concentration did not climb above 2.5 mM within 35 h, and a reduction rate for the ΔOMC strain could thus not be calculated (Fig. 1). However, this minimal Fe(III) citrate-reducing activity apparently increased substantially during prolonged incubation and several serial transfers. The accelerated iron reduction over time suggested the development of a suppressor mutation. In order to characterize a potential suppressor mutant, ΔOMC⁵, single cells were isolated using dilution series and subsequent plating on LB agar plates.

**Fe(III) citrate, Fe(III)-NTA and birnessite reduction without outer-membrane cytochromes**

It has been shown that *S. oneidensis* can couple growth to the reduction of ferric iron or manganese oxides (Myers & Nealson, 1988a, b). Therefore, ferrous iron concentrations were determined as a surrogate for growth in medium containing lactate as carbon and electron source and 50 mM Fe(III) citrate or 5 mM Fe(III)-NTA as electron acceptor. Iron reduction curves were recorded in the presence and absence of 1 mM arabinose as inducer for *mtrA* and *mtrB* expression (Fig. 1). Induction with 1 mM arabinose led to the expression of wild-type levels of MtrA, as determined by SDS-PAGE and staining of haem-containing proteins (Fig. S1). Of note, *S. oneidensis* cannot grow with arabinose as a substrate (Gralnick et al., 2006). As indicated in Fig. 1, Fe(III) citrate reduction of the suppressor mutant ΔOMC⁵ was strictly dependent on arabinose induction, upon which the reduction rate reached 84 % of the

![Fig. 1. Fe(III) citrate reduction. Iron reduction by *S. oneidensis* strains growing on minimal medium with 50 mM lactate and 50 mM Fe(III) citrate, and addition of 1 mM arabinose, where indicated. Symbols show means of triplicate measurements; error bars, ± SD. Solid lines represent curve-fits done with GraphPad Prism 4. The specific iron reduction rate, µ, in h⁻¹ was calculated for each biological replicate, and the mean ± SD values are: wild-type, 0.534 ± 0.045; ΔOMC⁵ + arabinose, 0.451 ± 0.104; reduction rates not determinable for ΔOMC⁵ and ΔOMC+ arabinose because curve-fits were not possible due to minimal reduction within the time frame of the experiment.](http://mic.sgmjournals.org)
wild-type rate (Fig. 1), while the original mutant ΔOMC showed almost no Fe(III) citrate reduction activity. Fe(III)-NTA, which is a smaller and more membrane-permeable iron chelate, was reduced rather slowly by ΔOMC but much faster by the suppressor mutant ΔOMC<S>, reaching 34% of the wild-type reduction rate (Fig. 2).

Birnessite, as a mineral-phase electron acceptor, was used to assess the ability of ΔOMC<S> to reduce oxidized manganese species (Fig. 3). While ΔOMC had only background reducing activity, ΔOMC<S> was able to reduce birnessite 10-fold faster compared with the original mutant and with 20% of the wild-type rate. However, the phenotype of birnessite reduction of ΔOMC<S> was not as pronounced as with Fe(III) citrate as electron acceptor.

Reduction of electron shuttles by ΔOMC<S>

Electron shuttles have been proposed to be involved in S. oneidensis-catalysed metal and mineral reduction. Either they can be produced and secreted by the cell endogenously, mainly in the form of riboflavin (Marsili et al., 2008; von Canstein et al., 2008), or they can be present in soils, typically in the form of dissolved or solid-phase humic substances (Jiang & Kappler, 2008; Roden et al., 2010). As humic substances are a structurally diverse group of substances, AQDS can be used as a surrogate for small fulvic acid molecules and for quinone moieties in humic acids (Jiang & Kappler, 2008). Reduction of large humic acid molecules was measured with PPHA.

For riboflavin and AQDS, the reduction activity was recorded over 20 min using washed cell suspensions of anoxically grown cells that should be free of any additional electron shuttle. As Fig. 4(a) indicates, reduction of riboflavin was rather slow for ΔOMC and ΔOMC<S> (12 and 21% of wild-type activity, respectively). In contrast to this, AQDS was reduced by the suppressor mutant with 81% of the wild-type rate, while the original ΔOMC strain reduced AQDS with only 32% of the wild-type rate (Fig. 4b). This effect was even more pronounced when the cells were pregrown on Fe(III) citrate, leading to a ΔOMC<S> AQDS reducing activity of 99% compared with the wild-type (Fig. 4b).

The reduction of PPHA was measured over a time frame of 18 days in a growth experiment (Fig. 5). For the reduction of PPHA, the situation resembled the results of the riboflavin reduction experiments. The maximal PPHA reduction rate reached in ΔOMC was 18% and in ΔOMC<S> 35% of wild-type activity (Fig. 5).

