Partial redundancy in the synthesis of the D-arabinose incorporated in the cell wall arabinan of Corynebacterineae

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The major cell wall carbohydrate of Corynebacterineae is arabinogalactan (AG), a branched polysaccharide that is essential for the physiology of these bacteria. Decaprenylphosphoryl-D-arabinose (DPA), the lipid donor of D-arabinofuranosyl residues of AG, is synthesized through a series of unique biosynthetic steps, the last one being the epimerization of decaprenylphosphoryl-β-D-ribose (DPR) into DPA, which is believed to proceed via a sequential oxidation–reduction mechanism. Two proteins from Mycobacterium tuberculosis (Rv3790 and Rv3791) have been shown to catalyse this epimerization in an in vitro system. The present study addressed the exact function of these proteins through the inactivation of the corresponding orthologues in Corynebacterium glutamicum (NCgl0187 and NCgl0186, respectively) and the analysis of their in vivo effects on AG biosynthesis. We showed that NCgl0187 is essential, whereas NCgl0186 is not. Deletion of NCgl0186 led to a mutant possessing an AG that contained half the arabinose and rhamnose, and less corynomycolates linked to AG but more trehalose mycolates, compared with the parental strain. A candidate gene that may encode a protein functionally similar to NCgl0186 was identified in both C. glutamicum (NCgl1429) and M. tuberculosis (Rv2073c). While the deletion of NCgl1429 had no effect on AG biosynthesis of the mutant, the gene could complement the mycolate defect of the AG of the NCgl0186 mutant, strongly supporting the concept that the two proteins play a similar function in vivo. Consistent with this, the NCgl1429 gene appeared to be essential in the NCgl0186-inactivated mutant. A detailed bioinformatics analysis showed that NCgl1429, NCgl0186, Rv3791 and Rv2073c could constitute, with 52 other proteins belonging to the actinomycetales, a group of closely related short-chain reductases/dehydrogenases (SDRs) with atypical motifs. We propose that the epimerization of DPR to DPA involves three enzymes that catalyse two distinct steps, each being essential for the viability of the bacterial cells.

Abbreviations: AG, arabinogalactan; Ara, arabinose; Araf, arabinofuranosyl; DPA, decaprenylphosphoryl-D-arabinose; DPPR, decaprenyl-monophosphoryl-5-phosphoribose; DPR, decaprenyl-monophosphoryl-β-D-ribose; Gal, galactose; mAGP, mycolyl-arabinogalactan-peptidoglycan; PAD, pulsed amperometric detector; ppRp, 5-phosphoribofuranosyl pyrophosphate; Rha, rhamnose; SDR, short-chain dehydrogenase/reductase; Suc, sucrose; TDCM, trehalose dicorynomycolate; TFA, trifluoroacetic acid; TMCM, trehalose monocorynomycolate.

Two supplementary tables, listing the primers used in this study and the orthologous group of proteins obtained by computational analysis using protein sequence alignments of strains examined in this study, are available with the online version of this paper.
INTRODUCTION

The Corynebacterineae is a suprageneric actinomycetes group that includes the causative agents of the human diseases tuberculosis, leprosy and diphtheria, Mycobacterium tuberculosis, Mycobacterium leprae and Corynebacterium diphtheriae, respectively. The cell wall core of these bacteria is made up of an unusual matrix composed of a cross-linked peptidoglycan covalently bound to arabinogalactan (AG) chains, which in turn are linked to mycolic acids, allowing the attached lipids to form the inner leaflet of an outer membrane. This complex is often referred to as mycolyl-arabinogalactan-peptidoglycan (or mAGP) complex (Daffe & Draper, 1998).

Structural motifs of the AG polymer were first established for M. tuberculosis (Daffe et al., 1999). However, except for some variations in glycosyl linkage compositions, notably in the arabinosyl portion, the major structural features of AG are conserved among all Corynebacterineae examined so far (Daffe et al., 1993). In mycobacteria the polysaccharide is composed of two distinct regions: a homogalactan portion, composed of alternating 5-linked and 6-linked β-galactofuranosyl residues (Daffe et al., 1999, 1993), which is covalently linked to the peptidoglycan via a disaccharide phosphate (McNeil et al., 1990), and arabinan chains, formed from arabinofuranosyl (Araf) residues, attached to carbon 5 of some of the 6-linked galactosyl (Gal) residues (Daffe et al., 1990) at their reducing termini (Besar et al., 1995) and esterified with mycolic acids at their non-reducing termini (McNeil et al., 1991). In addition to galactose (Gal) and arabinose (Ara), rhamnose (Rha) is also found in AG of Corynebacterium glutamicum (Alderwick et al., 2005; Gebhardt et al., 2007).

In the last few years an increasing number of studies have shown that C. glutamicum constitutes a good model for investigating biosynthetic pathways of specific cell wall compounds of Corynebacterineae, such as mycolic acids and AG. Indeed, while these components are essential for the viability of mycobacteria, C. glutamicum mutants either totally or severely deprived of mycolates or synthesizing a truncated AG devoid of the arabinan domain are viable, although they grow slowly (Alderwick et al., 2005, 2006b; Portevin et al., 2004, 2005; Tropis et al., 2005).

The synthesis of the arabinan domain of AG results from the sequential addition of Ara residues to the galactan portion, composed of alternating 5-linked and 6-linked β-D-galactofuranosyl residues (Daffe et al., 1990). In mycobacteria the polysaccharide is composed of two distinct regions: a homo-galactan portion, composed of alternating 5-linked and 6-linked β-D-galactofuranosyl residues (Daffe et al., 1999, 1993), which is covalently linked to the peptidoglycan via a disaccharide phosphate (McNeil et al., 1990), and arabinan chains, formed from arabinofuranosyl (Araf) residues, attached to carbon 5 of some of the 6-linked galactosyl (Gal) residues (Daffe et al., 1990) at their reducing termini (Besar et al., 1995) and esterified with mycolic acids at their non-reducing termini (McNeil et al., 1991). In addition to galactose (Gal) and arabinose (Ara), rhamnose (Rha) is also found in AG of Corynebacterium glutamicum (Alderwick et al., 2005; Gebhardt et al., 2007).

form decaprenylphosphoryl-5-phosphoribose (DPPR); (ii) removal of the 5′ phosphate giving decaprenylphosphoryl ribose (DPR); and (iii) epimerization of DPR to DPA, which likely occurs via a sequential oxidation–reduction mechanism involving an intermediate (DPX), product of DPR oxidation and precursor of DPA (Fig. 1). Although not unambiguously identified, DPX is probably a decaprenylphosphoryl-2-keto-β-D-erythro-pentofuranose.

