A case of *Exophiala dermatitidis* infection in a child after allogeneic stem cell transplantation: case report and literature review of paediatric cases

Dominika Tanuskova,1,* Julia Horakova,1 Darina Buzassyova,2 Miroslava Poczova,3 Ivana Bodova,1 Peter Svec,1 Alica Chocholova,1 Jaroslava Adamicakova,1 Tomas Sykora,1 Miroslava Pozdechova,1 Lucia Geczova1 and Alexandra Kolenova1

**Abstract**

**Introduction.** *Exophiala dermatitidis* is a relatively common environmental black yeast with worldwide distribution and is a rare cause of fungal infection, mostly in patients with certain predisposing factors. Due to the rarity of the infection, little is known about the specific predisposing factors, way of infection or treatment.

**Case presentation.** Here, we report what is to our knowledge the first case of *E. dermatitidis* infection in a child after allogeneic stem cell transplantation. We also review all paediatric cases reported in the literature since 1993.

**Conclusion.** This is, to our knowledge, the first reported case of *E. dermatitidis* infection in a child after allogeneic stem cell transplantation. This report should increase the awareness of *E. dermatitidis* in immunocompromised paediatric patients, particularly after stem cell transplantation.

**INTRODUCTION**

*Exophiala dermatitidis* is a yeast that can cause a skin, subcutaneous or systemic infection and is a well-known cause of lung infection in patients with cystic fibrosis [1–4]. It is a very rare infection in children. To our knowledge, this is the first case of *E. dermatitidis* infection in a child after haematopoietic stem cell transplantation.

**CASE REPORT**

An 8-year-old boy with secondary acute myeloid leukemia in complete remission after therapy for relapsed medulloblastoma was admitted to Bone Marrow Transplantation Unit in Bratislava for a planned allogeneic stem cell transplantation. The bone marrow transplantation from a 9/10 HLA (human leukocyte antigen) -matched unrelated donor was performed [day (D) 0] after conditioning consisting of treosulfan, cyclophosphamide and melphalan. Anti-thymocyte globulin and cyclosporin A were used for graft-versus-host disease prophylaxis. From D+2, the patient’s clinical and laboratory status started rapidly worsening. The patient was subfebrile up to 37.2 °C, tachy dyspnoeic with insufficient diuresis (oliguria) and fluid retention. The patient was pancytopenic with: high inflammation activity – 158 mg C-reactive protein l⁻¹, 1.02 µg procalcitonin l⁻¹; nephropathy – 9.9 mmol urea l⁻¹, 98 µmol creatinine l⁻¹; metabolic acidosis – pH 7.23, pCO₂3.9 kPa, HCO₃ 11.9 mmol l⁻¹. Due to respiratory and renal failure, the patient was transferred on D+3 to a paediatric intensive care unit. Despite the clinical and laboratory signs of sepsis, the aetiology (the pathogen) was not recognized with repeatedly negative/sterile blood cultures. On D+7, the patient required mechanical ventilation, and from D+9 continuous peritoneal dialysis. On D+9, while positioning him, an asystole occurred and cardiopulmonary resuscitation was required. The cardiopulmonary resuscitation was successful, but from then onwards continuous adrenalin was needed. On D+13, due to progression of pleural effusion, pleural puncture with a pigtail catheter insertion was performed. Repeatedly tested (cultivation, *Pneumocystis jirovecii* PCR, galactomannan) pleural and peritoneal effusions were negative, as were all routinely monitored swabs, virus PCRs and galactomannan from blood. The patient was treated with wide-spectrum empirical antimicrobial therapy, since no specific pathogen was...
recognized. After a short period of clinical stabilization, the patient’s clinical and laboratory status started worsening again. He was haemodynamically unstable with progression of multiorgan failure. From D+16, signs of bullous epidermolysis occurred, the skin became dark, even black, on the scrotum, and grey sputum appeared from bronchi aspiration. On D+20, a yeast, *E. dermatitidis*, was identified from several tracheobronchial aspirations and galactomannan was positive from the aspirations – 2.23 (positivity index). The same yeast was then identified from a pleural effusion from D+18, and throat and nose swabs on D+21. The yeast was susceptible to posaconazole, voriconazole and amphotericin B (Table 1). The patient had antimycotic prophylaxis – micafungin from D–13, at a dose of 1 mg kg\(^{-1}\), from D +3 the dose was increase to a therapeutic 2 mg kg\(^{-1}\), because his condition worsened on empiric broad spectrum antibiotics without any proof of the infection pathogen. Due to kidney failure requiring peritoneal dialysis, treatment with intravenous posaconazole 1×150 mg (the patient was 22 kg at that time) was started. Despite the targeted therapy, the patient died of multiorgan failure on D+25. To the day of his death he did not engraft. Autopsy was not performed. Surprisingly, *E. dermatitidis* was not cultivated from the black lesions of the patient’s skin.