Reduction of Fe(III) citrate by washed cell suspensions

Cell suspension experiments were conducted to assess the ability of ΔOMC<S> cells to reduce Fe(III) citrate without the putative involvement of endogenous electron shuttles. The different S. oneidensis strains were pre-grown on minimal medium with fumarate or Fe(III) citrate as electron acceptors. Cells were then washed twice in buffer without electron acceptor under anoxic conditions and thereafter used for Fe(III) citrate reduction assays. Fumarate-grown ΔOMC<S> cells reduced Fe(III) citrate with 28% of the wild-type rate, while ΔOMC showed no detectable activity. When the cells...
were pre-grown on Fe(III) citrate, the activity of ΔOMCS increased to 51 % of the wild-type ferric iron reduction rate (Fig. 4c).

Putative involvement of other outer-membrane protein complexes

Several experimental results point towards the role of the outer-membrane c-type cytochromes MtrC and OmcA as the final metal and flavin reductases of S. oneidensis (Bücking et al., 2010; Coursolle et al., 2010). A recent study described the modularity of different outer membrane-spanning complexes in S. oneidensis (Coursolle & Gralnick, 2010). Hence, it was asked whether another surface-localized terminal reductase could functionally replace outer-membrane cytochromes. The only other enzyme fulfilling this prerequisite is the DMSO reductase DmsAB (Gralnick et al., 2006). The genome of S. oneidensis has two gene clusters encoding DMSO reductases (dmsA-1 and dmsB-1, SO_1429–30 and dmsA-2 and dmsB-2, SO_4357–58). Both loci are still present in ΔOMC, the parental strain of suppressor mutant ΔOMCS. Hence, dmsAB-1 and dmsAB-2 were deleted in ΔOMCS and the effect of these deletions on the Fe(III) citrate reduction rate of this strain was sought. The deletions did not lead to a change of the ΔOMCS phenotype with respect to Fe(III) citrate reduction (data not shown). Hence, an involvement of these proteins in the described iron reduction phenotype was excluded.

The type II secretion system transfers outer-membrane cytochromes to the outer surface of the cell in S. oneidensis. Consequently, this export machinery is essential for metal reduction (DiChristina et al., 2002; Shi et al., 2008). If other proteins could functionally replace the outer-membrane cytochromes in ΔOMCS, one would expect a possible involvement of type II secretion in protein translocation. The genes for the type II secretion systems are clustered in the gsp locus. Deletion of gspD, a key gene within the cluster, is sufficient to disable the function of the secretion system (Shi et al., 2008). As expected, gspD

---

Fig. 4. Cell suspension experiments. Reduction rates of S. oneidensis wild-type, ΔOMC mutant and ΔOMCS mutant strains with riboflavin, AQDS and Fe(III) citrate. Washed cell suspensions were prepared from cultures grown under anoxic conditions with fumarate or Fe(III) citrate as terminal electron acceptor. All experiments were carried out under anoxic conditions. Maximal reduction rates (means of triplicate measurements) were normalized to the protein content. The reduction of riboflavin (a) and AQDS (b) was measured in a continuous spectrophotometric assay using absorption maxima at 450 and 436 nm, respectively. Fe(III) citrate reduction rates (c) were measured by determining Fe(II) content discontinuously using the ferrozine reagent.

---

Fig. 5. Humic acid reduction. Electrons transferred to PPHA by different S. oneidensis strains. Values are given in electron equivalents measured with Fe(III) citrate in an electron shuttling assay. Symbols show means of duplicate measurements; error bars, SD. Solid lines represent curve-fits done with GraphPad Prism 4. The specific PPHA reduction rate, μ, in h⁻¹ was calculated for each biological replicate, and the mean ± SD values are: wild-type, 0.534 ± 0.045; ΔOMCSΔ + arabinose, 0.451 ± 0.104; ΔOMCSΔ, not determinable; ΔOMCSΔ + arabinose, not determinable.

---

http://mic.sgmjournals.org 2149
deletion led to a complete loss of ferric reductase activity using the wild-type as parental strain (Fig. 6). In contrast to this, the ΔOMC^SΔgspD (JG444) strain was not affected in Fe(III) citrate reduction, since the maximal reduction rates of ΔOMC^S and ΔOMC^SΔgspD were almost identical. Only a slightly longer lag phase of the ΔOMC^SΔgspD strain was detectable, which might be due to a slightly lower initial cell number (Fig. 6).

