INTRODUCTION

Campylobacter jejuni is the most important cause of bacterial diarrhoea of humans in developing and developed countries (Altekruse et al., 1999; Friedman et al., 2000; Mead et al., 1999). The spectrum of the disease can vary from mild watery diarrhoea to bloody diarrhoea and is characterized by fever and abdominal cramps. In addition, the Guillain–Barré syndrome might emerge as a post-infection complication (Allos, 2001). At the beginning of the infection, C. jejuni colonizes the small intestine before moving to the colon, which represents the target organ. C. jejuni is generally considered to be invasive; the level of invasiveness varies depending on the strain (Black et al., 1988; van Vliet & Ketley, 2001). In contrast to the infection of humans, C. jejuni is only a commensal organism in chickens and other birds, where the pathogen preferentially resides in the caecum, which contains many anaerobic fermentative bacteria (Barnes et al., 1972; Lu et al., 2003). Within those natural habitats, C. jejuni is able to metabolize products like free amino and keto acids generated by these accompanying bacterial species or from the host itself (Lee & Newell, 2006). Since C. jejuni lacks the glycolytic enzyme phosphofructokinase, glucose cannot be catabolized, and other products serve as electron donors for respiration, for instance succinate, malate, formate, D-lactate, hydrogen and NAD(P)H (Hoffman & Goodman, 1982; Parkhill et al., 2000; Kelly, 2001; Sellars et al., 2002; Velayudhan & Kelly, 2002). It has also been shown that sulphite can be exploited as a respiratory electron donor. Like chemolithotrophic bacteria, C. jejuni possesses a sulphite:cytochrome c oxidoreductase (SOR), which is encoded by the genes cj0005c and cj0004c. The cj0004c gene encodes a monohaem cytochrome c oxidoreductase, while cj0005c encodes a molybdopterin oxidoreductase (Myers & Kelly, 2005). The possession of a sulphite oxidation system may be important for the pathogen to survive in foodstuffs that have been treated with sulphite for preservative reasons, or in the human host, since it has been shown that sulphite is also released by neutrophils as part of the host defence (Mitsuhashi et al., 1998). Thus a potential antimicrobial chemical can be used by C. jejuni for the generation of electrons, which are subsequently supplied to the respiratory chain of the pathogen and therefore serve for the provision of energy.

In this report we describe that the possession of SOR and the ability to utilize sulphite as an electron donor significantly influences bacterial motility, gene regulation,
autoagglutination and the invasiveness of \( C. jejuni \) towards host cells. Bacterial proliferation and intracellular ATP level do not correlate with the presence of this enzyme.

**METHODS**

**Bacterial strains, media and culture conditions.** \( C. jejuni \) strain B2, isolated from a patient suffering from gastroenteritis, was grown on Columbia agar supplemented with 5% defibrinated sheep blood at 42 °C in microaerobic conditions (85% N\(_2\), 10% CO\(_2\), 5% O\(_2\)) (Schmidt-Ott et al., 2005; Dasti et al., 2007). When needed, kanamycin and chloramphenicol were added at concentrations of 50 \( \mu \)g ml\(^{-1}\) and 30 \( \mu \)g ml\(^{-1}\), respectively. Growth experiments were carried out at 42 °C in Mueller–Hinton (MH) broth under microaerobic conditions in the presence or absence of 10 mM Na\(_2\)SO\(_3\). Cloning experiments were performed in *Escherichia coli* strain DH5\(_{\alpha}\). The *E. coli* bacteria were grown on Luria Bertani (LB) agar or in LB broth at 37 °C. When necessary, ampicillin (100 \( \mu \)g ml\(^{-1}\)) was added.

**Preparation of competent cells and electroporation.** *E. coli* DH5\(_{\alpha}\) was grown in 500 ml LB broth to an OD\(_{600}\) of 0.35–0.45. The culture was stored on ice for 10 min and centrifuged at 4000 g at 4 °C. After washing the bacteria three times in ice-cold water, the pellet was resuspended in 15 ml 10% (v/v) glycerol and centrifuged again. Finally, the pellet was dissolved in 1 ml 10% (v/v) glycerol and 100 \( \mu \)l aliquots were used for electroporation. \( C. jejuni \) cells were collected from Columbia agar plates. Then the cells were washed four times in ice-cold washing buffer containing 272 mM sucrose and 15% (v/v) glycerol. Finally, the bacteria were resuspended in 400 \( \mu \)l washing buffer and aliquots of 100 \( \mu \)l were used for electroporation.