**DISCUSSION**

*E. dermatitidis*, previously known as *Wangiella dermatitidis*, belongs to the group of the so-called black yeasts, which is a polyphyletic morphological group within the Ascomycetes that is characterized by melanized cells and yeast-like growth states (multilateral and polar budding cells) in addition to hyphal growth [5]. *Exophiala* spp. are ubiquitous in nature, but are also found in foodstuffs, steam-bath facilities and bathrooms [6–9]. They are increasingly being recognized as a cause of human disease resulting in skin, subcutaneous tissue and systemic infections [1–3]. Compared with other *Exophiala* species, *E. dermatitidis*, in particular, appears to be associated frequently with systemic infection, as well as with poorer outcomes [10, 11]. It is well known that *E. dermatitidis* frequently colonizes the airways of patients with cystic fibrosis, can trigger antibody production and may cause significant airway infection in these patients [4]. Matsumoto et al. summarized 37 cases of infection published between 1960 and 1992; Suzuki et al. summarized 30 cases published between 1993 and 2011, where 80% of patients had invasive (non-superficial) infections and most of them had predisposing factors (e.g. peritoneal dialysis, leukemia, steroid use, human immunodeficiency virus (HIV) infection, cancer, bronchiectasis and diabetes mellitus) [3, 12]. Chalkias et al. in 2014 were the first people who published a case of *E. dermatitidis* infection in an adult patient after allogeneic stem cell transplantation in a setting of severe graft-versus-host-disease [13]. To our knowledge, there has been no case of this infection in a child after allogeneic stem cell transplantation. We have summarized all published paediatric cases of *E. dermatitidis* infection since 1993, including our case, in Table 2. Only nine cases of the infection have been reported in the paediatric population since 1993. Interestingly, only a minority of paediatric patients had predisposing factors in comparison with adults. Our patient presented shortly after allogeneic stem cell transplantation with myeloablative regimen, he did not engraft; therefore, he was pancytopenic and already in a sepsis of unknown origin. The risk of any fungal infection was high. Other predisposing factors are underlying diseases such as cystic fibrosis, acute leukemia and HIV infection.

Another interesting fact is the neurotropism of *E. dermatitidis* in China – both children reported from China were immunocompetent and had brain abscess, which none of other children had. Involvement of the central nervous system is associated with poor prognosis and both children died. This neurotropism in Asia had been described before, but the explanation remains difficult to understand [14]. Matos et al. investigated the molecular diversity of *E. dermatitidis*. On the basis of internal transcribed spacer sequence, fingerprint and small subunit intron data, strains of *E. dermatitidis* were subdivided into groups A and B. All strains from Asia were of the A genotype. Members of group B have been found in Europe and America, but not in Asia [15].

Few treatments were ultimately found to be effective, and the optimal antifungal therapy for these infections is unknown [16]. Data on *in vitro* susceptibility are sparse and seem to poorly correlate with *in vivo* drug efficacy [17]. As it is difficult to treat the disease after the disease has become fulminated, measures should be taken to initiate appropriate therapy as soon as possible [18]. In our case, despite immediate targeted treatment according to the MIC with intravenous posaconazole, the patient died after 5 days of treatment.

The *E. dermatitidis* infection occurred when the patient had already been in a critical state in the paediatric intensive care unit on mechanical ventilation for 17 days. Therefore, it is impossible to know the degree of contribution of the *E. dermatitidis* infection to the death of the patient. However, his state worsened after he acquired *E. dermatitidis* infection and he died shortly after.

---

### Table 1. MICs of the *E. dermatitidis* isolate as reported by the Department of Mycology, HPL Ltd, Bratislava, Slovakia

<table>
<thead>
<tr>
<th>Antifungal agent</th>
<th>Susceptibility, MIC (μg ml(^{-1}))</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fluconazole</td>
<td>Semi- susceptible, 3.0</td>
</tr>
<tr>
<td>Itraconazole</td>
<td>Susceptible, 0.125</td>
</tr>
<tr>
<td>Voriconazole</td>
<td>Susceptible, 0.064</td>
</tr>
<tr>
<td>Posaconazole</td>
<td>Susceptible, 0.094</td>
</tr>
<tr>
<td>Amphotericin</td>
<td>Susceptible, 0.094</td>
</tr>
<tr>
<td>Caspofungin</td>
<td>Resistant, 32</td>
</tr>
<tr>
<td>Anidulafungin</td>
<td>Resistant, 32</td>
</tr>
<tr>
<td>Micafungin</td>
<td>Resistant, 32</td>
</tr>
</tbody>
</table>
The prognosis of any rare fungal infection is always very serious and this type of infection is often a cause of high mortality. 4 out of 9 children with *E. dermatitidis* infection died, which is higher than in adults in the review by Suzuki et al., where 4 out of 17 patients died [12].