**Confirmation of outer-membrane integrity in ΔOMC^S**

A higher permeability of the outer membrane could cause the observed phenotype of ΔOMC^S, since it is known that periplasmic cytochromes can catalyse ferric iron reduction as well (Pitts et al., 2003; Qian et al., 2011b; Schuetz et al., 2009). Still, outer-membrane cytochromes are usually necessary, since electron acceptors such as Fe(III) citrate or birnessite cannot pass the outer membrane. Therefore, three tests were performed to exclude the possibility that the general stability or permeability of the outer membrane might be the reason for the observed phenotype of the ΔOMC^S-mutant. First ethidium bromide uptake rates were determined by measuring the increase in fluorescence over time. Ethidium bromide is a large, hydrophobic dye (about 12 Å wide) with a delocalized positive charge that makes it unlikely to diffuse through narrow porin channels (7 × 11 Å in *E. coli* OmpF; Cowan et al., 1992). It can therefore be used to measure the permeability of the outer membrane (Murata et al., 2007). The detected ethidium bromide uptake kinetics of wild-type, ΔOMC^S and ΔOMC^S were almost identical (Fig. S2). As a positive control, cells were treated with polymyxin B, which destabilizes the outer membrane (Vaara, 1992). The ethidium bromide uptake rate increased approximately twofold (Fig. S2a) but was again identical in the three tested strains. A similar experiment was performed with Fe(III) citrate as a second test to exclude the possibility that a more specific Fe(III) citrate transporter could cause the observed reduction phenotype. The amount of Fe(III) citrate diffusion into the cell was determined after 30 min incubation. The total cellular iron content after washing the cells was nearly identical for all strains but increased when polymyxin B was added (Fig. S2b).

As a third test, the antibiotic susceptibility of the different strains was compared using kanamycin A as an aminoglycoside, colistin as a polymyxine, and chloramphenicol, which forms its own group of small molecules with an aromatic ring. As Fig. S3 indicates, the inhibitory zones around the test plates containing the antibiotic drugs were either very similar for wild-type, ΔOMC^S and ΔOMC^S or smaller for ΔOMC^S compared with the other two strains. Therefore, the results of the ethidium bromide and Fe(III) citrate uptake tests were verified, and it seems unlikely that an altered outer-membrane permeability caused the observed phenotype of ΔOMC^S.

**Identification of point mutations as a genetic basis of ΔOMC suppression**

The genetic source of the ΔOMC^S mutant phenotype was addressed using Solexa sequencing of the genome and mapping to the published sequence of *S. oneidensis* MR-1 (accession no. NC_004347). The genome was nearly completely covered (>99.9 % coverage). Analysis of the assembled Solexa reads revealed 85 point mutations but no larger insertions or rearrangements of genes (Table S2). Two of the point mutations raised our interest, since they were insertions or rearrangements of genes (Table S2). As Fig. S3 indicates, the inhibitory zones around the test plates containing the antibiotic drugs were either very similar for wild-type, ΔOMC^S and ΔOMC^S or smaller for ΔOMC^S compared with the other two strains. Therefore, the results of the ethidium bromide and Fe(III) citrate uptake tests were verified, and it seems unlikely that an altered outer-membrane permeability caused the observed phenotype of ΔOMC^S.

Both proteins are crucial for dissimilatory metal reduction as well as reduction of extracellular electron shuttles. Furthermore, arabinose induction was necessary for extracellular respiration in ΔOMC^S. Therefore, it was our aim to assess whether these point mutations in mtrA and mtrB caused the observed ΔOMC^S phenotype. Hence, both genes were cloned in their wild-type and suppressor mutant form (designated mtrA and mtrB^S) in a pBAD202 expression vector.
vector, resulting in plasmids pmtrAmtrB and pmtrA\(^5\)mtrB\(^5\) (Table 2). These plasmids were transferred into the parental strain \(\Delta\)OMC and the resulting strains were tested for their ability to reduce Fe(III) citrate (Fig. 7). Expression of the wild-type allele (strain JG450) did not lead to an alteration of the \(\Delta\)OMC phenotype, but expression of the allele carrying the point mutations (strain JG449) resulted in a phenotype highly similar to \(\Delta\)OMC\(^S\) (Fig. 7). Since these data demonstrated that the point mutations in \(mtrAS\) and \(mtrBS\) were essential for suppression of the \(\Delta\)OMC\(^S\) mutation, it was concluded that the other observed point mutations might be interesting but were of minor importance for this study.

A further question arising this study was whether a single mutated gene is sufficient for ferric iron reduction without outer-membrane cytochromes. Therefore, \(mtrA\(^5\)\) and \(mtrB\(^5\)\) were cloned into separate vectors, resulting in pmtrA\(^5\) and pmtrB\(^5\) (strains JG462 and 463). Expression of both proteins could be shown by SDS-PAGE (Fig. S4). Only \(mtrB\(^5\)\) expression (strain JG463) resulted in iron reduction, although at a lower level compared with \(\Delta\)OMC\(^5\) or the \(\Delta\)OMC strain carrying pmtrA\(^5\)mtrB\(^5\) (Fig. 7). Hence, both variants, MtrA\(^5\) and MtrB\(^5\), have to be present to enable rapid reduction of Fe(III) citrate.

