Glucosyltransferase A (GtfA) and inulosucrase (Inu) of *Lactobacillus reuteri* TMW1.106 contribute to cell aggregation, *in vitro* biofilm formation, and colonization of the mouse gastrointestinal tract

Jens Walter,1,3 Clarissa Schwab,2 Diane M. Loach,1 Michael G. Gänzle2 and Gerald W. Tannock1

1Department of Microbiology and Immunology, University of Otago, Dunedin, New Zealand
2Department of Agricultural Food and Nutritional Science, University of Alberta, Edmonton, Canada
3Department of Food Science and Technology, University of Nebraska, Lincoln, NE 68583-0919, USA

Members of the genus *Lactobacillus* are common inhabitants of the proximal gastrointestinal tract of animals such as mice, rats, chickens and pigs, where they form epithelial biofilms. Little is known about the traits that facilitate biofilm formation and gut colonization. This study investigated the ecological role of a glucosyltransferase (GtfA) and inulosucrase (Inu) of *Lactobacillus reuteri* TMW1.106 and a fructosyltransferase (FtfA) of *L. reuteri* LTH5448. *In vitro* experiments using isogenic mutants revealed that GtfA was essential for sucrose-dependent autoaggregation of *L. reuteri* TMW1.106 cells under acidic conditions, while inactivation of Inu slowed the formation of cell aggregates. Experiments using an *in vitro* biofilm assay showed that GtfA and Inu contributed to biofilm formation of *L. reuteri* TMW1.106. Experiments using *ex-Lactobacillus*-free mice revealed that the ecological performance of the *inu* mutant, but not of the *gtfA* or *ftfA* mutant, was reduced in the gastrointestinal tract when in competition with the parental strain. In the absence of competition, the *gtfA* mutant showed delayed colonization of the murine gut relative to the wild-type. In addition, the *gtfA* mutant showed reduced ecological performance in competition experiments with *Lactobacillus johnsonii* #21. From the evidence provided in this study we conclude that GtfA and Inu confer important ecological attributes of *L. reuteri* TMW1.106 and contribute to colonization of the mouse gastrointestinal tract.

**INTRODUCTION**

The gastrointestinal tract of mammals is colonized by a complex collection of micro-organisms (the gut microbiota) that influences biochemical, physiological, immunological and non-specific disease-resistance characteristics of the host (Gordon & Pesti, 1971). Bacteria of the genus *Lactobacillus* are commonly detected as inhabitants of gut ecosystems and predominate in the proximal (gastric) regions of animals such as pigs, chickens, mice and rats (Tannock, 1992; Walter, 2005). The high population levels of lactobacilli in the gut of these animals are facilitated through bacterial adherence to the non-secretory, stratified, squamous epithelia present in the forestomach, pars oesophagea and crop of mice and rats, pigs and chickens, respectively. The epithelial associations formed by lactobacilli show characteristics of bacterial biofilms (Donlan & Costerton, 2002) because the bacteria are firmly attached to a surface (epithelium) and are embedded in a matrix of extracellular polymeric substances (Fuller & Brooker, 1974; Savage et al., 1968). The mechanisms by which lactobacilli form these epithelial associations are not well understood, but preliminary *in vitro* investigations have shown that carbohydrate molecules are likely to be involved, while a large surface protein (Lsp) appears to initiate adherence *in vivo* (Tannock, 1997; Walter et al., 2005).