For each electroporation of *E. coli* and \( C. jejuni \), 0.5–3 \( \mu \)g plasmid DNA was added to the competent cells and the mixture was transferred into an ice-cold electroporation cuvette. After incubation on ice for 30 min, electroporation was carried out at 2.5 kV, 25 \( \mu \)F and 200 \( \Omega \) with a BTX Electro Cell manipulator before 500 \( \mu \)l SOC medium (2% [w/v] tryptone, 0.5% [w/v] yeast extract, 10 mM NaCl, 2.5 mM KCl, 20 mM MgSO\(_4\), 20mM glucose) was added. In the case of \( C. jejuni \) electroporation, the cells were incubated on non-selective Colombia blood agar plates overnight at 37 °C under microaerobic conditions. Then, the cells were transferred onto plates supplemented with appropriate antibiotics and incubated under microaerobic conditions at 42 °C. Transformed *E. coli* bacteria were directly plated on selective LB agar and incubated at 37 °C overnight.

**Cultivation of Caco2 cells.** Human colon carcinoma Caco2 cells were cultivated in Dulbecco’s minimal essential medium (DMEM) supplemented with 10% fetal bovine serum, 1x non-essential amino acids, 100 U penicillin ml\(^{-1}\) and 100 \( \mu \)g streptomycin ml\(^{-1}\) in a humidified atmosphere of 95% air and 5% CO\(_2\) at 37 °C.

**Invasion and adhesion assays.** Bacterial invasion assays were carried out according to Everest et al. (1992). Caco2 cells were grown to approximately 80% confluence in a six-well plate, then the cells were washed with PBS and inoculated with 400 \( \mu \)l *C. jejuni* suspension adjusted to an OD\(_{600}\) of 0.5, which represents an m.o.i. of 100. In the experiments in which the bacteria were brought into contact with the Caco2 cells, the six-well plates were centrifuged at 600 g for 15 min. Two hours post-infection, the bacterial suspension was removed and the cells were washed three times with PBS before further incubation with culture medium supplemented with 100 \( \mu \)g gentamicin ml\(^{-1}\). Then the Caco2 cells were lysed by treatment with 1% Triton X-100 for 10 min and the number of viable bacteria was determined by counting c.f.u. on Columbia blood agar plates after incubation for 48 h at 42 °C under microaerobic conditions.

For the investigation of bacterial adhesion, Caco2 cells were incubated with the bacteria for only 30 min. Then the monolayer was washed with PBS, the cells were lysed and the bacteria were plated on Columbia blood agar plates, where the number of recovered bacteria was determined as described above. Every experiment was repeated six times.

**ATP assays.** To determine the intracellular ATP concentration of bacterial samples, the BacTiter-Glo assay from Promega was used. After growing the bacteria in MH broth at 42 °C under microaerobic conditions, cultures were adjusted to an OD\(_{600}\) of 0.5. Then, serial dilutions of all samples were prepared according to the manufacturer’s instructions. Following incubation of the samples at room temperature in a 96-well plate, luminescence was measured together with an ATP standard using a VICTOR \(^{\text{X}}\)V Multilabel Counter (Perkin Elmer).

**Motility assays.** One microlitre of an overnight culture that was adjusted to an OD\(_{600}\) of 0.05 was stabbed onto the centre of a 0.4% MH agar plate. After incubation of the plate at 42 °C for 36 h, motility was judged by measuring the diameter of the halo. Every experiment was carried out four times.

**Autoagglutination assays.** Autoagglutination assays were carried out as initially described by Misawa & Blaser (2000). Bacteria grown on plates were suspended in PBS and adjusted to an OD\(_{600}\) of approximately 1.0. A 2 ml sample of each suspension was incubated under microaerobic conditions for 24 h at 37 °C. Then 1 ml of the bacterial suspension was carefully removed and the OD\(_{600}\) was determined. The parental strain B2, its knockout mutant and the complemented mutant were tested five times each. The data obtained were normalized according to Howard et al. (2009); the OD\(_{600}\) measured after 24 h was subtracted from the OD\(_{600}\) from the start of the experiment; the values obtained were divided by the initial OD\(_{600}\) and were finally multiplied by 100 in order to give the percentage autoagglutination.

**Isolation of nucleic acids.** Genomic DNA of *C. jejuni* was isolated with the QIAamp DNA Mini kit (Qiagen), following the manufacturer’s instructions. Plasmid DNA was prepared using the GeneElute Plasmid Miniprep kit (Sigma) according to the manufacturer’s protocol. Bacterial RNA from *C. jejuni* was extracted with the RiboPure Bacter kit (Ambion). DNA contaminations were removed by the treatment with DNase I (Ambion) as recommended by the protocol. Prior to RT-PCR analysis, PCR assays with the isolated bacterial RNA as a template were performed to exclude remaining traces of DNA.

**Sequencing of genomic DNA.** Genomic DNA sequencing was performed by SeqLab (Göttingen, Germany) using 10 \( \mu \)g genomic *C. jejuni* DNA and 10 pmol KanF primer (for all primer sequences, see Supplementary Table S1, available with the online version of this paper).