**Complementation of \(\Delta mtrA\) and \(\Delta mtrB\) mutants using \(mtrA\(^5\)\) and \(mtrB\(^5\)\)**

MtrA and MtrB form a complex in the outer membrane of *S. oneidensis* cells (Hartshorne et al., 2009). Furthermore, MtrA is necessary for the periplasmic stability of MtrB (Schickberger et al., 2011). Since this interaction seems to be highly specific, it was asked whether the observed point mutations led to variants of the corresponding proteins that were only able to operate in the presence of the other variant. In other words, can MtrB\(^5\) only operate in an MtrA\(^5\) strain or is an interaction with wild-type MtrA still possible? Consequently, \(mtrA\(^5\)\) and \(mtrB\(^5\)\) were expressed in *S. oneidensis* \(\Delta mtrA\) and \(\Delta mtrB\) strains, respectively. As Fig. S5 indicates, both genes can also complement the corresponding deletion mutants in a wild-type background. Hence, the point mutations do not seem to be localized to positions that disable interaction with the wild-type partner proteins.

**Table 2. Plasmids used in this study**

<table>
<thead>
<tr>
<th>Plasmid</th>
<th>Relevant genotype</th>
<th>Source or reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>pBAD202</td>
<td>Expression vector, Km(^r), arabinose-inducible promoter</td>
<td>Invitrogen</td>
</tr>
<tr>
<td>pMQ150</td>
<td>Deletion vector, cen/arsh, ura3, Km(^r), sacB</td>
<td>Shanks et al. (2006); accession no. EU546823</td>
</tr>
<tr>
<td>pmtrA(^5)</td>
<td>(mtrA) with nucleic base exchange C869A in pBAD202</td>
<td>This study</td>
</tr>
<tr>
<td>pmtrB(^5)</td>
<td>(mtrB) with nucleic base exchange C656A in pBAD202</td>
<td>This study</td>
</tr>
<tr>
<td>pmtrAmtrB</td>
<td>(mtrA) and (mtrB) in pBAD202</td>
<td>This study</td>
</tr>
<tr>
<td>pmtrA(^5)mtrB(^5)</td>
<td>(mtrA) C869A and (mtrB) C656A in pBAD202</td>
<td>This study</td>
</tr>
<tr>
<td>pmtrAmtrB(^5)</td>
<td>(mtrA) and (mtrB) C656A in pBAD202</td>
<td>This study</td>
</tr>
</tbody>
</table>

**Fig. 7. \(\Delta\)OMC complementation with pmtr plasmids. Fe(III) citrate reduction by *S. oneidensis* \(\Delta\)OMC mutant strains, complemented with different plasmids and growing on minimal medium with 50 mM lactate and 50 mM Fe(III) citrate. Arabinose was added to all experiments at a concentration of 1 mM. Means of triplicate measurements are shown; error bars, SD. Solid lines represent curve-fits done with GraphPad Prism 4. The specific iron reduction rate, \(\mu\), in h\(^{-1}\) was calculated for each biological replicate, and the mean±SD values are: \(\Delta\)OMC pmtrA\(^5\)mtrB\(^5\), 0.602±0.169; \(\Delta\)OMC pmtrAmtrB, not determinable; \(\Delta\)OMC pmtrA\(^5\), not determinable; \(\Delta\)OMC pmtrB\(^5\), 0.218±0.062; \(\Delta\)OMC pmtrAmtrB\(^5\), not determinable; \(\Delta\)OMC\(^5\), 0.451±0.104. Some reduction rates were not determinable because curve-fits were not possible due to minimal reduction within the time frame of the experiment.**
MtrAB clusters in microbial genomes