The only enzyme identified in the DPA biosynthesis pathway by gene inactivation and mutant analysis is the DPPR synthase, the product of the essential M. tuberculosis gene Rv3806c (Huang et al., 2005), and named UbIA in C. glutamicum (Alderwick et al., 2005). Inactivation of ubia in this bacterium produces a mutant totally devoid of arabinan (Alderwick et al., 2005). Very recently, on the basis of the similarity with proteins implicated in the arabinosylation of the Azorhizobium caulinodans nodulation factor, Mikusova et al. (2005) have proposed that M. tuberculosis genes Rv3790 and Rv3791, which are both annotated as probable oxidoreductases, could encode the two enzymes expected to be involved in the epimerization of DPR into DPA. The two genes have been cloned and expressed in Escherichia coli, and the corresponding proteins purified. The use of an in vitro assay has shown that the presence of both proteins is required to transform DPR to DPA, suggesting that the two proteins work in concert to catalyse the epimerization. It was thus interesting to address the question of the precise role of these enzymes in the biosynthesis of AG in vivo. As Rv3790 and Rv3791 are presumably essential in M. tuberculosis (Sassetti et al., 2003) we attempted to delete their C. glutamicum orthologues, NCgl0187 and NCgl0186, respectively, and characterize the resulting AG modifications.

METHODS

Strains and culture conditions. C. glutamicum (ATCC 13032 Res167) (Dusch et al., 1999) was cultured in Brain Heart Infusion (BHI) medium (Difco) at 30 °C. E. coli DH5α and Top10 (Invitrogen) were used for the construction of plasmids and were grown on Luria–Bertani (LB) medium (Difco) at 37 °C. Ampicillin, kanamycin (Km), chloramphenicol (Cm) and sucrose (Suc) were added when required at final concentrations of 10, 25, 30 μg ml−1 and 10% (w/v) (for E. coli), or 6 μg ml−1 and 10% (w/v) (for C. glutamicum), respectively. Transformation of C. glutamicum by electroporation was performed as described by Bonamy et al. (1990).

DNA manipulations. C. glutamicum chromosomal DNA was extracted as described by Ausubel et al. (1987). Oligonucleotide primers were synthesized by Genosys. All DNA sequencing was carried out by Cogenics.

Construction of plasmids and strains. In order to delete NCgl0186 or NCgl0187, we used the strategy described by Schäfer et al. (1994). In brief, two DNA fragments overlapping the gene to be deleted at its 5′ and 3′ extremities were amplified by PCR from C. glutamicum total DNA by using appropriate primers (186/1/186-2 and 186-3/186-4 for NCgl0186, and 187-1/187-2 and 187-3/187-4 for NCgl0187; see Supplementary Table S1) and cloned in the non-replicative vector pK18mobSacA186 and
pK18mobsacΔ187) were sequenced and independently transferred into C. glutamicum ATCC 13032 RES167 by electroporation. Transformants in which the construct was integrated into the chromosome by single crossing over were selected on BHI plates containing Km. The second crossover event was selected by plating KmR clones on BHI plates containing Suc. KmS and SucR colonies were screened by PCR for the correct deletion of the gene using primers binding upstream and downstream of the NCgl0186 sequence. After verification of PCR products by sequencing, one strain carrying the NCgl0186 deletion (Cg-D186) was selected for further studies.

Construction of a mutant strain carrying a deletion of the NCgl0187 gene was not successful. We thus tried to inactivate this gene. For this purpose, an internal fragment was amplified using primers 187-5 and 187-6 (see Supplementary Table S1) and cloned into plasmid pCR 2.1-TOPO using the TOPO TA cloning kit (Invitrogen) to give plasmid TOPO : : 187. This plasmid was used to electrotransform the ATCC 13032 RES167 strain. No transformant carrying the correct plasmid integration could be detected after selection on BHI plates containing Km and PCR analysis.

Deletion of the NCgl1429 gene was done using a strategy described previously (Portevin et al., 2004). In brief, two DNA fragments overlapping the NCgl1429 gene at its 5′ and 3′ extremities were amplified by PCR from C. glutamicum total DNA using primers 1429-AMXho/1429-AMSac and 1429-AVSac/1429-AVBgl, respectively (Supplementary Table S1). These fragments were inserted flanking a km resistance cassette into plasmid pMCS5 (MoBiTec). The resulting plasmid was transferred into C. glutamicum ATCC 13032 RES167 by electroporation, and transformants were selected on plates containing Km. Transformants in which allelic replacement had occurred were selected by PCR analysis using combinations of primers binding upstream and downstream of NCgl1429 and in the aphIII sequences. After sequencing of the PCR products, one strain (Cg-D1429) was selected for further studies.

For inactivation of NCgl1429 in Cg-D186, two plasmids were constructed. First, an internal fragment was amplified using primers 1429-1 and 1429-2 (see Supplementary Table S1) and cloned into plasmid pCR2.1-TOPO to give plasmid TOPO::187. This plasmid was used to electrotransform the ATCC 13032 RES167 strain. No transformant carrying the correct plasmid integration could be detected after selection on BHI plates containing Km and PCR analysis.

Deletion of the NCgl1429 gene was done using a strategy described previously (Portevin et al., 2004). In brief, two DNA fragments overlapping the NCgl1429 gene at its 5′ and 3′ extremities were amplified by PCR from C. glutamicum total DNA using primers 1429-AMXho/1429-AMSac and 1429-AVSac/1429-AVBgl, respectively (Supplementary Table S1). These fragments were inserted flanking a km resistance cassette into plasmid pMCS5 (MoBiTec). The resulting plasmid was transferred into C. glutamicum ATCC 13032 RES167 by electroporation, and transformants were selected on plates containing Km. Transformants in which allelic replacement had occurred were selected by PCR analysis using combinations of primers binding upstream and downstream of NCgl1429 and in the aphIII sequences. After sequencing of the PCR products, one strain (Cg-D1429) was selected for further studies.