Extracellular polysaccharides (exopolysaccharides, EPS) are synthesized by a wide variety of bacteria including lactic acid bacteria, and they have been shown to contribute to dental biofilm formation and cell aggregation of streptococci (Burne et al., 1996; Munro et al., 1991; Yamashita et al., 1993). Many strains of lactobacilli produce homopolysaccharides (HoPS) and oligosaccharides (OS) consisting of either glucose residues (glucans and glucooligosaccharides, GOS) or fructose residues (fructans and fructo-oligosaccharides, FOS) (Gänzle & Schwab, 2005; Korakli & Vogel, 2006; van Hijum et al., 2006). These
Compounds are synthesized from sucrose by the single action of extracellular enzymes termed glycosyltransferases, or more specifically glucosyltransferases and fructosyltransferases, respectively. Research on EPS of lactobacilli has focused on their properties as viscosifying, stabilizing, emulsifying, gelling, water-binding and prebiotic agents in the food industry (Bello et al., 2001; Korakli & Vogel, 2006; van Hijum et al., 2006). However, whereas the importance of streptococcal glycosyltransferases for colonization of the oral cavity is clearly established, the ecological importance of these enzymes and their products has not been revealed for lactobacilli (Korakli & Vogel, 2006).

Strains of Lactobacillus reuteri are commonly detected in the gastrointestinal tract of humans, pigs, chickens, mice and rats (Reuter, 2001; Walter, 2005). They often produce glucans and fructans of different linkage types, and some glycosyltransferases responsible for their production have been biochemically characterized (Kralj et al., 2002, 2004; Tieking et al., 2005; van Hijum et al., 2002, 2006). HoPS and OS formation of L. reuteri TMW1.106 and LTH5448 and its regulation under different environmental conditions has been previously investigated in detail (Gänzle & Schwab, 2005; Schwab & Gänzle, 2006; Schwab et al., 2007). L. reuteri TMW1.106 forms large amounts of a high molecular mass glucan and GOS and low amounts of FOS from sucrose and expresses the gtfA and inu genes encoding a glucosyltransferase and an inulosucrase, respectively (Schwab & Gänzle, 2006; Tieking et al., 2003). L. reuteri LTH5448 produces a high molecular mass frucan and FOS and expresses the ftfA gene encoding a levansucrase (Schwab & Gänzle, 2006). Insertional inactivation of these genes eliminated the synthesis of corresponding poly- and oligosaccharides in L. reuteri TMW1.106 and L. reuteri LTH5448, but did not impair the growth and metabolism of maltose and glucose (Schwab et al., 2007).

The prevalence of HoPS and OS production among lactobacilli isolated from gut ecosystems and the importance of these compounds in cell aggregation and biofilm formation of bacteria indigenous to the oral cavity led us to hypothesize that EPS production might constitute an important phenotypic trait of lactobacilli in colonization of the gastrointestinal tract. The availability of well-characterized, isogenic gtfA, inu and ftfA mutants of L. reuteri (Schwab et al., 2007) and the Lactobacillus-free mouse model (Tannock et al., 1988) provided an excellent opportunity to test this hypothesis.

METHODS

Bacterial strains, media and incubation conditions. The bacterial strains used in this study are listed in Table 1. The mutants were derived by insertional mutagenesis of genes encoding the respective glycosyltransferases by site-specific integration of pORI28 into the chromosome (Schwab et al., 2007). Cultures were propagated anaerobically at 37 °C in Lactobacilli MRS medium (Difco) unless otherwise stated. Erythromycin (3 µg ml⁻¹) was added to the growth medium for culture and maintenance of the mutant strains. Given that the plasmid insertion of the mutants showed high stability in the absence of antibiotics (Schwab et al., 2007), functional assays and phenotypic comparisons were performed using media without erythromycin. Bacterial growth in vitro was investigated by monitoring optical densities (OD₆₀₀) as described previously (Tannock et al., 2005).

Aggregation assays. Strains listed in Table 1 were grown for 16 h (0.5% inoculum) in either MRS medium or MRS medium supplemented with 1% (w/v) sucrose. Cells were harvested from 1 ml of culture by centrifugation at 6000 g for 7 min at room temperature, washed once in 1 ml phosphate-buffered saline (PBS) adjusted to either pH 4 or 7, and resuspended in the initial volume of the same solution. Cell suspensions were kept at room temperature and autoaggregation was monitored visually. For determination of coaggregation, two tubes containing 1 ml cell suspension were prepared as described above using PBS (pH 4). Then 100 µl of a 10 × concentrated cell suspension of the second strain was added to one of the tubes. After 1 h at room temperature, the optical densities were determined in both tubes. The degree of coaggregation between the two strains was recorded as the difference in optical densities following addition of cells of the second strain.