Respiration of extracellular electron acceptors without outer-membrane cytochromes may be a strategy that is used by other micro-organisms as well. Hence, it was asked whether clusters consisting of genes encoding a periplasmic decahaem cytochrome and β-barrel protein can be found in the database that are not accompanied by a gene encoding an outer-membrane decahaem cytochrome like MtrF or MtrC directly upstream or downstream. Twenty-five gene clusters with similarities to mtrAB from S. oneidensis were detected using the integrated microbial genomes tool (Table 3). Six of these clusters belong to purple bacteria. One of these purple bacteria strains, Rhodopseudomonas palustris TIE-1, has been shown to be dependent on this gene cluster when ferrous iron is added as electron donor for photolithotrophic growth (Jiao & Newman, 2007). The pioAB genes are similar to mtrAB, and they are accompanied by a gene encoding a periplasmic high-potential iron protein that has been suggested to enable periplasmic electron transfer through PioAB and, tentatively, the photosystem in the inner membrane of the organism (Bird et al., 2011). The similarities between pioABC of R. palustris TIE-1 and all other mtrAB-like clusters in Rhodopseudomonas and Rhodomicrobium strains suggest a similar function in these purple bacteria as well. Interestingly, Gallionella capsiferriformans (also known as Gallionella ferruginea, subsp. capsiferriformans) and Sideroxydans lithotrophicus also contain mtrAB-like gene cassettes. Both organisms are known to use the neutrophilic oxidation of ferrous iron with oxygen as energy source (Emerson et al., 2010). Furthermore, similar clusters were detected in other organisms, including nitrifiers, sulfate reducers and magnetotactic bacteria. Of note, a number of Geobacter strains contain similar clusters as well. Some of these clusters contain a β-barrel protein that has either low or almost no similarity to MtrB from S. oneidensis. So far it is not possible to ascribe a function to these gene clusters, but it seems possible that they enable some kind of extracellular respiration, be it electron uptake as in the case of PioABC or electron disposal.

DISCUSSION

Mesophilic dissimilatory metal reducers like S. oneidensis MR-1 have developed an extended respiratory chain to the cell surface. This respiratory chain is mainly composed of c-type cytochromes (Shi et al., 2007). In previous work we have shown that expression of the cytoplasmic membrane c-type cytochrome CymA is sufficient to transform E. coli into a dissimilatory iron reducer. Still, this respiratory process is possible only when soluble, outer-membrane-permeable iron chelates such as Fe(III)-NTA are used (Gescher et al., 2008). Environmentally relevant electron acceptors such as ferrihydrite, haematite or birnessite are poorly soluble or effectively insoluble at neutral pH (Thamdrup, 2000). Hence, their reduction depends on electron transfer through the periplasm and outer membrane. As already mentioned in the Introduction, even some chelated ferric iron sources like Fe(III) citrate are apparently not membrane-permeable (Dobbin et al., 1996; Gescher et al., 2008; Pitts et al., 2003). Large, polymeric Fe(III) citrate complexes that are observed under certain conditions might explain this effect (Spiro et al., 1967). It is therefore not surprising that the catabolic reduction of Fe(III) citrate in S. oneidensis depends on the catalytic activity of outer-membrane cytochromes. Interestingly, Qian and co-workers proposed an OMC-independent iron chelate uptake mechanism catalysed by SO_2907 and subsequent periplasmic ferric iron reduction (Qian et al., 2011a). This would certainly challenge the assumption that Fe(III) citrate can be used as a model substance to study extracellular respiration. Nevertheless, the ΔOMC strain is unable to reduce Fe(III) citrate at detectable rates (Bücking et al., 2010). Furthermore, several different laboratories have demonstrated that mutants in mtrC, mtrB or mtrA are unable to grow with Fe(III) citrate as electron acceptor (Beliaev & Saffarini, 1998; Beliaev et al., 2001; Borloo et al., 2007; Hartshorne et al., 2009; Myers & Myers, 2002b). MtrA, MtrB and MtrC build the electron conduit to the cell surface, and MtrC is probably the most influential terminal reductase for extracellular respiration in S. oneidensis (Belchik et al., 2011; Beliaev et al., 2001; Coursolle et al., 2010; Jiao et al., 2011; Marshall et al., 2006; Reardon et al., 2010). All this seems to be in disagreement with the experiments conducted by Qian and co-workers. Those authors found that a deletion mutant in a gene encoding a putative TonB-dependent receptor protein (SO_2907) was affected in ferric iron reduction. Still, they observed that the corresponding protein is an efficient Fe(III)-NTA binding protein. Hence, the observed growth phenotype might also be caused by the lack of an efficient iron acquisition protein in the outer membrane of an SO_2907 deletion mutant. Furthermore, and unfortunately, they did not show complementation of the mutant using SO_2907. Hence, it is not absolutely clear whether a polar effect of the mutation also accounts at least partly for the observed growth disadvantage.