**Insertional knockout of cj0005c.** Primers Cj0005cF and Cj0005cR (Supplementary Table S1) were used to amplify a 1239 bp DNA fragment corresponding to cj0005c. The PCR fragment was XbaI digested and inserted into the XbaI-restricted and dephosphorylated plasmid vector pBluescript II KS (Stratagene) to obtain pBcj0005c. For the insertion of a kanamycin-resistance cassette, the aphA-3 gene including a 5′-promoter was amplified from plasmid pSB1699 (Coley et al., 2001) using the 5′-phosphorylated primers Kan1 and Kan2 (Supplementary Table S1). A PCR containing 10 mM Tris/ HCl pH 8.3, 50 mM KCl, 1.5 mM MgCl\(_2\), all four dNTPs (each 0.2 mM), 10 pmol primer and 1 U Pfu DNA polymerase was carried out to obtain a blunt-ended PCR product. Initial incubation at 95 °C for 1 min was followed by 30 cycles at 93 °C for 30 s, 55 °C for 30 s
and 72 °C for 1 min, with a final incubation at 72 °C for 10 min. After purification of the PCR product using the QIAquick PCR Purification kit (Qiagen), the kanamycin-resistance cassette was ligated into plasmid pBcj0005c, which was BsaBI-restricted and dephosphorylated with Antarctic Phosphatase (New England Biolabs) to obtain pBcj0005cKan.

Cloning of cj0005c into pRRC. The C. jejuni gene cj0005c was amplified by PCR and cloned into C. jejuni expression vector pRRC (Karlyshev & Wren, 2005). The primers Cj0005cF and Cj0005cR with Xbal restriction sites are listed in Supplementary Table S1. PCR was carried out using a TRIO-Thermocycler (Biometa) with 10 ng genomic DNA of C. jejuni strain B2 as a template. The PCR mixture contained 0.02 U KOD Hot Start DNA Polymerase (Novagene), dNTPs (each 0.2 mM), 1.5 mM MgSO4, 10 pmol of each primer and 1 x KOD Hot Start buffer. After initial incubation at 95 °C for 3 min, 40 cycles at 95 °C for 30 s, 55 °C for 30 s and 72 °C for 1 min were carried out with a final incubation at 72 °C for 5 min. After PCR, the PCR amplon and the expression vector pRRC were Xbal digested. Following dephosphorylation of pRRC with Antarctic Phosphatase (New England Biolabs), the restricted PCR fragment was ligated into pRRC using the Quick Ligase kit (New England Biolabs), following the manufacturer’s recommendations.

Real-time RT-PCR analysis. Semiquantitative real-time RT-PCR analysis was performed with the LightCycler 1.5 instrument (Roche) and the Quantifast SYBR Green RT-PCR kit (Qiagen) as described previously (Tareen et al., 2010). In brief, 10 ng samples of RNA isolated from C. jejuni B2 wild-type strain, its knockout mutant B2A:: cj0005c and its complemented knockout mutant were used to compare the transcription levels of the genes summarized in Supplementary Table S1. Amplicons of the 23S gene were used to normalize the RNA pools under investigation based on the crossing points obtained. 23S RT-PCR was carried out as follows. Initial reverse transcription for 20 min at 50 °C and denaturation for 5 min at 95 °C was followed by 30 cycles of denaturation (95 °C; 10 s), annealing (50 °C; 10 s) and elongation (72 °C; 5 s). After adjustment of the RNA samples with regard to the crossing points of the 23S gene, real-time RT-PCR assays were undertaken in order to compare the transcription level of all other investigated genes. Initially, RNA was reverse transcribed for 20 min at 50 °C and denatured for 5 min at 95 °C. Subsequently, 28–35 cycles of denaturation (95 °C; 10 s), annealing (58 °C; 10 s) and elongation (72 °C; 10 s) were executed. The primers used are listed in Supplementary Table S1. Every assay was run in triplicate. Semiquantitative real-time RT-PCR analysis follows:

\[ x = 2^{\Delta \text{Cp}} , \]  
\[ \Delta \text{Cp} \text{ stands for the difference in crossing points (Cp1} - \text{Cp2) of the sample pools compared.} \]

Statistical analysis. The STATISTICA software (StatSoft) was used to perform Student’s t-tests. P-values <0.01 were considered to be significant.

RESULTS

Identification of cj0005c

In order to detect genes of C. jejuni that are important for the pathogen to invade host cells, we screened a transposon-based mutant library. A total of 660 mutants were examined for their capacity to infect Caco2 cells (Tareen et al., 2010). One of the mutants that showed a clearly reduced infectivity carried a transposon insertion in the gene cj0005c, which was mapped by direct sequencing of genomic DNA using a primer that binds directly to the 5’ region of the kanamycin-resistance cassette (Supplementary Table S1). Gene cj0005c encodes a molybdopterin oxidoreductase, which, together with the adjacent gene cj0004c encoding the monohaem cytochrome c oxidoreductase, constitutes a sulphite : cytochrome c oxidoreductase (SOR). In three independent gentamicin protection assays, the percentage of recovered colonies of mutants compared to the parental strain B2 was 24.5, 26.5 and 25 %, respectively, with a mean value of 25.34 % (P<0.001).