Biofilm experiments. L. reuteri TMW1.106, the gtfA mutant and the inu mutant were grown for 12 h in MRS medium. Cells were

### Table 1. Lactobacillus strains used in this study

<table>
<thead>
<tr>
<th>Strain</th>
<th>Relevant characteristics</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>L. reuteri</td>
<td></td>
<td></td>
</tr>
<tr>
<td>TMW1.106</td>
<td>Sourdough isolate; produces large amounts of glucan HoPS and GOS and small amounts of FOS; GtfA⁻ Inu⁺, Em⁺</td>
<td>Schwab &amp; Gänzle (2006)</td>
</tr>
<tr>
<td>TMW1.106 gtfA mutant</td>
<td>TMW1.106 mutated in the gtfA gene; does not produce glucan; gtfA::pORI28, Em⁻</td>
<td>Schwab et al. (2007)</td>
</tr>
<tr>
<td>TMW1.106 inu mutant</td>
<td>TMW1.106 mutated in the inu gene; large amounts of glucan and GOS without FOS production; inu::pORI28, Em⁺</td>
<td>Schwab et al. (2007)</td>
</tr>
<tr>
<td>LTH5448</td>
<td>Sourdough isolate producing fructan HoPS and FOS; FTF⁺, Em⁺</td>
<td>Schwab &amp; Gänzle (2006)</td>
</tr>
<tr>
<td>LTH5448 ftfA mutant</td>
<td>LTH5448 mutated in the ftfA gene; absence of fructan/FOS production; ftfA::pORI28, Em⁺</td>
<td>Schwab et al. (2007)</td>
</tr>
<tr>
<td>100-23</td>
<td>Rodent gut isolate; weak EPS production on sucMRS; Ftf⁻, Em⁺</td>
<td>Wesney &amp; Tannock (1979)</td>
</tr>
<tr>
<td>L. johnsonii #21</td>
<td>Rodent gut isolate; weak EPS production on sucMRS; Em⁺</td>
<td>Bateup et al. (1995)</td>
</tr>
<tr>
<td>100-100</td>
<td>Rodent gut isolate; absence of EPS production on sucMRS; Em⁺</td>
<td>Lundeen &amp; Savage (1990)</td>
</tr>
</tbody>
</table>
harvested from 0.5 ml culture by centrifugation at 6000 g for 7 min at room temperature and resuspended in 1 ml prewarmed (37 °C) half-strength MRS medium containing 0.5 % sucrose (pH 4.5). Disposable flow cells (Stovall Life Science) were connected to a sterile reservoir of the same medium stored at 37 °C. Flow was generated using a Barnant Manostat Multichannel Pump (Carter). The flow rate was adjusted individually for each channel to 12 ml h⁻¹. The flow condition used was at least 50 times faster than the doubling time of *L. reuteri* TMW1.106. Medium flow was stopped, and the flow cell was aseptically inoculated with bacterial cultures using sterile syringes as described by the manufacturer of the flow cell. After 1 h, flow was started again, and biofilm formation was monitored at different time points using a Nikon Optiphot microscope. Images of mature biofilms were acquired using an Olympus SZX12 dissecting microscope.

**Experiments with mice.*** *Lactobacillus*-free mice (males and females) were maintained throughout the experiments in isolators using gnotobiotic technology (Tannock et al., 1988). The *Lactobacillus*-free status was verified on a regular basis. The animals had free access to water and a standard rat diet (NRM Diet 86, Tegel). The feed contained about 1.25 % sucrose from molasses. The animals were 3–6 weeks of age at the start of the experiments. All animal experiments were conducted with approval from the University of Otago Animal Ethics Committee (approval number 104/02).