As pointed out above, MtrC is believed to be the main terminal metal and flavin reductase. The phenotype of a ΔmtrC mutant can only be rescued if MtrC itself or the similar cytochrome MtrF is used for complementation (Bücking et al., 2010; Myers & Myers, 2001). Hence, it was a great surprise to see that a quintuple mutant in all outer-membrane cytochrome-encoding genes (ΔOMC) was able to regain its ability to grow as a dissimilatory metal reducer after a lag phase of several weeks. The new strain, ΔOMC₅, reduced Fe(III) citrate nearly as fast as the wild-type in growth experiments. Interestingly, the suppressor mutant reduced extracellular electron acceptors with varying activities when compared with the wild-type in cell suspension experiments. Fe(III) citrate and AQDS were reduced with rates ranging from >50 to 99% compared with the recorded wild-type reduction rates. In contrast, 

C. Bücking and others
riboflavin, PPHA and birnessite were reduced at only 20–35% compared with the wild-type. The different redox potentials of these compounds are unlikely to be the reason for the observed differences. Fe(III) citrate has a potential of +372 mV (Straub et al., 2001), while AQDS, with −184 mV, is more than half a volt below this value (Fultz & Durst, 1982). Birnessite has a redox potential of +612 mV (Robie et al., 1995) and is therefore an even better electron acceptor than Fe(III) citrate. The redox potential of riboflavin is, at −210 mV (Stare, 1935), very similar to the potential of AQDS. For naturally occurring humic acids, no defined redox potentials, but rather ranges similar to the potential of AQDS. Still, we cannot definitely discount the possibility that other surface-localized proteins could interact with MtrA and/or MtrB and hence could also be involved in the observed phenotype.

Unfortunately, it is so far not possible to show how the point mutations that exchange asparagine residues for lysine residues would enable this hypothesized modified structure of the MtrAB complex. The crystal structure of the two important proteins, MtrA and MtrB, is not known so far. The mutation in MtrB was predicted to localize to a hydrophilic stretch outside the β-barrel, and one could speculate that this stretch might be important for the interaction between MtrA and MtrB. The structure of MtrA was modelled recently by Firer-Sherwood et al. (2011) based on small-angle X-ray scattering. They proposed an elongated form of MtrA, which fits well with a partial insertion into the barrel of MtrB to form a complex. The point mutation in MtrA is near the C-terminal region and could be embedded into MtrB.

The ability of the suppressor mutant to reduce extracellular electron acceptors without outer-membrane cytochromes raises the question of whether this suppressor mutant resembles an evolutionary intermediate. The correct localization of outer-membrane cytochromes is a complex process. Lipid anchors have to be added to periplasmic cytochromes. These modified cytochromes have to be recognized by the type II secretion machinery. This machinery is a multiprotein complex itself, but furthermore the interaction with target proteins is highly specific and still not fully understood (Francetić & Pugsley, 2005; Sandkvist, 2001). Last but not least, these outer-membrane cytochromes have to be connected to modules such as MtrAB that thereafter can deliver electrons that originate from the cytoplasmic membrane. Due to this complexity it seems likely that the addition of outer-membrane cytochromes to existing outer-membrane conduits might be a typical example of upstream evolution. We were able to show that extracellular metal respiration is possible without outer-membrane cytochromes, although the environment might have finally selected for organisms with higher reduction rates realized by outer-membrane cytochromes. Interestingly, bioinformatic investigations revealed similarities to known ferrous iron oxidizers such as the chemolithotrophs Gallionella capsiferriformans and Sideroxydans lithotrophicus, and the photolithotrophic purple bacterium R. palustris TIE-1. For R. palustris TIE-1 and Sideroxydans lithotrophicus ES-1 it is known that genes similar to MtrAB are necessary for ferrous iron oxidation (Jiao & Newman, 2007; Liu et al., 2012). For the latter, it has been shown that MtoA from Sideroxydans lithotrophicus can functionally replace MtrA in Shewanella (Liu et al., 2012). One could envisage a succession from a module necessary for ferrous iron oxidation in one species to a module that is necessary for the reverse reaction in another species. It should be noted that there might be further metabolic processes that depend on MtrAB-like complexes, since similar gene clusters were also detected in other organisms. Future research will reveal their role in the individual organisms.

Finally, it should be mentioned that the studied suppressor mutant nicely displays the plasticity of S. oneidensis. In a previous publication a similar case was observed but not followed. Myers & Myers (2002b) could show in their experiments that a ΔmtrC mutant was capable of reducing Fe(III) citrate after a 3-day lag phase. The cells that were growing after 72 h might also have been suppressor mutants. What we can learn is that the results of mutant studies have to be treated carefully, at least in S. oneidensis.
Table 3. Gene clusters with similarities to mtrAB from *S. oneidensis*

(a), (b), (c): These genomes contain two or more MtrAB homologues.