For inactivation of NCgl1429 in Cg-D186, two plasmids were constructed. First, an internal fragment was amplified using primers 1429-1 and 1429-2 (see Supplementary Table S1) and cloned into plasmid pCR2.1-TOPO to give plasmid TOPO::1429. Then, a fragment corresponding to the entire NCgl1429 ORF under the cspB promoter was amplified from plasmid pCGL2330 (see below), digested with HindIII and SseI, and ligated to TOPO::1429 digested with the same restriction enzymes. The resulting plasmids, TOPO::1429-cspB186 and TOPO::1429, were both used to electrotransform strain Cg-D186, and transformants were selected

Fig. 1. The biosynthetic pathway proposed for DPA synthesis from pRpp (Mikusova et al., 2005). The isoprenoid moiety is drawn to conform to the structure of decaprenyl phosphate identified in mycobacteria. The structure of the corneybacterial equivalent is not known. Although the structure of DPX is not certain, this intermediate is very likely a decaprenylphosphoryl-2-keto-β-D-erythropentofuranose. The role of NCgl2782 (Rv3807c) in the dephosphorylation step is putative and arises from the position of the gene on the chromosome, just beside ubiA (Rv3806c), and from its annotation as a phosphatase.
on BHI plates containing Km. Genomic DNAs from mutants were analysed by three PCR amplifications using two primers binding upstream of the 5' end and downstream of the 3' end of the NCGl1429 gene, and the M13 forward and reverse primers binding in the pCR2.1-TOPO vector. The PCR products corresponding to the gene disruption borders were sequentially sequenced.

Expression vectors encoding NCgl0186 (pCGL2330), Rv3791 (pCGL2331) and NCgl1429 (pCGL2333) were constructed using pCGL482 (Peyret et al., 1993) as the cloning vector. We chose to clone these three ORFs under the control of the cspB promoter (Peyret et al., 1993). Coding sequences from C. glutamicum ATCC 13032 chromosomal DNA were amplified by PCR using primer pairs 187-3/186-7 for NCgl0186 and 1429-3/1429-4 for NCgl1429. The Rv3791 ORF was amplified using primers 3791-1 and 3791-2 from the pSTbule1-Rv3791 plasmid (Mikusova et al., 2005). A DNA fragment containing the cspB promoter was generated using primers pcspBBam and pcspBNco and pCGL824 (Peyret et al., 1993) as template DNA. All the ampiclons were digested with the appropriate restriction endonucleases (see Supplementary Table S1). Plasmids were obtained by simultaneous ligation of fragments containing the cspB promoter, fragments containing one ORF and the appropriately digested pCGL482. Transformants were selected on Cm-containing plates.

**AG preparation.** Cell walls were prepared as described previously (Daffe et al., 1990). Briefly, wet cells were resuspended in PBS buffer (50 mM, pH 7.2) at the concentration of 5 g in 20 ml and broken in a cell disruptor (2.7 x 10^7 Pa). After three cell disrupter cycles, intact cells were removed by centrifugation (3000 g) and the supernatant containing the cell walls was then centrifuged (27000 g, 1 h). The resulting cell wall pellet was treated with aqueous 2 % (w/v) SDS at 95 °C for 1 h and sedimented again at 27000 g. This was followed by two acetone washes and the pellet thus obtained was mAGP. mAGP was further treated with 2 M NaOH and stirred gently at 80 °C for 16 h to remove the mycolic acid residues and cleave peptidoglycan to solubilize AG. The pellet was discarded and the pH of the supernatant was adjusted to 7.0 with glacial acetic acid. This neutralized AG was dialysed for at least 3–4 h with three 1 l changes of MilliQ water. The AG obtained was concentrated by rotary evaporation to a final volume of 1 ml, and was freeze-dried and stored till further use.

**Enzymic treatments of soluble AG.** The soluble AG was treated with partially purified endogenous arabinase (Dong et al., 2006) at 37 °C for 12 h, followed by extraction with CHCl3/CH3OH/H2O (10:10:3, by vol.). After centrifugation, the supernatant, which contained most of the released arabinoxylans, was used for MALDI-TOF MS analysis.

To further elucidate the cell wall AG configuration, the soluble AG was treated with *Cellulomonas* arabanase obtained as described previously (McNeil et al., 1994), which specifically releases Ara2 and Ara4 from AG.

**Analysis of AG.** The glycosyl composition of purified cell walls was determined by hydrolysing an aliquot of mAGP with 2 M trifluoroacetic acid (TFA) for 2 h at 110 °C, followed by trimethylsilylation (Sweeney et al., 1963) and GC analysis of the resulting products, or by alditol acetate formation, as described previously (Daffe et al., 1990).

HPLC analysis of soluble AG samples treated with arabinase was carried out on a Dionex system equipped with a CarboPac PA1 analytical column (4 x 250 mm). Total run time was 50 min, the NaOH concentration (100 mM) being consistent throughout. Sodium acetate concentration was varied from 0 to 450 mM for a period of 50 min. The eluted samples were detected using a pulsed amperometric detector (PAD).

For MALDI-TOF MS analysis, samples were peracetylated with acetic anhydride (100 μl) in the presence of pyridine (100 μl) at room temperature overnight and recovered after removal of the solvent in chloroform prior to MALDI-TOF MS. The acetylated AG samples were run in positive ion reflector mode on a Bruker Ultraflex MALDI-TOF/TOF mass spectrometer (Bruker Daltonics). One microtitre of the sample was mixed with 1 μl matrix (10 mg ml^-1 dihydroxybenzoic acid in 50 % acetonitrile/0.1 % TFA) and allowed to air dry on the MALDI target plate.

For 1H- and 13C-NMR analysis, AGs were solubilized in H2O (20 ml) and analysed at 600 and 150.9 MHz, respectively, on a Bruker Avance 600 MHz NMR spectrometer equipped with a TCI cryoprobe. 1H-decoupled 13C spectra were obtained with a DEPT 135 pulse sequence in a DQD acquisition mode.