**Testing the ability of *L. reuteri* TMW1.106 and LTH5448 to colonize *Lactobacillus*-free mice.*** The ability of *L. reuteri* strains TMW1.106 and LTH5448 to colonize the gastrointestinal tract of *Lactobacillus*-free mice was tested by distributing 10 ml of overnight culture (~5 × 10⁸ bacteria) per cage over the feed and fur of the animals (three mice per bacterial strain). Animals were killed 14 days later, and gut samples (foresomach, jejunum, caecum) were collected for bacteriological culture of serial dilutions of organ homogenates on Rogosa SL agar (Difco), as described previously (Walter et al., 2003).

**Ecological competitiveness of the *gftA*, *inu* and *ftfA* mutants.*** Measurements of ecological competitiveness of strains were made by inoculating *Lactobacillus*-free mice (intragastric gavage) with 1:1 mixtures (total dose of 10⁶ lactobacilli) of mutant and respective wild-type strains as described previously (Walter et al., 2005). Quantification of the mutant strain and the total *Lactobacillus* populations in faecal (after 7 days colonization), foresomach and caecal (14 days colonization) samples was achieved by determining counts on Rogosa SL agar plates with or without erythromycin as described previously (Walter et al., 2005). In competition experiments with *L. reuteri* TMW1.106 and *Lactobacillus johnsonii* #21 (both erythromycin-sensitive), differentiation of strains was achieved by plating gut samples (foresomach and caecum) after 7 days on Rogosa SL agar containing 0.05 % bromocresol green (Sigma). Each strain was quantified based on its distinctive colony morphologies. Correct differentiation of TMW1.106 and #21 was confirmed by picking representative colonies to MRS agar supplemented with or without erythromycin. Ten per cent of colonies were identified correctly based on colony morphology on bromocresol green agar.

**Experiments in *Lactobacillus*-free mice with pure cultures of *L. reuteri* TMW1.106, *gftA* mutant and *inu* mutant to study colonization dynamics, in vivo metabolism and biofilm formation.*** *Lactobacillus*-free mice were inoculated by intragastric gavage with pure cultures (dose of 10⁷ lactobacilli) of either TMW1.106, or the *gftA* or *inu* mutant strain. Bacterial populations were quantified in faecal samples 2 days after inoculation by plating dilutions of homogenates on Rogosa SL agar (Difco) with or without erythromycin. The animals were killed 7 days after inoculation, and the size of *Lactobacillus* populations in the caecum was determined.

Comparison of colony counts on media with or without erythromycin indicated that the mutational insert of both mutants was stably maintained throughout the experiment (data not shown).

Stomach contents of seven *Lactobacillus*-free mice and of five mice colonized for 7 days by either *L. reuteri* TMW1.106 or the *gftA* or *inu* mutant were collected, pooled and freeze-dried. Carbohydrates in stomach contents were extracted using double-distilled H₂O at 80 °C for 2 h. Maltose, glucose and lactate were separated using an Aminex HPX 87H column (Bio-Rad) with 5 mM H₂SO₄ as solvent at 0.4 ml min⁻¹ and detected with a refractive index detector as described previously (Schwab et al., 2007).

Bacterial associations formed on the epithelial surface of the murine forestomach were investigated by transmission electron microscopy (TEM) as described previously (Walter et al., 2007). Briefly, stomachs from two mice colonized by either *L. reuteri* TMW1.106, the *inu* mutant or the *gftA* mutant (see above), excised from the animals, were fixed in a 0.1 M cacodylate buffer (pH 7) containing 3 mg ruthenium red ml⁻¹ and 3 % glutaraldehyde for 4 h at room temperature and then at 4 °C for 4–5 days. Forestomach pieces of ~1 mm³ were successively fixed, dried and washed before embedding in resin. Sections of 80 nm thickness were examined by TEM.