<table>
<thead>
<tr>
<th>Genome</th>
<th>MtrA homologue</th>
<th>Expected E value</th>
<th>Number of amino acid residues</th>
<th>MtrB homologue</th>
<th>Expected E value</th>
<th>Number of amino acid residues</th>
<th>Gene upstream of MtrA homologue</th>
<th>Size of intergenic region before MtrA</th>
<th>Function of gene upstream of MtrA</th>
</tr>
</thead>
<tbody>
<tr>
<td>Desulfobacterium autotrophicum HRM2</td>
<td>HRM2_13960</td>
<td>4E-44</td>
<td>323</td>
<td>HRM2_13950</td>
<td>−</td>
<td>583</td>
<td>HRM2_13970</td>
<td>466 bp</td>
<td>Dicarboxylate transporter protein†</td>
</tr>
<tr>
<td>Desulfitomonas acetoxidans DSM 684</td>
<td>Dace_1556</td>
<td>9E-43</td>
<td>354</td>
<td>Dace_1557</td>
<td>−</td>
<td>805</td>
<td>Not sequenced</td>
<td>Not sequenced</td>
<td>Cytochrome c class I (very small, 112 aa)‡</td>
</tr>
<tr>
<td>Geobacter benedictensis Bem, DSM 16622</td>
<td>Gbem_3118</td>
<td>4E-38</td>
<td>334</td>
<td>Gbem_3117</td>
<td>−</td>
<td>718</td>
<td>Gbem_3119</td>
<td>1 bp</td>
<td>ABC transporter-related protein</td>
</tr>
<tr>
<td>Geobacter sp. FRC-32</td>
<td>Geob_2912</td>
<td>3E-39</td>
<td>331</td>
<td>Geob_2911</td>
<td>−</td>
<td>701</td>
<td>Geob_2913</td>
<td>1 bp</td>
<td>ABC transporter-related protein</td>
</tr>
<tr>
<td>Geobacter sp. M18 (a)</td>
<td>GM18_0932</td>
<td>2E-08</td>
<td>192</td>
<td>GM18_0933</td>
<td>−</td>
<td>660</td>
<td>GM18_0931</td>
<td>60 bp</td>
<td>NHL repeat-containing protein</td>
</tr>
<tr>
<td>Geobacter sp. M18 (b)</td>
<td>GM18_1019</td>
<td>3E-07</td>
<td>192</td>
<td>GM18_1020</td>
<td>−</td>
<td>653</td>
<td>GM18_1018</td>
<td>18 bp</td>
<td>NHL repeat-containing protein</td>
</tr>
<tr>
<td>Geobacter sp. M18 (c)</td>
<td>GM18_3089</td>
<td>1E-42</td>
<td>340</td>
<td>GM18_3088</td>
<td>−</td>
<td>762</td>
<td>GM18_3090</td>
<td>1 bp</td>
<td>ABC transporter-like protein</td>
</tr>
<tr>
<td>Geobacter sp. M21 (a)</td>
<td>GM21_0397</td>
<td>7E-66</td>
<td>299</td>
<td>GM21_0398</td>
<td>3E-16</td>
<td>807</td>
<td>GM21_0396</td>
<td>367 bp</td>
<td>Transcriptional regulator</td>
</tr>
<tr>
<td>Geobacter sp. M21 (b)</td>
<td>GM21_1144</td>
<td>3E-44</td>
<td>334</td>
<td>GM21_1145</td>
<td>−</td>
<td>716</td>
<td>GM21_1143</td>
<td>1 bp</td>
<td>ABC transporter-related protein</td>
</tr>
<tr>
<td>Geobacter uranireducens RH4 (a)</td>
<td>Gura_2291</td>
<td>7E-46</td>
<td>335</td>
<td>Gura_2290</td>
<td>−</td>
<td>712</td>
<td>Gura_2292</td>
<td>1 bp</td>
<td>ABC transporter-related protein</td>
</tr>
<tr>
<td>Geobacter uranireducens RH4 (b)</td>
<td>Gura_3626</td>
<td>1E-61</td>
<td>299</td>
<td>Gura_3627</td>
<td>1E-17</td>
<td>835</td>
<td>Gura_3625</td>
<td>486 bp</td>
<td>Transcriptional regulator</td>
</tr>
<tr>
<td>Magnetospirillum magnetinum AMB-1</td>
<td>amb3017</td>
<td>6E-63</td>
<td>318</td>
<td>amb3018</td>
<td>2E-21</td>
<td>731</td>
<td>amb3016</td>
<td>3 bp</td>
<td>Hypothetical</td>
</tr>
<tr>
<td>Nitrosococcus halophilus Nc4 (a)</td>
<td>Nhal_1192</td>
<td>7E-59</td>
<td>325</td>
<td>Nhal_1191</td>
<td>2E-21</td>
<td>827</td>
<td>Nhal_1193‡</td>
<td>38 bp</td>
<td>NUDIX hydrolase</td>
</tr>
<tr>
<td>Nitrosococcus halophilus Nc4 (b)</td>
<td>Nhal_1655</td>
<td>1E-76</td>
<td>312</td>
<td>Nhal_1654</td>
<td>1E-75</td>
<td>755</td>
<td>Nhal_1656</td>
<td>75 bp</td>
<td>Peptidoglycan-binding LysM</td>
</tr>
<tr>
<td>Nitrosococcus oceani AFC27</td>
<td>NOC27_2654</td>
<td>7E-45</td>
<td>168</td>
<td>NOC27_3054</td>
<td>1E-07</td>
<td>461</td>
<td>NOC27_2862</td>
<td>51 bp</td>
<td>Hypothetical</td>
</tr>
<tr>
<td>Nitrosococcus oceani ATCC 19707</td>
<td>Noc_2748</td>
<td>−</td>
<td>315</td>
<td>Noc_2747</td>
<td>2E-27</td>
<td>827</td>
<td>Noc_2749</td>
<td>44 bp</td>
<td>NUDIX hydrolase†</td>
</tr>
<tr>
<td>Rhodomicrobium vanniellii ATCC 17100</td>
<td>Rvan_3213</td>
<td>2E-48</td>
<td>476</td>
<td>Rvan_3214</td>
<td>2E-06</td>
<td>844</td>
<td>Rvan_3212</td>
<td>468 bp</td>
<td>Exonuclease RecJ</td>
</tr>
<tr>
<td>R. palustris BixA53</td>
<td>RPE_0831</td>
<td>9E-49</td>
<td>632</td>
<td>RPE_0832</td>
<td>2E-13</td>
<td>793</td>
<td>RPE_0830</td>
<td>754 bp</td>
<td>Alpha/beta hydrolase</td>
</tr>
<tr>
<td>R. palustris BixB18</td>
<td>RPC_2960</td>
<td>1E-45</td>
<td>577</td>
<td>RPC_2959</td>
<td>1E-14</td>
<td>830</td>
<td>RPC_2961†</td>
<td>545 bp</td>
<td>Putative flavoprotein involved in K⁺ transport</td>
</tr>
<tr>
<td>R. palustris CGA009</td>
<td>RPA0746</td>
<td>5E-48</td>
<td>534</td>
<td>RPA0745</td>
<td>1E-08</td>
<td>810</td>
<td>cysA</td>
<td>775 bp</td>
<td>ABC transporter</td>
</tr>
</tbody>
</table>
Table 3. cont.