**Extraction and quantification of corynomycolic acids.** Lipids were extracted from wet cells for 16 h with CHCl3/CH3OH (1:2, v/v) at room temperature; the cells were re-extracted with CHCl3/CH3OH (1:1, v/v) and CHCl3/CH2OH (2:1, v/v) for 16 h. The three organic phases were pooled and concentrated by means of rotary evaporation. The crude lipid extracts were partitioned between the aqueous and the organic phases arising from a mixture of CHCl3/H2O (1:1, v/v). The lower organic phases were collected and evaporated to dryness to yield the crude lipid extracts from each strain. Subsequently, they were examined comparatively using TLC on silica gel-coated plates (Durasil-25, 0.25 mm thickness, Macherey–Nagel) developed with CHCl3 and CH3OH/H2O (30:8:1 or 65:25:4, by vol.). Lipids were detected by spraying plates with 0.2 % anthrone in concentrated H2SO4, followed by heating. The corynomycolate content of extractable lipids was determined in three independent experiments as follows. Lipid extracts (100 mg) of the various strains were dried under vacuum and saponified (Daffe et al., 1983); the saponified products were acidified with 20 % H2SO4. The resulting fatty acids were extracted with diethyl ether, washed with water, converted to methyl esters with diazomethane and dried under vacuum, and weighed. The fatty acid methyl esters were separated from contamination on a silica gel column irrigated with different concentrations of diethyl ether in petroleum ether (0, 5, 10, 20 and 100 %, v/v). Fractions were analysed by TLC developed with CH3Cl2. Lipids were detected by spraying plates with rhodamine B and fractions containing corynomycolates were pooled and weighed. The AG-bound mycolic acids present in the delipidated cells, i.e. the material after the CHCl3/CH3OH extraction (described above), were saponified with 40 % KOH in 2-methoxyethanol (1:7, v/v) at 110 °C for 3 h (Daffe et al., 1983) and the solution was neutralized using 20 % H2SO4. The released mycolic acids were extracted with diethyl ether and methylated with diazomethane. The resulting fatty acid methyl esters were examined using TLC on silica gel-coated plates developed as described above. When significant amounts of fatty acids other than mycolic acids were present in the saponification products, corynomycolates were isolated by chromatography on a silica gel column as described above and weighed.

**Epimerase assay.** The cell wall or cell membrane (70 μl, 5.5 mg protein ml^-1), 10 mM ATP (1 μl), 10 mM each of FAD, NAD and NADP (3 μl each), and [34Cl]Rpp (20 μl, 10 000 c.p.m. μl^-1) were added to MOPS buffer (pH 8 with MgCl2) to give a final volume of 160 μl. The reaction mixture was incubated for 90 min at 37 °C. Subsequently, the mixture was extracted with CHCl3/CH3OH/H2O (8:4:3, by vol.), centrifuged briefly at 4500 g, and the organic layer was separated and dried. This was subjected to TFA (2 M) hydrolysis for 1 h at 110 °C, cooled, dried down in CH3OH and analysed for Ara and ribose content by HPLC. The samples (neutral sugars after TFA hydrolysis) were analysed using a CarboPac PA01 analytical column (4 x 250 mm). The sugars were separated with an isocratic solvent system consisting of 10 % NaOH (100 mM) in water for 45 min. The
14C-containing components were detected using a Beta ram detector. The same HPLC system was connected to a PAD to detect non-radioactive standards of the respective sugars for comparative quantification.

RESULTS

Genomic comparison of the 3790–3791 locus
Data from Mikusova et al. (2005) strongly suggest that in M. tuberculosis the products of Rv3790 and Rv3791 work in concert to catalyse the conversion of DPR to DPA. Rv3790 and Rv3791 are located upstream of the genes encoding the arabinosyltransferases AftA, EmbA, EmbB and EmbC (Fig. 2). All these genes are part of a large cell wall biosynthesis cluster containing, among others, Rv3806c, which encodes the enzyme involved in the first step of DPA synthesis, the transfer of a 5-phosphoribosyl residue from pRpp to decaprenyl phosphate to form DPPR (Huang et al., 2005). An in silico analysis revealed that orthologues of Rv3790 and Rv3791 are present in all the available genome sequences of Corynebacterineae. The two genes are always organized in a putative operon and, except in Corynebacterium jeikeium, are adjacent to the aftA-emb cluster (Fig. 2). In C. glutamicum, Rv3790 and 3791 orthologues correspond to genes NCgl0187 and NCgl0186, respectively. Sequence comparisons at protein level show 65% identity for Rv3790 and NCgl0187 and 57% identity for Rv3791 and NCgl0186. As Rv3790 and Rv3791 are presumably essential in M. tuberculosis (Sassetti et al., 2003), attempts were made to delete their C. glutamicum orthologues in order to address the question of the functions of the corresponding enzymes in the biosynthesis of AG in vivo, a strategy already successfully used to decipher the functions of mycobacterial essential genes (Alderwick et al., 2006a; Portevin et al., 2004, 2005).

Construction and growth characteristics of CgΔ186
In an attempt to delete either NCgl0186 or NCgl0187, non-replicative pK18mobsac-derivative plasmids were constructed (pK18mobsacΔ186 and pK18mobsacΔ187), each carrying sequences adjacent to the gene to be deleted (Schafer et al., 1994). KmS and SucR clones resulting from two recombination events between plasmid and chromosome were obtained for both deletions. In the case of NCgl0186, 50 clones were analysed by PCR and half of them were found to have a gene deletion at the expected position. In comparison to the wild-type strain, however, the ΔNCgl0186 mutant (thereafter named CgΔ186) exhibited no significant phenotypic changes, had similar growth and did not aggregate in liquid cultures. These results indicated that although NCgl0186 may be involved in the synthesis of DPA, the mutant strain has a behaviour very different from that previously observed for the mycolate-less or AG-truncated mutants, i.e. a rough and matte colony phenotype, instead of the smooth and shiny one of the wild-type, and very slow growth and strong aggregation in liquid cultures (Alderwick et al., 2006a; Portevin et al., 2004, 2005).

In the case of NCgl0187 about 200 clones were analysed by PCR but, unlike NCgl0186, all of the clones showed the wild-type genotype, indicating either a strong disadvantage of NCgl0187 deletion or the essentiality of the gene even in the corynebacterial host. Furthermore, even when the analysis was continued up to 3 weeks to eventually obtain slow-growing rough-textured colonies as described for emb and aftA mutants in C. glutamicum (Alderwick et al., 2005, 2006b), no such colonies were visible. As no deletion mutant could be obtained for NCgl0187, we attempted to inactivate the gene by plasmid integration, but no integration mutants could be obtained by this procedure.
either. We thus concluded that NCg0187 is likely an essential gene.

**Glycosyl compositional analysis of the cell wall and AG from Cg-Δ186 and ATCC 13032 strains**

Purified cell walls from Cg-Δ186 and its parental ATCC 13032 wild-type of *C. glutamicum* were analysed for glycosyl composition. As shown in Table 1, Cg-Δ186 still produced Ara but contained only half of the amount observed in the wild-type strain, resulting in a lower Ara:Gal ratio than that of the ATCC 13032 strain. A similar diminution of the Rha:Gal ratio was also observed for the mutant strain, suggesting that the rhamnolysyl residues are linked to the arabinian moiety of AG. Confirmation of these data came from the analysis of AG released from the peptidoglycan using base treatment, as described previously for mycobacterial cell walls (Daffe´ et al., 1990), which showed a twofold decrease in the Ara:Gal and Rha:Gal ratios of the mutant compared with those of the parental strain (data not shown).