**RESULTS**

**Autoaggregation and coaggregation.*** *L. reuteri* TMW1.106 was the only strain shown in Table 1 whose cells autoaggregated. At pH 7, cells aggregated whether sucrose was present in the growth medium or not, and autoaggregation was not affected by inactivation of the *gftA* and *inu* genes (data not shown). At pH 4, *L. reuteri* TMW1.106 cells aggregated when grown with sucrose, and autoaggregation of the *gftA* and the *inu* mutant was adversely affected. Aggregation of the *inu* mutant cells was slower, while the *gftA* mutant did not aggregate (Fig. 1a, b). None of the strains aggregated at pH 4 when grown in the absence of sucrose, but autoaggregation could be initiated for *L. reuteri* TMW1.106 and the *inu* mutant, but not for the *gftA* mutant, by the addition of ≥ 1 mg sucrose ml⁻¹ to the cell suspensions and incubation at 37 °C for 5 h (data not shown). These findings indicated that glucan production mediated by GtfA was involved in autoaggregation of TMW1.106 cells.

To gain information about the aggregative mechanism of *L. reuteri* TMW1.106, the ability of the *gftA* mutant to coaggregate with cells producing glucan at pH 4 was tested. Addition of wild-type and *inu* mutant cells, but not of the *gftA* mutant, resulted in a reduction in optical density of the *gftA* mutant cell suspension (Fig. 1c). Therefore, *gftA* cells could still coaggregate with *L. reuteri* TMW1.106 and the *inu* mutant. The ability of the *gftA* mutant to coaggregate was not dependent on sucrose, but required sucrose in the growth medium of the wild-type strain (data not shown).

Biofilms in the proximal gut are composed of different *Lactobacillus* strains and species. Therefore the effect of mutation of *gftA* and *inu* on coaggregation with other
strains of *L. reuteri* (LTH5448, its ftfA mutant and 100-23) and *L. johnsonii* (strains #21 and 100-100) was tested. All the strains coaggregated with *L. reuteri* TMW1.106 at pH 4, resulting in a reduction in optical density (Fig. 1c). Inactivation of gtfA, but not inu, adversely affected coaggregation with strains of *L. reuteri*. Inactivation of ftfA of *L. reuteri* LTH5448 had no effect. Coaggregation of *L. reuteri* TMW1.106 with strains of *L. johnsonii* (#21 and 100-100) was not affected by inactivation of gtfA.

### Evaluation of *in vitro* biofilm formation of *L. reuteri* TMW1.106, the gtfA mutant and the inu mutant

Biofilm formation was assessed by direct, non-destructive examination of the cells adhering to a glass surface. As shown in Fig. 2(a), wild-type TMW1.106 formed a biofilm that was visible under a light microscope within 18 h. The gtfA and the inu mutant showed impaired biofilm formation. Intense cell aggregation could be observed for the wild-type and the inu mutant. In contrast, cell aggregation of the gtfA mutant was virtually absent at 12 h, and greatly reduced at 18 h. Biofilms became macroscopically visible after around 30 h. When observed with a dissecting microscope, the biofilms formed by the mutants showed reduced density compared to the wild-type (Fig. 2b). Although clearly impaired when compared with the wild-type, the inu mutant performed slightly better than the gtfA mutant in two independent experiments (Fig. 2).

### Ecological performance of the gtfA, inu and ftfA mutants in the murine gut in competition experiments with the respective wild-type strain

*L. reuteri* TMW1.106 and LTH5448 colonized the murine gastrointestinal tract of *Lactobacillus*-free mice (*n*=3) after a single inoculation (data not shown), and formed population sizes comparable to the rodent strain *L. reuteri* 100-23 used in previous studies (Tannock et al., 2003, 2005, 2007). Further animal experiments were carried out to determine whether the *L. reuteri* glycosyltransferase mutant strains exhibit an impaired competitiveness in the murine gut. Competitive colonization experiments using 1:1 mixtures of wild-type and mutant strains as inoculum (*n*=9–11) revealed that the inu mutant competed poorly with *L. reuteri* TMW1.106 (see Fig. 3b). The gtfA and ftfA mutant populations were not different from that of the wild-type TMW1.106 and LTH5448, respectively (Fig. 3a, c).