<table>
<thead>
<tr>
<th>Genome</th>
<th>MtrA homologue</th>
<th>MtrB homologue</th>
<th>Size of intergenic region before MtrA</th>
<th>Gene upstream of MtrA</th>
<th>Gene upstream of MtrB</th>
<th>Function of gene upstream of MtrA</th>
<th>Function of gene upstream of MtrB</th>
</tr>
</thead>
<tbody>
<tr>
<td>R. palustris DX-1</td>
<td>Rpdx1_0796</td>
<td>Rpdx1_0797</td>
<td>548 bp</td>
<td>Sulfitatericase, ATPase subunit</td>
<td>Rpdx1_0796</td>
<td>7E-48</td>
<td>ATPase subunit</td>
</tr>
<tr>
<td>R. palustris TIE-1</td>
<td>Rpal_0817</td>
<td>Rpal_0818</td>
<td>810 bp</td>
<td>2E-11</td>
<td>Rpal_0817</td>
<td>2E-08</td>
<td>9E-11</td>
</tr>
<tr>
<td>Sideroxydans</td>
<td>Slit_2497</td>
<td>Slit_2498</td>
<td>45 bp</td>
<td>Cytochrome c, class I</td>
<td>Slit_2497</td>
<td>4E-19</td>
<td>Decaheme c type cytochrome (671 aa)*</td>
</tr>
<tr>
<td>HMPREF9464_01980</td>
<td>HMPREF9464_01979</td>
<td>HMPREF9464_01979</td>
<td>1E-11</td>
<td>Decaheme type cytochrome (671 aa)*</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*These genes are predicted to encode β-barrel proteins (http://www.cbs.dtu.dk/services/LipoP/), but the E value of a BLAST search against MtrB is above 1E-5.

**These proteins are not predicted to be lipoproteins by LipoP (http://www.cbs.dtu.dk/services/LipoP/).

---

The shorter the duration of an experiment, the more one can exclude the possibility that the observed phenotype might not be due to the inserted genetic modification but to a suppressor mutation. Therefore, Fe(III) citrate seems to be a good model substance to study extracellular electron transport pathways, since it allows fast growth and reduction rates when compared with mineral phase electron acceptors such as ferrihydrite or haematite.

In conclusion, this study characterizes for the first time MtrA and MtrB as a final reductase necessary for extracellular respiration. Only two point mutations were necessary to gain this activity. It seems possible that this form of dissimilatory metal reduction resembles a preliminary form which might have been the intermediate state between lithotrophic ferrous iron oxidation and hetero-trophic ferric iron respiration.

REFERENCES