Complementation of Cg-Δ186 with plasmid pCGL2330 (containing NCg0186) led in both cases to the restoration of a normal level of Ara (Table 1). This indicated that the rhamnolysyl protein exhibited an in vivo function similar to that of the corynebacterial NCg0186. The Rha:Gal ratio was, however, only slightly increased (10%) by complementation of Cg-Δ186 with the plasmid containing NCg0186 and unchanged by the presence of a plasmid containing Rv3791, a phenomenon likely to be due to a problem with co-regulation of the synthesis of the Rha-containing arabinian segment of AG. We thus concluded that NCg0186 is directly or indirectly involved in the reaction of epimerization of DPR into DPA, but that another enzyme possessing a similar function is likely to be present in *C. glutamicum*. These data are consistent with the normal growth phenotype observed for the Cg-Δ186 strain.

**Table 1.** Quantitative analysis of the sugar constituents of the cell walls extracted from *C. glutamicum* ATCC 13032, its isogenic ΔNCg0186 (Cg-Δ186) mutant and the mutant complemented with either the corynebacterial gene [Cg-Δ186(pCGL2330)] or its mycobacterial orthologue [Cg-Δ186(pCGL2331)]

<table>
<thead>
<tr>
<th>Strain</th>
<th>Ara : Gal</th>
<th>Rha : Gal</th>
</tr>
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<tbody>
<tr>
<td>Wild-type</td>
<td>4.50 ± 0.5</td>
<td>1.0 ± 0.1</td>
</tr>
<tr>
<td>Cg-Δ186</td>
<td>2.20 ± 0.2</td>
<td>0.56 ± 0.05</td>
</tr>
<tr>
<td>Cg-Δ186(pCGL2330)</td>
<td>4.60 ± 0.5</td>
<td>0.60 ± 0.05</td>
</tr>
<tr>
<td>Cg-Δ186(pCGL2331)</td>
<td>4.30 ± 0.3</td>
<td>0.60 ± 0.05</td>
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In **vivo** epimerization of DPR into DPA by cell membrane and cell wall from Cg-Δ186 and wild-type strains

To prove the direct involvement of NCg0186 in DPA biosynthesis in *C. glutamicum* we determined the effect of mutation of the coding gene on the epimerization of DPR into DPA. Thus, both membranes and cell walls prepared from Cg-Δ186 and its parental strain, ATCC 13032, were independently incubated with radioactive [14C]Rpp and decaprenyl phosphate. The resulting mixtures of [14C]R and DP[14C]A were hydrolysed and the radioactive sugars released were analysed by HPLC. The epimerization of DPR into DPA was observed when wild-type cell membranes and, to a lesser extent, wild-type cell walls were used as enzyme sources (Table 2). When the experiments were done with Cg-Δ186 cell walls and cell membranes we observed a 50% decrease in the DPA formed as compared with the wild-type. This change was partially restored by complementation with NCg0186. Overexpression of NCg0186 in the wild-type bacterium did not lead to increased epimerization (Table 2), probably due to the fact that NCg0186 was not limiting for epimerization in the wild-type bacterium. These data established the direct involvement of NCg0186 in the epimerization of DPR into DPA and confirmed the contribution of another enzyme with similar function.

**Structural analysis of the AG from the cell walls of Cg-Δ186 and wild-type strains**

In order to investigate the potential effects of NCg0186 deletion on the structure of AG we performed different types of analysis. First, the soluble AG from Cg-Δ186 and wild-type were treated with an arabinase partially purified from *Mycobacterium smegmatis* (Dong et al., 2006), and the reaction products were analysed by MALDI-TOF MS. Similar fragments were observed in the degradative products of the AG from the wild-type and Cg-Δ186 strains; they were composed of oligomers of Ara of up to nine residues. Second, the soluble AGs from both strains were treated with a mixture of arabinases and galactofuranases from *Cellulomonas* (McNeil et al., 1994), and analysed by HPLC and MALDI-TOF MS. Again, no significant difference could be detected between the patterns obtained from Cg-Δ186 and ATCC 13032 strains by this method (data not shown). Consistent with these observations, the glycosyl-linkage compositions of the AG from the wild-type and mutant strains of *C. glutamicum* were very similar to one another and to those of other strains of the species (Alderwick et al., 2005; Gebhardt et al., 2007). Finally, since 13C-NMR spectroscopy of AGs has proven to be an appropriate tool for comparative qualitative analysis of AGs, AG from the two isogenic strains was compared by this technique (Daffe´ et al., 1990, 1993; Gebhardt et al., 2007). The 13C-NMR spectra obtained from Cg-Δ186 and ATCC 13032 were superimposable (data not shown) and identical to that published.
previously for another strain of *C. glutamicum* (Gebhardt *et al.*, 2007). We thus concluded that the AGs from the two strains were composed of the same structural motifs.

**Lipid analysis of the Cg-Δ186 and wild-type strains**

In view of the decrease in the Ara content of the AG from Cg-Δ186, we reasoned that this might affect the lipid profile of the mutant, notably the amount of corynomycolates linked to AG. Accordingly, we performed a comparative lipid analysis of Cg-Δ186 and the wild-type strain, focusing on corynomycolate-containing compounds, i.e. cell wall and trehalose monocorynomycolate (TMCM) and trehalose dicorynomycolate (TDCM) (Puech *et al.*, 2001). Cells were labelled with acetate and the lipids were extracted with organic solvents. Analysis of the extractable lipids by TLC and quantification of the corynomycolate esterifying trehalose showed that the mutant strain accumulated 57 % more corynomycolates than did the parent strain (Table 3). Concomitantly, the Cg-Δ186 strain contained 40 % less corynomycolates covalently bound to AG (Table 3). These data showed that NCgl0186 deletion affected the tethering of corynomycolates to AG, probably through the reduction of the number of sites of attachment of these lipids. Consistent with this finding, this phenotype was completely reversed as DPA is still produced by the Cg-Δ186 mutant, as confirmed by enzyme assay (Table 2). As a consequence, the occurrence of Ara in the mutant could be explained by the redundancy of NCgl0186 in *C. glutamicum* or, alternatively, NCgl0186 could have a function in corynebacteria different from that of Rv3791 in mycobacteria, despite the homology and the similar genomic environment of the two genes, i.e. Rv3791 and NCgl0186. However the last hypothesis seems unlikely in view of our data that show that the expression of Rv3791 is able to complement the deficit of Cg-Δ186 in Ara. In order to identify genes encoding one or more proteins functionally equivalent to NCgl0186 in *C. glutamicum*, BLAST searches were performed against this genome using NCgl0186 as a query sequence (Altschul *et al.*, 1990). On the basis of the score values, only one sequence, the product of the NCgl1429 gene, could be considered to encode a paralogous protein. This protein shows 35 % identity with NCgl0186 (Fig. 3). We checked for the occurrence of an NCgl1429 orthologue in the *M. tuberculosis* genome and found Rv2073c. As shown in Fig. 3 amino acid alignments of NCgl1429 and Rv2073c with NCgl0186 and Rv3791 revealed well-conserved blocks, and led us to propose NCgl1429 as the protein possessing a functional redundancy with NCgl0186. It should be noted that, in contrast to NCgl0186, no paralogue was found for NCgl0187 in *C. glutamicum*. We thus attempted to delete NCgl1429 both in ATCC 13032 and in Cg-Δ186.

