Exposure of sputum to phenol disinfectant in conjunction with universal sample processing solution provides safety to laboratory workers during smear microscopy

Sputum smear microscopy is the backbone of any tuberculosis-control programme. Of paramount importance is the safety of laboratory workers, especially in programme settings, who are involved daily in handling a large number of Mycobacterium tuberculosis-infected samples. Increased incidence of tuberculosis amongst health-care workers, especially laboratory technicians, has been well documented (Baussano et al., 2011). In developing countries, infectious samples are often handled on open benches or in hoods without biosafety certification. Moreover, sample concentration techniques are often used to increase the sensitivity of smear microscopy. Increased incidence of tuberculosis amongst health-care workers during smear microscopy, would be highly welcome to the safety of laboratory workers, especially laboratory technicians, has been well documented (Baussano et al., 2011). In developing countries, infectious samples are often handled on open benches or in hoods without biosafety certification. Moreover, sample concentration techniques are often used to increase the sensitivity of smear microscopy (Steingart et al., 2006). The processing of samples containing live tubercle bacilli is associated with the risk of aerosol-mediated spread of infection and contamination of the laboratory environment. Early sample disinfection during processing, that does not compromise the performance of smear microscopy, would be highly welcome to laboratory personnel not having access to stringent biosafety laboratories. Unfortunately, mycobacteria are extremely resistant to disinfection processes. Importantly, heat fixing of slides has not proved to be effective in deactivating M. tuberculosis, and thus the process may potentially lead to the infection of laboratory workers (Allen, 1981; Chedore et al., 2002). Various disinfectants have been evaluated for their ability to deactivate M. tuberculosis with the aim of rendering samples safe for handling and disposal (Chedore et al., 2002; Best et al., 1988, 1990; Rikimaru et al., 2002; Selvakumar et al., 2002; Hall et al., 2007; Hegna, 1977). In the first phase, various disinfectants selected based on earlier studies (Chedore et al., 2002; Best et al., 1988, 1990; Rikimaru et al., 2002; Selvakumar et al., 2002; Hegna, 1977), were tested on negative sputum samples that were spiked with $1 \times 10^5$–$1 \times 10^6$ c.f.u. M. tuberculosis H37Rv ml$^{-1}$. Briefly, each disinfectant (5% phenol in ethanol, 2.5% phenol ammonium sulphate (PhAS), 7.5% hydrogen peroxide, 2% sodium or calcium hypochlorite), was individually added to 1 ml aliquots of spiked sputum, simultaneously with 2 ml USP solution, to achieve a final concentration of the disinfectant as mentioned above. After exposure of the samples to the disinfectant for 30 min at room temperature, 10 ml sterile water was added and the samples were centrifuged at 3500 g for 15 min at room temperature. The resulting pellet was washed twice with 10 ml sterile water, pelleted again by centrifugation as above and the final pellet was resuspended in 500 µl 0.05% Tween 80. Two drops of this suspension were used to make a smear on a glass slide, and four to five drops were inoculated on Löwenstein–Jensen medium slants and one to two drops in Middlebrook 7H9 medium, which were incubated at 37°C for 8 weeks. Aliquots of unspiked negative sputum samples were also processed similarly as negative controls. The Ziehl–Neelsen (ZN)-stained disinfectant + USP solution smears were compared to direct smears or USP solution smears with respect to their smear grade status and staining characteristics, including background staining and interference as per standard World Health Organization criteria (WHO, 2006).

Smears prepared after sputum treatment with USP solution in the presence of phenol, PhAS or hydrogen peroxide were found to be of an acceptable quality. Viability testing indicated that 5% phenol was effective, as no growth of acid fast bacilli (AFB) was observed in any of the aliquots treated with this disinfectant. Hydrogen peroxide had a

### Table 1. Comparison of sputum smear status

<table>
<thead>
<tr>
<th>ZN-staining smear grade</th>
<th>Direct</th>
<th>USP*</th>
<th>USP + phenol</th>
</tr>
</thead>
<tbody>
<tr>
<td>3+</td>
<td>39</td>
<td>56</td>
<td>51</td>
</tr>
<tr>
<td>2+</td>
<td>6</td>
<td>4</td>
<td>5</td>
</tr>
<tr>
<td>1+</td>
<td>11</td>
<td>5</td>
<td>7</td>
</tr>
<tr>
<td>Scanty</td>
<td>6</td>
<td>4</td>
<td>7</td>
</tr>
<tr>
<td>Negative</td>
<td>8</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>Total</td>
<td>70</td>
<td>70</td>
<td>70</td>
</tr>
<tr>
<td>Background staining debris</td>
<td>Present</td>
<td>Absent</td>
<td>Absent</td>
</tr>
</tbody>
</table>

*ZN-stained smear by USP solution concentration method.
similar sterilizing effect, although one positive liquid culture was obtained. However, the inherent unstable nature of this chemical precludes its routine use for sample treatment. PhAS treatment for 30 min was not found to be effective in sterilizing the mycobacterial load as ~18% of the sputum aliquots treated with this disinfectant showed growth of AFB. Treatment of the spiked sputum samples with sodium and calcium hypochlorite caused precipitates on the smear, and also reduced the grading of the smear as well as decreasing the staining intensity of the AFB. Moreover, the poor efficacy of short-term bleach treatment in killing \textit{M. tuberculosis} present in sputum has been demonstrated recently (Githui \textit{et al.}, 2007). Therefore, these disinfectants were not tested further.

Upon satisfactory completion of the first phase study wherein the sterilizing action of phenol did not compromise the performance of USP solution smear microscopy in spiked sputum samples, the study was extended to sputum specimens obtained from tuberculosis subjects. Seventy smear-positive sputum samples were obtained from New Delhi Tuberculosis Centre, India. Their direct smear grade status was decoded after completion of the study and found to vary from ‘3+’ to ‘scanty’. USP solution was added to sputum and the uniform slurry so obtained was distributed into two equal portions. One portion was processed by the standard USP solution methodology and the second portion was treated with USP solution + phenol as described above. Two drops of the resuspended material were examined by ZN staining and the remaining portion was used for viability testing by plating serial dilutions on Middlebrook 7H11 agar. The viable counts obtained from USP solution versus USP solution + phenol-treated samples were compared.

The comparative smear grades obtained by the two methods are given in Table 1 and representative smears are shown in Fig. 1. All 70 USP solution-treated aliquots showed growth of AFB with loads ranging from $1 \times 10^4$ to $6 \times 10^7$ c.f.u. ml$^{-1}$, while none of the corresponding aliquots treated with USP solution + phenol yielded any growth of AFB at the end of 8 weeks of incubation (data not shown). Importantly, phenol-mediated disinfection did not compromise smear microscopy at all. This study demonstrates that phenol is tuberculocidal even in the presence of USP solution and organic matter in sputum. Standard safety precautions while handling phenol need to be observed (http://www.osha.gov/SLTC/healthguidelines/phenol/recognition.html). We recommend that sputum treatment with USP solution + phenol can be effectively used to deactivate \textit{M. tuberculosis} and provide sensitive smear microscopy. Furthermore, because the USP solution method is compatible with all kinds of pulmonary and extrapulmonary samples (Chakravorty and Tyagi, 2005), its use in combination with phenol is expected to provide safety to laboratory personnel engaged in the handling of all types of infectious material containing pathogenic mycobacteria in basic laboratory settings.

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