In conclusion, to our knowledge, this is the first reported case of *E. dermatitidis* infection in a child after allogeneic stem cell transplantation. This report should increase the awareness of *E. dermatitidis* in immunocompromised pediatric patients, particularly after stem cell transplantation.

---

**Table 2. Summary of cases of *E. dermatitidis* infection in children without cystic fibrosis since 1993**

<table>
<thead>
<tr>
<th>No.</th>
<th>Age (years)/gender</th>
<th>Manifestation</th>
<th>Predisposing factor</th>
<th>Diagnostic method</th>
<th>Treatment</th>
<th>Outcome</th>
<th>Geographical region</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>3/M</td>
<td>Fungaemia</td>
<td>Acute leukemia</td>
<td>Culture</td>
<td>Catheter removal, AMPH-B, 5-FC</td>
<td>Survived</td>
<td>Germany</td>
<td>[19]</td>
</tr>
<tr>
<td>2</td>
<td>3/M</td>
<td>Fungaemia</td>
<td>HIV infection</td>
<td>Culture</td>
<td>Catheter removal, AMPH-B, ITCZ</td>
<td>Survived</td>
<td>USA</td>
<td>[20]</td>
</tr>
<tr>
<td>3</td>
<td>8/M</td>
<td>Systemic</td>
<td>None</td>
<td>Biopsy</td>
<td>AMPH-B, VRCZ</td>
<td>Died</td>
<td>Turkey</td>
<td>[21]</td>
</tr>
<tr>
<td>4</td>
<td>3/M</td>
<td>Brain abscess, meningitis</td>
<td>None</td>
<td>Biopsy, culture, PCR</td>
<td>AMPH-B, FLCZ, ITLC</td>
<td>Died</td>
<td>China</td>
<td>[14]</td>
</tr>
<tr>
<td>5</td>
<td>11/F</td>
<td>Liver cirrhosis</td>
<td>None</td>
<td>Biopsy, culture, PCR</td>
<td>VRCZ, liver transplantation</td>
<td>Survived</td>
<td>Korea</td>
<td>[22]</td>
</tr>
<tr>
<td>6</td>
<td>17/M</td>
<td>Phaeohyphomycosis</td>
<td>None</td>
<td>Biopsy</td>
<td>ITCZ, Op</td>
<td>Survived</td>
<td>Argentina</td>
<td>[23]</td>
</tr>
<tr>
<td>7</td>
<td>8/M</td>
<td>Brain abscess</td>
<td>None</td>
<td>Biopsy</td>
<td>AMPH-B, VRCZ, 5-FC</td>
<td>Died</td>
<td>China</td>
<td>[24]</td>
</tr>
<tr>
<td>8</td>
<td>16/F</td>
<td>Pneumonia</td>
<td>Cystic fibrosis</td>
<td>Culture</td>
<td>ITCZ, VRCZ</td>
<td>ns</td>
<td>USA</td>
<td>[25]</td>
</tr>
<tr>
<td>9</td>
<td>8/M</td>
<td>Pneumonia</td>
<td>AML after BMT</td>
<td>Culture</td>
<td>PSCZ</td>
<td>Died</td>
<td>Slovakia</td>
<td>Our case</td>
</tr>
</tbody>
</table>

5-FC, 5-Fluorocytosine; AML, acute myeloid leukemia; AMPH-B, amphotericin B; BMT, bone marrow transplantation; F, female; FLCZ, fluconazole; ITCZ, itraconazole; M, male; ns, not stated; Op, operation; PSCZ, posaconazole; VRCZ, voriconazole.

---

**Funding information**
The authors received no specific grant from any funding agency.

**Conflicts of interest**
The authors declare that there are no conflicts of interest.

**Ethical statement**
The parents of the subject gave informed consent to the work.

**References**


---

Five reasons to publish your next article with a Microbiology Society journal

1. The Microbiology Society is a not-for-profit organization.
2. We offer fast and rigorous peer review – average time to first decision is 4–6 weeks.
3. Our journals have a global readership with subscriptions held in research institutions around the world.
4. 80% of our authors rate our submission process as ‘excellent’ or ‘very good’.
5. Your article will be published on an interactive journal platform with advanced metrics.

Find out more and submit your article at microbiologyresearch.org.