Construction of a knockout mutant and complementation of the mutant phenotype

Since it could not be ruled out that the observed invasion-deficient phenotype of C. jejuni B2A:: cj0005c was not due to the transposon insertion into cj0005c but rather to genetic rearrangements in the genome, we created a knockout mutant to confirm the above-described observations. After transformation of C. jejuni B2 with pBcj0005cKan, PCR analysis was carried out by using genomic DNA from the obtained C. jejuni clones for the detection of a single band corresponding to the length of cj0005c including the aphA-3 cassette (B2:: cj0005c) (Fig. 1b). Subsequent gentamicin protection assays with the generated knockout mutant B2:: cj0005c confirmed the observed phenotype of C. jejuni strain B2 harbouring the transposon within cj0005c, clearly demonstrating that the inactivation of cj0005c was responsible for the invasion-deficient phenotype (Fig. 1c). To restore the phenotype of the parental strain B2, we introduced the C. jejuni expression plasmid pRRC carrying cj0005c into B2:: cj0005c to obtain B2:: cj0005c-comp-cj0005c. Because in pRRC the chloramphenicol-resistance cassette and downstream gene cj0005c are flanked by 16S and tRNA sequences, a double recombination event leads to the integration of both genes into one of the three 16S gene clusters. After verification of the intact cj0005c gene by PCR (Fig. 1b), we repeated invasion assays with wild-type strain B2, both mutants and the complemented knockout mutant. Taking the number of recovered B2 colonies as 100 %, the mean values of invasion capacity of B2A:: cj0005c, B2:: cj0005c and B2:: cj0005c-comp-cj0005c were 25 %, 29.8 % and 85.6 %. The altered invasion capacities of both mutants were clearly significant (P<0.0001). In contrast, the P-value for the complemented knockout mutant was only 0.16, indicating no significant difference compared to the parental strain B2 (Fig. 1c). An overview of the genetic arrangement of cj0005c in the parental strain, the parental strain with the transposon insertion in cj0005c, the cj0005c knockout mutant and the complemented mutant is shown in Fig. 1(a).
strains in the presence and the absence of sodium sulphite. While in the absence of sodium sulphite no differences in the proliferation of the parental strain B2, the knockout mutant B2::cj0005c and the complemented mutant were observed, the growth of the mutant was clearly reduced as compared to B2 and B2::cj0005c-comp-cj0005c when sodium sulphite was added to the medium at a concentration of 10 mM (Fig. 2).

**ATP levels do not differ in the wild-type, mutant and complemented mutant**

Since we could detect no variations in bacterial proliferation of the populations examined in a medium that did not contain the particular substrate for SOR, we asked whether also the ATP level is identical in the parental strain B2 and the cj0005c-deficient mutant. Upon measuring serial dilutions of the bacteria under investigation with regard to their intracellular ATP content, we could not detect any differences (Fig. 3), demonstrating that the ATP level is not deficient phenotype of B2Δcj0005c could be confirmed after knockout of cj0005c (B2::cj0005c). Complementation of the knockout mutant restored the parental phenotype. The data are means ± SD.
altered even in the absence of an important enzyme, which is an integral part of the energy metabolism.

**Reduced motility of C. jejuni in the absence of a functional SOR**

We further asked if the motility of the pathogen is diminished by *cj0005c* mutation, since motility is an energy-consuming process. Upon comparing the parental strain B2, its knockout mutant and the complemented knockout mutant, we clearly detected a motility loss of the mutant B2::*cj0005c* that could be reverted completely to the level of the wild-type strain after introduction of a functional *cj0005c* gene into the mutant. In four independent motility assays, the mean diameter (±SD) of the B2 motility zone was 40 ± 4.6 mm, while the mutant B2::*cj0005c* was significantly less motile (18.2 ± 1.7 mm, *P* < 0.01). Introduction of a functional copy of *cj0005c* restored the motility to wild-type level again (38.7 ± 4.1 mm). These findings implied that the reduced infectivity may be primarily due to a lack of motility in the absence of a functional SOR.