A km resistance cassette flanked by arms about 500 bp identical to the 5' and 3' parts of NCgl1429 was cloned into

**Table 2.** Epimerization of DPR to DPA by cell membranes and cell walls of *C. glutamicum* ATCC 13032, its isogenic ΔNCgl0186 (Cg-Δ186) mutant and the complemented mutant Cg-Δ186(pCGL2330), and the wild-type strain in which NCgl0186 was overexpressed [ATCC 13032(pCGL2330)].

Epimerase activity was assayed by measuring the ability of cell wall or cell membrane extracts to transform pRpp into DPA. The percentages of Ara and ribose were determined relative to the masses of the delipidated cells and extractable lipids, respectively. The values are the means±SD of at least three independent experiments.

<table>
<thead>
<tr>
<th>Strain</th>
<th>Cell wall as enzyme source Ara (%)</th>
<th>Cell membrane as enzyme source Ara (%)</th>
<th>Ribose (%)</th>
<th>Ribose (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ATCC 13032</td>
<td>72±4</td>
<td>83±2</td>
<td>28±4</td>
<td>17±3</td>
</tr>
<tr>
<td>Cg-Δ186</td>
<td>41±3</td>
<td>51±3</td>
<td>59±3</td>
<td>49±3</td>
</tr>
<tr>
<td>Cg-Δ186(pCGL2330)</td>
<td>44±3</td>
<td>63±4</td>
<td>56±3</td>
<td>37±4</td>
</tr>
<tr>
<td>ATCC 13032(pCGL2330)</td>
<td>62±2</td>
<td>78±3</td>
<td>38±2</td>
<td>22±3</td>
</tr>
</tbody>
</table>

**Table 3.** Quantitative analysis of cell wall-linked and extractable corynomycolates from *C. glutamicum* ATCC 13032, its isogenic mutant ΔNCgl0186 (Cg-Δ186) and the mutant complemented with either the corynebacterial gene [Cg-Δ186(pCGL2330)] or its mycobacterial orthologue [Cg-Δ186(pCGL2331)]

The percentages of corynomycolates in cell walls and extractable lipids were determined relative to the masses of the delipidated cells and extractable lipids, respectively. The values are the means±SD of at least three independent experiments. Corynomycolates were isolated by saponification of the delipidated cells and extractable lipids, purified by chromatography and weighed, as described in Methods.

<table>
<thead>
<tr>
<th>Strain</th>
<th>Wall-linked corynomycolates (%)</th>
<th>Corynomycolates in extractable lipids (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ATCC 13032</td>
<td>1.0±0.1</td>
<td>33±2</td>
</tr>
<tr>
<td>Cg-Δ186</td>
<td>0.6±0.1</td>
<td>52±3</td>
</tr>
<tr>
<td>Cg-Δ186(pCGL2330)</td>
<td>1.1±0.1</td>
<td>29±2</td>
</tr>
<tr>
<td>Cg-Δ186(pCGL2331)</td>
<td>1.0±0.1</td>
<td>28±2</td>
</tr>
</tbody>
</table>

**Search for an NCgl0186 orthologue and construction of the corresponding mutant**

As DPA has been shown to be the only donor of Ara in *C. glutamicum* (Alderwick *et al.*, 2005), these results indicated that DPA is still produced by the Cg-Δ186 mutant, as confirmed by enzyme assay (Table 2). As a consequence, the occurrence of Ara in the mutant could be explained by the redundancy of NCgl0186 in *C. glutamicum* or, alternatively, NCgl0186 could have a function in corynebacteria different from that of Rv3791 in mycobacteria, despite the homology and the similar genomic environment of the two genes, i.e. Rv3791 and NCgl0186. However the last hypothesis seems unlikely in view of our data that show that the expression of Rv3791 is able to complement the deficit of Cg-Δ186 in Ara. In order to identify genes encoding one or more proteins functionally equivalent to NCgl0186 in *C. glutamicum*, BLAST searches were performed against this genome using NCgl0186 as a query sequence (Altschul *et al.*, 1990). On the basis of the score values, only one sequence, the product of the NCgl1429 gene, could be considered to encode a paralogous protein. This protein shows 35 % identity with NCgl0186 (Fig. 3). We checked for the occurrence of an NCgl1429 orthologue in the *M. tuberculosis* genome and found Rv2073c. As shown in Fig. 3 amino acid alignments of NCgl1429 and Rv2073c with NCgl0186 and Rv3791 revealed well-conserved blocks, and led us to propose NCgl1429 as the protein possessing a functional redundancy with NCgl0186. It should be noted that, in contrast to NCgl0186, no paralogue was found for NCgl0187 in *C. glutamicum*. We thus attempted to delete NCgl1429 both in ATCC 13032 and in Cg-Δ186.

A km resistance cassette flanked by arms about 500 bp identical to the 5’ and 3’ parts of NCgl1429 was cloned into
a vector unable to replicate in corynebacteria. This construct was transferred into both C. glutamicum ATCC 13032 and Cg-Δ186. In both cases kmR transformants were obtained and analysed by PCR using various combinations of primers. From the wild-type strain one clone (Cg-D1429) among 20 gave the amplification pattern consistent with allelic replacement of NCgl1429 with the km cassette. This exchange was the result of two recombination events. In the Cg-D186 host only the first recombination event, corresponding to plasmid integration, could be obtained, as revealed by the analysis of 200 clones. This absence of deletions led us to suspect that, as for NCgl0187, inactivation of both genes (i.e. NCgl0186 and NCgl1429) was not possible because of the essentiality of the function associated with the two proteins in the epimerization of DPR to DPA. To conclusively demonstrate this hypothesis we tried to inactivate NCgl1429 in Cg-Δ186 by plasmid integration in the presence or absence of a functional copy of NCgl0186. For that purpose a pCR2.1-TOPO plasmid derivative containing an ~500 bp NCgl1429 internal fragment was first constructed (TOPO::1429). We then transferred a copy of NCgl0186 under the control of the cspB promoter into this plasmid (TOPO::1429-cspB186). No integration mutants could be obtained when Cg-Δ186 was transformed with TOPO::1429, while integration took place very easily when TOPO::1429-cspB186 was used as integration vector. These data demonstrated that NCgl1429 is essential in the absence of NCgl0186.