### Colonization dynamics of *L. reuteri* TMW1.106, gtfA and inu mutants in the gut of mice

As the ftfA mutant of *L. reuteri* LTH5448 was not adversely affected in aggregation or gut colonization, we focused further studies on strain *L. reuteri* TMW1.106 and its gtfA and inu mutants. Groups of *Lactobacillus*-free mice were inoculated with a single strain to determine the population sizes in faecal (2 days) and caecal (7 days) samples, as well as to detect growth substrates, bacterial metabolites and biofilm formation in the stomach (see below). Inactivation of gtfA and inu of *L. reuteri* TMW1.106 did not result in decreased population levels in the caecal samples when the bacteria were given time to establish (data not shown).
However, colonization by the gtfA mutant was delayed because mice \( (n=7) \) colonized with this strain had statistically significant (Student’s \( t \)-test, \( P=0.013 \)) lower numbers of lactobacilli \( (6.91 \log_{10} \text{c.f.u.}) \) in faecal samples 2 days after inoculation compared to mice inoculated with the wild-type \( (7.63 \log_{10} \text{c.f.u.; } n=7) \). Counts of the inu mutant were lower than wild-type counts \( (7.40 \log_{10} \text{c.f.u.; } n=6) \), but the difference was not statistically significant.

**Metabolism of lactobacilli during growth in the stomach of mice**

Lactobacilli inhabiting the forestomach utilized maltose and glucose and produced lactate (Table 2). Sucrose was present in the stomach of mice not colonized with lactobacilli but the peak was too small for quantification (data not shown). Interestingly, stomach contents of mice colonized with the inu mutant contained higher maltose and lower lactate concentrations compared to stomach contents of mice colonized with the wild-type or the gtfA mutant, indicating that the utilization of maltose by the inu mutant was reduced \( \text{in vivo} \). We compared the \( \text{in vitro} \) growth of \( L. \ reuteri \) TMW1.106, the gtfA and the inu mutant in MRS broth containing glucose or maltose as the only carbohydrates, but the mutant strains did not differ from the wild-type with respect to growth rate (data not shown).

**Epithelial association studied by electron microscopy**

TEM of forestomach epithelium after 7 days of colonization revealed that \( L. \ reuteri \) TMW1.106 adhered to the surface of the stratified squamous epithelium of the forestomach (Fig. 4). The organisms formed cell layers and aggregates on the epithelium, and bacterial densities were highest in inversions (infoldings) of the epithelium and on epithelial cells that had been shed into the lumen. The gtfA mutant and the inu mutant formed epithelial associations that were qualitatively similar to the wild-type (Fig. 4b, c).

**Competition experiments with \( L. \) Johnsonii**

Given the possibility that glucan produced by wild-type cells might have complemented the gtfA mutant in competition experiments with the mice, we tested if GtfA contributes to the ecological performance of \( L. \ reuteri \) TMW1.106 when colonizing the gut in competition with an organism that does not produce glucan. Competitive colonization assays (six mice) with \( L. \) Johnsonii \#21 revealed that \( L. \ reuteri \) TMW1.106 formed an average of 39.7% (SD 27.4) and 36% (SD 6.0) of the total population in the forestomach and caecum, respectively. The proportion of the total population formed by the gtfA mutant was...
9.8 % (SD 6.9) and 22.2 % (SD 20.2) respectively (n=6). The difference in population levels between L. reuteri TMW1.106 and the gtfA mutant was statistically significant in the forestomach (Student’s t-test, \( P < 0.027 \)).