**Downregulation of flhA, flIR and rpoN transcription in the *cj0005c* mutant**

To gain an insight into how the lack of a functional SOR is linked to reduced motility, we investigated the regulation of genes which have been shown to be important for the composition of a functional flagellar apparatus (Hendrixson & DiRita, 2003). We carried out semiquantitative real-time RT-PCR assays to compare the transcription of genes *rpoN*, *flgR*, *flgS*, *flhA*, *flhB*, *fliP*, *flIR* and *maf7* in parental strain B2 and the mutant. While the transcription levels of *flgS*, *flgR*, *maf7*, *flhB* and *fliP* were not altered relevantly, we detected a significant downregulation of *rpoN*, *flIR* and *flhA* in the mutant strain compared to strain B2; the downregulation of *flhA* was particularly marked (Table 1). Subsequent testing of the transcription levels of genes significantly downregulated in the mutant revealed only minor differences of the respective crossing points in the parental strain B2 compared to the complemented mutant. The results are shown in Supplementary Table S2.

**Reduced motility contributes to decreased invasiveness**

To investigate whether the reduced motility is the only reason for the clearly reduced invasion capacity of the *cj0005c*-deficient mutant, we performed modified gentamicin protection assays in which motility was mimicked by centrifugation of the bacteria onto the Caco2 host cells. Taking the number of colonies recovered in the case of wild-type strain B2 as 100%, we detected only 48.6% of colonies (*P* < 0.01) after centrifugation of the knockout mutant B2::*cj0005c*. The complemented mutant gave 87% of recovered colonies, which was not significantly different compared to the parental strain (Fig. 4). As compared to the gentamicin protection assays without centrifugation (Fig. 1c) we could clearly observe an increase in recovered mutant colonies from 29.8% up to 48.6%. Nevertheless, this finding was surprising since one would have expected to shift the mutant invasiveness up to wild-type level after bypassing motility as influencing factor. In contrast, this result indicates that altered motility is one, but not the only factor responsible for the clearly reduced invasiveness of the *cj0005c*-deficient mutant.

**Reduced invasiveness is also caused by diminished adherence**

To characterize the process of invasion in more detail, we investigated whether the adherence capability of the *cj0005c*-deficient mutant was altered. For this, we carried out adhesion assays of the wild-type isolate B2, the mutant B2::*cj0005c* and the complemented mutant. After centrifugation of the bacteria onto the host cells, we could clearly determine an adhesion-deficient phenotype of the mutant, which could be restored to wild-type behaviour after complementation. Taking the number of recovered colonies from the parental strain B2 as 100%, the mean value of colonies from the mutant B2::*cj0005c* was 67% (±5.1, *P* < 0.001) in contrast to B2::*cj0005c*-comp-*cj0005c* with a recovery rate of 94% (±2.2) (Fig. 4).

**The absence of a functional SOR downregulates the transcription of virulence-associated factors**

After the confirmation that the reduced motility in combination with diminished adherence is a reason for the decreased invasiveness of the *cj0005c* mutant, we examined the transcription level of genes involved in virulence and, in particular, in the adherence of *C. jejuni* to host cells. The investigated genes were *groEL* and *dnaJ*,

![Fig. 3. Intracellular ATP levels of parental strain B2, mutant B2::*cj0005c* and complemented mutant. ATP contents of four serial dilutions of the bacteria under investigation were measured. The results are shown as means ± SD of data from triplicate wells of a representative experiment. No differences in intracellular ATP content between the bacterial strains could be detected (*P* > 0.1).](image-url)
encoding heat-shock proteins (Thies et al., 1999; Konkel et al., 1998), the chemotaxis regulator gene cheY (Yao et al., 1997), the putative chemoreceptor tlp7 (Marchant et al., 2002), genes that encode proteins important for binding and adhesion of C. jejuni, like cadF, peb1, jlpA, porA and fliA (Konkel et al., 1997; Pei & Blaser 1993; Jin et al., 2001; Flanagan et al., 2009; Konkel et al., 2010) and genes known to be responsible for O-linked glycosylation of the flagella by the synthesis of pseudaminic acid and legionaminic acid (LegAm) (Legan et al., 2008). In addition, we compared the transcription level of Campylobacter invasion antigen gene ciaB (Rivera-Amill et al., 2001; Konkel et al., 2004), the cytolethal distending toxin subunit B gene cdiB (Pickett & Whitehouse, 1999) and flaA (Guerry et al., 1991) in the wild-type strain B2 and its mutant. Semiquantitative real-time RT-PCR assays detected a statistically significant 5.16-fold downregulation of dnaJ transcription in the mutant compared to the transcription level of this gene in wild-type strain B2 (P<0.001). In addition, the transcription of ptmB, ptmC and ptmE, which contribute to a set of genes responsible for the synthesis of LegAm were significantly downregulated by a factor of 4.31–7.67 (P<0.001) as outlined in Table 2. The transcription of other genes, including genes of LegAm metabolism, were also downregulated significantly, but, to a lesser extent (ptmA, ptmF, ptmG). Other virulence factors investigated, e.g. for the biosynthesis of pseudaminic acid, were not controlled or only regulated by a factor of uncertain biological significance. The results are summarized in Supplementary Table S3.