### Analysis of the NCgl1429 mutant strain

Cell walls from Cg-Δ1429 were analysed for glycosyl composition and corynomycolate content as described for Cg-Δ186 and ATCC 13032. Surprisingly, and in sharp contrast with Cg-Δ186, no change in the Ara:Gal and Rha:Gal ratios could be detected in the mutant strain, compared with the wild-type. Likewise, no significant difference was observed in terms of amounts of extractable and bound corynomycolates of the mutant, compared with the parental ATCC 13032 strain. These results strongly suggest that in the presence of a functional NCgl0186, NCgl1429 has no detectable contribution in vivo. Based on the data obtained previously with the partially redundant mycoloyltransferases of M. tuberculosis (Puech et al., 2002),

![Multiple alignments of protein sequences of NCgl0186, Rv3791, NCgl1429, Rv2073c and the deduced consensus sequence (consensus 1). Identical residues are boxed and similar amino acids have light-grey backgrounds. The line above the sequence indicates the conserved block chosen for computational analysis.](image-url)
we tested the in vivo effect of NCgl1429 overexpression on the epimerization reaction in the absence of NCgl0186. For that purpose, a plasmid containing the NCgl1429 ORF under the control of the strong promoter of cspB was constructed (pCGL2333) and introduced into the Cg-Δ186 strain. As we have shown that the quantification of corynomycolates linked to AG reasonably reflects the complementation of the phenotype of the Cg-Δ186 mutant (Table 3), we performed a quantitative analysis of the cell wall corynomycolates of Cg-Δ186 (pCGL2333). The percentage of cell wall-linked corynomycolates of Cg-Δ186 (pCGL2333) was found to be 1 % (± 0.1), a value close to that obtained with the wild-type strain and with the Cg-Δ186 strain complemented with either NCgl0186 or Rv3791 (Table 3). This result indicated that overexpression of NCgl1429 in vivo could restore the defect in AG production by the absence of NCgl0186, thus pointing to a partial redundancy of the two enzymes similar to that demonstrated for the three mycoloyltransferases of M. tuberculosis (Puech et al., 2002).

Conservation and redundancy of NCgl0186 orthologous proteins among Corynebacterineae

NCgl0186, NCgl1429, Rv3791 and Rv2073c belong to the superfamily of short chain dehydrogenases/reductases (SDRs), which contains enzymes catalysing NAD(P)(H)-dependent oxido-reduction (Jornvall et al., 1995; Oppermann et al., 2003). This is a large and functionally heterogeneous protein family with low sequence identities but with conserved motifs and common structural organization. In bacteria the number of SDR members deduced from genome analysis varies considerably from one organism to another, but it should be noted that this number is especially large in M. tuberculosis (57 members) (Jornvall et al., 1999). We wanted to evaluate the level of sequence conservation and of redundancy of NCgl0186 orthologues among the Corynebacterineae. For this purpose, one of the conserved blocks, SsxAGxRxxNx-VYGSxKAGD (shown in Fig. 3), was chosen. It contains the three conserved residues of the catalytic active triad (S, Y and K) characteristic of SDR proteins (Oppermann et al., 2003). This motif was searched for in the NCBI Reference Sequence (RefSeq) collection (http://www.ncbi.nlm.nih.gov/RefSeq/). Forty-seven sequences were found to match with this motif totally and six more with one mismatch. Pair-wise alignments showed that 52 of the 53 sequences possessed between 36 and 100 % identities with Rv3791 and thus can be considered to be closely related. All these protein sequences belong to Actinomycetales, mostly to Corynebacterineae (see Supplementary Table S2). This analysis shows that all members of Corynebacterineae for which the genome sequence is known possess at least one, and the great majority two, orthologous genes.

The SDR superfamily is divided into five families on the basis of protein length and specific sequence motifs. According to their length (between 242 and 260 aa, except for MAP1819c from Mycobacterium paratuberculosis, which has only 187 residues) the proteins identified in this work are classical SDRs. CLUSTAL W alignment (Fig. 3) and secondary structure predictions performed on these protein sequences allowed the identification of only two SDR motifs specific to the classical family among the seven proposed by Kallberg et al. (2002) (Table 4). In particular, the typical pattern of three Gly residues (TGxGxGxG) in the N-terminal domain characteristic of the NAD(P)(H)-binding site is missing. Nevertheless, a conserved motif, G/G/AxxSxxGA, localized in the β1–α1 predicted structure that resembles the patterns of both intermediate SDRs (G/AxxGxxG/A) and divergent SDRs (GxxxxxxAxxxSxxA), could constitute an atypical dinucleotide-binding motif (Fig. 3, Table 4).

DISCUSSION

The mAGP complex, which constitutes the cell wall skeleton of the very singular cell envelope of Corynebacterineae, represents, with mycolic acids, the hallmark of all the members of this group of bacteria (Besra et al., 1995; Daffe, 2005; Dover et al., 2004; McNeil et al., 1991). Despite the physiological importance of this complex, its biosynthesis is far from being totally understood, mostly because of its essentiality for the viability of M. tuberculosis. However, thanks to the use of C. glutamicum as a model organism for Corynebacterineae, significant progress on understanding AG biosynthesis has been made recently, in particular with the discovery of new glycosyltransferases (Alderwick et al., 2006b; Seidel et al., 2007). As AG is made up of D-arabinofuranosyl and

Table 4. Conserved motifs in the NCgl0186 orthologue group and comparison with classical SDR motifs

Secondary structure elements were predicted using PredictProtein (Rost et al., 2004). In the motif, ‘a’ denotes an aromatic residue, ‘h’ a hydrophobic residue and ‘x’ any residue. Alternative amino acids at a motif position are given within brackets.