DISCUSSION

Mechanisms involved in attachment and coaggregation that may be relevant to biofilm formation remain largely unexplored for commensal gut bacteria (Flint et al., 2007). Here we present evidence that gtfA and inu contribute to cell aggregation and in vitro biofilm formation of L. reuteri TMW1.106. Furthermore, by using the well-characterized Lactobacillus-free mouse model, we could show that both genes contribute to the ecological performance of the organism during gut colonization. In contrast, inactivation of ftfA did not affect coaggregation or ecological performance of L. reuteri LTH5448 in the experiments presented here.

According to Rickard et al. (2003), bacterial aggregation is an integral process of biofilm formation which proceeds in the form of a succession of adhesion and multiplication events. Autoaggregation of L. reuteri TMW1.106 at neutral pH was independent of sucrose, and inactivation of gtfA and inu had no effect. In contrast, under acidic conditions (pH 4), autoaggregation required both sucrose and GtfA. These findings indicate that the glucan produced by GtfA from sucrose mediates aggregation of L. reuteri TMW1.106. Glucan production has been shown to play an important role in cell aggregation in Gram-positive bacteria (Banas & Vickerman, 2003; Gibbons & Fitzgerald, 1969; Lynch et al., 2007). A possible explanation for the impaired aggregation of the inu mutant is that Inu functions as a glucan-binding protein in L. reuteri TMW1.106. Accordingly, homologues of Inu function as glucan-binding proteins in Streptococcus mutans (Rozen et al., 2004; Russell et al., 1983). The gtfA mutant could still coaggregate with wild-type cells that had been grown in medium containing sucrose, showing that the ability to bind glucan was not mediated by GtfA. Coaggregation with non-aggregating strains of L. reuteri (100-23, LTH5448) was adversely affected (but not eliminated) by inactivation of gtfA. These findings indicated that some L. reuteri strains possess glucan-binding proteins that contribute to coaggregation, even though they do not produce glucan.

We tested the ability of L. reuteri TMW1.106, the gtfA mutant and the inu mutant to form biofilms under acidic conditions (pH 4.5) on a glass surface. Biofilm formation was adversely affected for both mutants. Microscopic evaluation revealed that cell aggregation on the glass slides was basically absent for the gtfA mutant after 12 h, while the inu mutant showed clear signs of cell aggregation. These findings are consistent with those obtained in the aggregation tests. We conclude that GtfA and Inu contribute to biofilm formation by allowing individual cells to form cell aggregates.

Table 2. Concentrations (mmol per g dried stomach content) of fermentable sugars and lactate in pooled stomach contents of Lactobacillus-free (n=8) and ex-Lactobacillus-free (n=5) mice colonized with L. reuteri TMW1.106, the gtfA and the inu mutant

<table>
<thead>
<tr>
<th></th>
<th>L. reuteri TMW1.106</th>
<th>gtfA mutant</th>
<th>inu mutant</th>
<th>No lactobacilli</th>
</tr>
</thead>
<tbody>
<tr>
<td>Maltose</td>
<td>109</td>
<td>73</td>
<td>194</td>
<td>759</td>
</tr>
<tr>
<td>Glucose</td>
<td>75</td>
<td>49</td>
<td>101</td>
<td>253</td>
</tr>
<tr>
<td>Lactate</td>
<td>168</td>
<td>216</td>
<td>83</td>
<td>34</td>
</tr>
</tbody>
</table>
Due to production of lactic acid by the bacteria and host acid secretion, the milieu encountered by lactobacilli during colonization of the forestomach is likely to be acidic (Baumgartner & Montrose, 2004). The lactate levels in the forestomach of mice (Table 2) correspond to those that are typical for stationary cultures of lactobacilli on cereal substrates, which show a pH of around 4 (Gänzle et al., 1998). The importance of GtfA and Inu for cell aggregation and biofilm formation under acidic conditions therefore provides a potential explanation for the decreased ecological performance of the two mutant strains when colonizing the gut of Lactobacillus-free mice. As shown in Fig. 4, L. reuteri TMW1.106 forms epithelial associations that are characterized by the formation of cell aggregates while colonizing the forestomach. These formations resemble natural biofilms detected on forestomach (rodents), crop (chicken), and pars oesophagea (pigs) epithelia, which are dominated by lactobacilli (Fuller & Brooker, 1974; Fuller et al., 1978; Savage et al., 1968). Although the gtfA and inu mutants were still able to form epithelial associations that were qualitatively similar to the wild-type, our in vitro data imply that GtfA and Inu enhance the ability to form these cell aggregates on the forestomach epithelium.