Further, we tested in the cj0005c-complemented mutant the transcription of the genes judged to be significantly downregulated in the cj0005c-mutant. The results showed crossing points on the same transcription level as for the parental strain (Supplementary Table S2).

### Autoagglutination deficiency of mutant B2::cj0005c

Howard et al. (2009) demonstrated that C. jejuni mutants lacking gene cj1324 or genes cj1321–1325/6, known to be responsible for the synthesis of LegAm, had significantly lower levels of autoagglutination compared to the parental strain. By comparing the transcription profile of the mutant B2::cj0005c with that of the wild-type strain B2 we were able to detect a significant downregulation of transcription of ptmG (cj1324) as well as that of other genes involved in the synthesis of LegAm as described above. We therefore tested whether the competence of mutant B2::cj0005c is also affected in regard to autoagglutination. We found that the autoagglutination capability of the mutant is indeed decreased. Whereas the percentage autoagglutination of wild-type strain B2 and the complemented mutant was 74.24 % (±0.25) and 73.65 % (±0.22), respectively, the mutant B2::cj0005c had an

### Table 1. Transcription levels of genes important for flagellar synthesis in parental strain B2 and mutant strain B2::cj0005c

The crossing point data are means ± SD. Significantly downregulated genes are in bold.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Crossing point</th>
<th>Fold change (t-test P-value)</th>
<th>Function</th>
</tr>
</thead>
<tbody>
<tr>
<td>B2::cj0005c</td>
<td>B2 wild-type</td>
<td></td>
<td></td>
</tr>
<tr>
<td>flhA</td>
<td>20.86 (±0.06)</td>
<td>17.98 (±0.32)</td>
<td>7.41 (0.0001) Flagellar biosynthesis</td>
</tr>
<tr>
<td>flhB</td>
<td>19.08 (±0.10)</td>
<td>17.64 (±0.14)</td>
<td>2.71 (0.065) Flagellar biosynthesis</td>
</tr>
<tr>
<td>flgR</td>
<td>16.16 (±0.10)</td>
<td>15.14 (±0.36)</td>
<td>2.02 (0.0051) Signal transduction regulation</td>
</tr>
<tr>
<td>flgS</td>
<td>20.52 (±0.18)</td>
<td>19.92 (±0.19)</td>
<td>1.51 (0.062) Signal transduction histidine kinase</td>
</tr>
<tr>
<td>fliP</td>
<td>20.80 (±0.06)</td>
<td>19.44 (±0.05)</td>
<td>2.56 (0.0051) Flagellar biosynthesis</td>
</tr>
<tr>
<td>fliR</td>
<td>20.24 (±0.10)</td>
<td>18.66 (±0.13)</td>
<td>3.24 (0.0001) Flagellar biosynthesis</td>
</tr>
<tr>
<td>maf7</td>
<td>24.46 (±0.01)</td>
<td>22.72 (±0.10)</td>
<td>3.34 (0.035) Unknown</td>
</tr>
<tr>
<td>rpoN</td>
<td>24.07 (±0.24)</td>
<td>21.98 (±0.15)</td>
<td>4.25 (0.0001) RNA polymerase σ54 subunit</td>
</tr>
</tbody>
</table>

**Fig. 4.** Invasion and adhesion assays with wild-type strain B2, knockout mutant B2::cj0005c and complemented mutant B2::cj0005c-comp-cj0005c. Dark grey bars: gentamicin protection invasion assays after centrifugation of the bacteria onto Caco2 cells. Taking the number of C. jejuni strain B2 colonies recovered as 100 %, the mean percentage of knockout mutant B::cj0005c colonies was 48.6 %. Complementation of the mutant partially restored the parental phenotype (87 % recovery). See text for details. Light grey bars: adhesion assays after centrifuging of the bacteria onto Caco2 cells. The percentage of adhered colonies of the mutant B2::cj0005c was only 67 % compared to wild-type isolate B2.
autoagglutination level of only 37.02% (± 0.61), which was significantly reduced compared to B2 (P<0.001).

**DISCUSSION**

*C. jejuni* possesses a highly branched and complex respiratory chain that allows the pathogen to use a variety of electron donors such as D-lactate, hydrogen, formate, succinate, malate, NAD(P)H and sulphite (Kelly, 2001; Sellars et al., 2002; Myers & Kelly, 2005; Weerakoon et al., 2009). The enzyme responsible for the oxidation of sulphite to sulphate and the release of electrons applied into the respiratory chain is a molybdenum-containing sulphite:cytochrome c oxidoreductase (SOR) which is composed of Cj0005c and Cj0004c and has homology to the corresponding enzyme of *Starkeya novella*. The two subunits were shown to constitute a functional SOR which respires sulphite in the presence of oxygen. The SOR-generated electrons, in turn, enter the respiratory chain after the bc1 complex at the level of cytochrome c (Kappler et al., 2000; Kappler & Dahl, 2001; Myers & Kelly, 2005).