<table>
<thead>
<tr>
<th>Secondary structure element</th>
<th>Classical SDR motif*</th>
<th>NCgl0186 orthologue group motif</th>
<th>Suggested function*</th>
</tr>
</thead>
<tbody>
<tr>
<td>β1 + α1</td>
<td>TGxxGhaG</td>
<td>G[GAK]xaS[EDQ]IM[G]</td>
<td>Structural role in cofactor-binding region</td>
</tr>
<tr>
<td>β3 + α3</td>
<td>DAx[Dv]</td>
<td>[DEN]xa[dV]</td>
<td>Binding of coenzyme</td>
</tr>
</tbody>
</table>

*Persson et al. (2003).
d-galactofuranosyl units, which are both rare sugars (McNeil et al., 1987), a complete understanding of their biosynthetic pathways could provide suitable targets for new chemotherapeutics. Ara is produced from DPA, the formation of which in mycobacteria has been shown to proceed by an unusual epimerization reaction that occurs when the sugar is linked to a prenyl phosphate (Mikusova et al., 2005). Previous experiments performed with purified recombinant Rv3790 and Rv3791 proteins from M. tuberculosis have shown that DPR could be converted into DPA in an in vitro system when both proteins were present in the reaction mixtures (Mikusova et al., 2005). Despite the fact that Rv3790 and Rv3791 were sufficient to catalyse the epimerization of DPR in vitro, there is no indication that, in a physiological context, other proteins are not also involved in this reaction. To investigate the in vivo functions of these proteins in the biosynthesis of AG, we deliberately chose to inactivate their orthologues NCgl0186 and NCgl0187 in C. glutamicum. This strategy was based on the fact that, unlike mycobacteria, corynebacteria can survive with a truncated AG totally devoid of the arabinan domain (Alderwick et al., 2005), and because of the synteny observed for the 3790/3791 locus from genome analysis of numerous Corynebacterineae, suggesting conserved functions for the orthologous proteins.

While the deletion of NCgl0186 was very easy to achieve, all our attempts to delete NCgl0187 were unsuccessful, probably reflecting the essentiality of this gene. The earlier generation of a ubiA mutant in C. glutamicum and its analysis has shown that DPA is the only Ara donor for AG biosynthesis and, as a consequence, its total absence leads to the production of a galactan instead of an AG (Alderwick et al., 2005, 2006a). Cg-Δ186 showed a different pattern: the strain possesses an AG with the same structural motifs as the wild-type strain but contains a significantly reduced quantity of arabinosyl residues. The fact that arabinan was reduced but not totally absent demonstrated a role for NCgl0186 in DPA synthesis, but either in a way different from that observed for Rv3791 in vitro or with functional redundancy in C. glutamicum. The first hypothesis is unlikely for two reasons. First, a direct involvement of NCgl0186 in DPR epimerization was verified by in vitro experiments. Second, a normal level of Ara was restored by in vivo Rv3791 expression in Cg-Δ186. An in silico search for paralogues of NCgl0186 revealed only one gene, NCgl1429, which is conserved among all Corynebacterineae except M. leprae and Mycobacterium bovis species. Although the genetic organization around NCgl1429 or its orthologues differs from one species to another, the gene of interest is always in close proximity, and likely in some cases in an operon together with genes encoding proteins predicted to be involved in precorrin biosynthesis. As in the case of NCgl0186, deletion of NCgl1429 was very easy to realize in the wild-type strain but, in contrast to Cg-Δ186, the resulting mutant synthesized an AG with an Ara:Gal ratio identical to that of the wild-type strain. However, the hypothesis that NCgl1429 is involved in DPA synthesis is strongly supported by the following observations. (i) NCgl1429 could be inactivated in the presence of a copy of NCgl0186 but not in its absence, indicating that NCgl1429 becomes essential when NCgl0186 is absent. (ii) A total recovery of cell wall mycolates was obtained when NCgl1429 was overexpressed in the absence of NCgl0186, indicating that NCgl1429 could fully replace NCgl0186 in the epimerization reaction under these conditions. We thus propose that epimerization of DPR into DPA involves three enzymes that catalyse two distinct steps, each being essential for the viability of the cell. One step would be catalysed by NCgl0187 and another by NCgl0186; in the absence of the latter enzyme, the reaction would be catalysed by NCgl1429. Although it has been shown previously that epimerization results from DPR oxidation followed by reduction to DPA (Mikusova et al., 2005), we were not able to dissect the two steps by identifying which enzyme is responsible of the oxidation reaction and which catalyses the reduction reaction.

By deleting NCgl0186 we obtained, for what is believed to be the first time, a mutant in which the DPA synthesis is lowered, allowing us to determine the influence of DPA level on AG biosynthesis in vivo. Considering the significant diminution of Rha and corynomycolates observed for the Cg-Δ186 mutant and the occurrence of similar structural motifs in the mutant and the wild-type in our structural analyses of AG, it seems that a decrease in DPA synthesis leads to a decrease in the number of branching arabinan chains rather than in the length of these chains. These results suggest that changes in the level of DPA could influence the number of priming sites for Araf deposition to the galactan core but not (or to a lesser extent) Ara polymerization.

The essentiality of the epimerization step was unexpected as the inactivation of the gene encoding the first enzyme involved in DPA synthesis, the DPPR synthase (UbiA), in C. glutamicum, produced a viable mutant (Alderwick et al., 2005). One possible reason is that absence of epimerization leads to a continued synthesis of DPR, which in turn significantly affects the pool of decaprenyl phosphate, so that this becomes limiting for peptidoglycan synthesis and lethal for the cell. This hypothesis has to be proven experimentally.

Apparent redundancy of gene function appears to be common among genes involved in a wide range of physiological processes. This redundancy has been found for some enzymes involved in cell wall biosynthesis in Corynebacterineae, such as mycoloyltransferases in corynebacteria (Brand et al., 2003; De Sousa-D’Auria et al., 2003) and mycobacteria (Puech et al., 2002), and EmbIs in mycobacteria (Escuyer et al., 2001). It could also be the case for the enzyme that dephosphorylates DPPR to DPR. This phosphatase is believed to be encoded by the gene located directly upstream of ubiA (Rv3807c in M. tuberculosis and NCgl2782 in C. glutamicum). Preliminary
experiments with an NCgl2782 mutant of C. glutamicum indicate that if this gene is effectively the phosphatase, a redundant enzyme must exist, as in the case of NCgl0186. In agreement with this, in silico analysis showed a paralogue (NCgl2785) located very close to NCgl2782. Further experiments will be needed to determine the precise role of the enzymic redundancies observed in the synthesis of DPA and more generally in cell wall compound biosynthesis.

ACKNOWLEDGEMENTS

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REFERENCES


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