A striking finding of this study was that the disruption of inu impaired colonization of mice by L. reuteri TMW1.106 in competition experiments using the wild-type strain, while disruption of gtfA had no adverse effect. On the other hand, GtfA did contribute to ecological performance when TMW1.106 colonized the gut alone or in competition with L. johnsonii #21. Thus both enzymes contribute to the colonization phenotype of L. reuteri TMW1.106 but they appear to play different roles. A possible explanation is that Inu is a glucan-binding protein and a receptor for the glucan produced by GtfA. Inactivation of Inu would then impair binding to the glucan matrix of the biofilm, resulting in decreased ecological performance in competition experiments with the wild-type. In contrast, assuming that GtfA is only involved in glucan synthesis but not in binding, the gtfA mutant would remain competitive when extracellular glucan is provided by the wild-type strain.

Fig. 4. Characterization of epithelial associations of L. reuteri TMW1.106 (a), the gtfA mutant (b) and the inu mutant (c) on the forestomach epithelium of ex-Lactobacillus-free mice 7 days after inoculation, assessed by TEM.
Determination of the exact role of GtfA and Inu in cell aggregation and biofilm formation requires further experimentation.

Although we clearly show the importance of GtfA and Inu for cell aggregation and biofilm formation, we cannot exclude other functions for these enzymes in the murine gut. Previous work showed that glycosyltransferases of L. reuteri could have several functions under different environmental conditions (Gänzle & Schwab, 2005; Schwab et al., 2007). For example, the gtfA and inu mutants showed a lower resistance to lactic acid when compared to wild-type TMW1.106 (Schwab et al., 2007). This could account for the reduced metabolic turnover of the inu mutant in the murine stomach (Table 2). However, growth rates of both mutants were not impaired in media containing maltose and glucose (Schwab et al., 2007), which are the main growth substrates available in the gut, and the preferred carbon sources for L. reuteri.

In conclusion, our study has shown the importance of two EPS-producing enzymes, GtfA and Inu, for the ecological performance of a gut commensal. Moreover, this is to our knowledge the first report on the importance of glycosyltransferases for cell aggregation and biofilm formation in the genus Lactobacillus. A high proportion of Lactobacillus isolates from gut environments produce EPS and possess orthologues of GtfA and Inu (Gänzle & Schwab, 2005; Korakli & Vogel, 2006; Tieking et al., 2003, 2005; van Hijum et al., 2006). Taken together, this study and the literature data indicate that GtfA and Inu confer important ecological attributes on L. reuteri TMW1.106 and contribute to colonization of the gastrointestinal tract.

ACKNOWLEDGEMENTS

We thank Richard Easingwood, Liz Girvan and the Otago Centre for Electron Microscopy for excellent support in electron microscopy. We are grateful to Greg Somerville and Yefei Zhu (University of Nebraska, Lincoln, USA) for introduction to the biofilm assay. This work was supported by grants of the DFG (Deutsche Forschungsgemeinschaft) and the University of Otago, Dunedin, New Zealand. Clarissa Schwab’s research period at the University of Otago was supported by a DAAD (Deutscher Akademischer Austauschdienst) scholarship. M. G. G. acknowledges the Canada Research Chairs Program for financial support.

REFERENCES


Edited by: P. W. O’Toole