In this report, we describe a *C. jejuni* cj0005c mutant that has strongly reduced motility and, therefore, a reduced capacity to invade host cells. In contrast to other motility-reduced mutants, no gene of the flagellar apparatus itself is affected, but SOR, an enzyme mediating the delivery of electrons into the respiratory chain leads to an impaired energy metabolism. Motility was shown to be important for *C. jejuni* to colonize and invade the respective host. Several studies in recent years have described the impact of genes and proteins of the flagellar apparatus on host colonization or the invasion of host cells in particular. One of the first publications, by Morooka et al. (1985), described that non-motile *C. jejuni* strains were cleared from the intestinal tract of suckling mice, whereas a wild-type strain colonized the lower portion of the intestine, the caecum and the colon. Furthermore, the colonization of rabbits was shown to depend on the flagellum, since a non-flagellated mutant was unable to survive inside the rabbit intestine (Pavlovskis et al., 1991). In addition to the studies mentioned above, the chicken intestinal tract was shown to be colonized only by fully motile strains of *C. jejuni*, which demonstrated that an intact flagellum is an important colonization factor (Nachamkin et al., 1993). Homologous recombination techniques were successfully applied to generate mutants that subsequently were investigated in cell culture experiments for their ability to invade host cells. Thereby, the importance of motility for the invasiveness of *C. jejuni* could be confirmed (Wassenaar et al., 1991; Yao et al., 1994; Syzmanski et al., 1995; Konkel et al., 2004). Meanwhile, many genes and proteins have been characterized which contribute to an intact flagellum in order to guarantee motility and, therefore, infectivity (Hendrixson, 2008). In order to find a link between the absence of a functional SOR on the one hand, and the reduced motility of the mutant on the other hand, we investigated the transcription levels of genes known to be important for flagellar composition. We detected a significant down-regulation of genes *fliR* and *flhA*, which encode proteins shown to be essential for the expression of FlgD and FlgE2, and of *rpoN*, encoding the RNA polymerase σ54 subunit of the pathogen, known to be responsible for the expression of the *flgDE2* locus as well as for the expression of minor flagellin FlaB (Hendrixson et al., 2001; Hendrixson & DiRita, 2003). These results indicate that the motility of the SOR mutant is negatively affected, although we have no data on protein level regarding the proteins whose expression is controlled by the gene products of *fliR*, *flhA* or *rpoN*.

The finding that the loss of a gene which is responsible for the supply of electrons into the respiratory chain leads to reduced motility raised the question whether the reduced motility is linked to the impaired energy metabolism. However, when we compared the intracellular ATP level of B2::cj0005c with the wild-type strain and the cj0005c-complemented mutant, we detected no differences. In addition, the cell proliferation of the mutant was not

**Table 2.** Transcription level of genes responsible for the synthesis of LegAm and the gene encoding heat-shock protein DnaJ in *C. jejuni* wild-type strain B2 and mutant B2::cj0005c

<table>
<thead>
<tr>
<th>Gene (cj)</th>
<th>Crossing point (B2::cj0005c)</th>
<th>Crossing point (B2 wild-type)</th>
<th>Fold change (t-test)</th>
<th>Function</th>
</tr>
</thead>
<tbody>
<tr>
<td>dnaJ (cj1260c)</td>
<td>23.34 (± 0.09)</td>
<td>20.97 (± 0.05)</td>
<td>5.16 (0.0001)</td>
<td>Heat shock response</td>
</tr>
<tr>
<td>ptmA (cj1332)</td>
<td>22.29 (± 0.55)</td>
<td>20.79 (± 0.17)</td>
<td>2.82 (0.0001)</td>
<td>LegAm synthesis</td>
</tr>
<tr>
<td>ptmB (cj1331)</td>
<td>19.40 (± 0.77)</td>
<td>16.83 (± 0.01)</td>
<td>5.93 (0.0001)</td>
<td>LegAm synthesis</td>
</tr>
<tr>
<td>ptmC (cj1327)</td>
<td>18.31 (± 0.47)</td>
<td>16.20 (± 1.19)</td>
<td>4.31 (0.0001)</td>
<td>LegAm synthesis</td>
</tr>
<tr>
<td>ptmD (cj1328)</td>
<td>20.53 (± 0.59)</td>
<td>20.16 (± 0.72)</td>
<td>1.29 (0.39)</td>
<td>LegAm synthesis</td>
</tr>
<tr>
<td>ptmE (cj1329)</td>
<td>18.11 (± 0.06)</td>
<td>15.17 (± 0.38)</td>
<td>7.67 (0.0001)</td>
<td>LegAm synthesis</td>
</tr>
<tr>
<td>ptmF (cj1330)</td>
<td>19.74 (± 0.50)</td>
<td>18.30 (± 0.89)</td>
<td>2.71 (0.0001)</td>
<td>LegAm synthesis</td>
</tr>
<tr>
<td>ptmG (cj1324)</td>
<td>19.16 (± 0.25)</td>
<td>17.64 (± 0.15)</td>
<td>2.86 (0.0001)</td>
<td>LegAm synthesis</td>
</tr>
<tr>
<td>ptmH (cj1325/6)</td>
<td>20.17 (± 0.26)</td>
<td>18.82 (± 0.12)</td>
<td>2.54 (0.017)</td>
<td>LegAM synthesis</td>
</tr>
</tbody>
</table>

The crossing point data are means ± SD. Strongly downregulated genes are in bold. See text for details.
negatively affected when the bacteria were grown in MH broth. Only in the presence of sodium sulphite was the mutant’s growth clearly impaired, probably due to the reduced ability of the mutant to clear this potentially antimicrobial chemical.

Surprisingly, when we eliminated motility as a virulence-associated factor by centrifuging the parental strain B2 and its mutant directly onto the host cells, we were able to shift the infectivity of the mutant from 29.8% to only 48.6%, indicating that not only reduced motility is responsible for the mutant’s impaired invasiveness compared to the wild-type strain. To obtain a more detailed insight into the mechanism of host cell invasion, adhesion assays were carried out. These experiments detected a reduced capacity of mutant B2::cj0005c compared to the wild-type strain in this initial host–pathogen interaction. Since many C. jejuni genes which contribute to the virulence of the pathogen and to the process of adhesion in particular have been described, we investigated whether the loss of a functional SOI has an impact on the transcription level of these genes. C. jejuni genes involved in stress responses, motility or chemotaxis were shown to be essential for the invasion of host cells. In addition, genes important for binding and adhesion were examined. Associated with the gene encoding the heat-shock protein DnaJ, the transcription of several genes responsible for the synthesis of legionaminic acid (LegAm) were shown to be significantly downregulated in the mutant B2::cj0005c.

The acetoamidino form of LegAm together with pseudaminic acid are two distinct nine-carbon sugars, which are O-linked to flagellin in most C. jejuni strains (Logan et al., 2008). Altogether eight genes have been identified which are required for the synthesis of LegAm. These so-called post-translational modification (ptm) genes are cj1324 (ptmG), cj1325/6 (ptmH), cj1327–cj1330 (ptmC, ptmD, ptmE, ptmF), cj1331 (ptmB) and cj1332 (ptmA) (McNally et al., 2007). Although knowledge of the importance of O-glycosylation by LegAm is limited, recent investigations by Howard et al. (2009) gave a first insight into its biological relevance. A C. jejuni cj1324 mutant showed full motility, but less hydrophobicity and impaired autoagglutination and biofilm formation due to the absence of two LegAm glycan modifications. Furthermore, this mutant had a reduced capacity to colonize chickens. When we investigated the transcription level of the LegAm genes in the cj0005c mutant, we detected only a 2.86-fold downregulation of cj1324 (ptmG). However, we also could clearly show reduced autoagglutination but no alterations in biofilm formation (data not shown). In contrast to Howard et al. (2009), we did not investigate the genes cj1321–cj1323, but we examined all genes required for LegAm synthesis with regard to altered transcription. This revealed transcription of ptmb (cj1331), ptmc (cj1327) and ptme (cj1329) to be strongly downregulated in the cj0005c mutant, whereas the transcription of genes ptma (cj1332), ptme (cj1330) and ptmg (cj1324) was significantly downregulated but to a lesser extent. However, we have no data regarding the corresponding proteins, and the data obtained so far and the resulting conclusions refer exclusively to our findings of reduced transcription. Since we have not performed transcription analysis covering the whole genome of the pathogen, alterations in the transcription level of other genes are likely. But the results obtained so far might indicate a correlation between the mutant’s altered biological properties (in particular diminished adhesion and autoagglutination) and the downregulation of genes responsible for LegAm synthesis.

In this study, we have characterized an invasion-deficient mutant, which owes its particular phenotype not to a mutation in a gene directly responsible in the process of motility, adherence or infection, but to a mutation within a gene involved in the energy metabolism of the pathogen. Despite this, we demonstrated an influence on events that are necessary for host cell invasion. It cannot be excluded that the reduced motility and the downregulation of virulence-associated genes are connected to the attempt of the pathogen to maintain the intracellular ATP level even in the absence of an enzyme which provides electrons for the respiratory chain. Other C. jejuni mutants with restricted energy metabolism should be examined in the future in order to investigate if a constant ATP level accompanying reduced biological functions and downregulated transcription is a more general phenomenon in C. jejuni.

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