We report to our knowledge the first case of human infection with *Ophiostoma piceae*. This *Sporothrix schenckii*-related fungus caused disseminated infection involving the lung and the brain in a patient with lymphoblastic lymphoma. The case emphasizes the significance of molecular techniques for identification of rare fungi in the clinical microbiology laboratory.
staining, which could not be further identified due to autolysis of the specimen. After 48 h incubation at 30 °C, plenty of whitish colonies of approximately 1 mm diameter were cultured on Sabaroud dextrose agar (SDA, bioMérieux; Fig. 2b). Microscopy of the colonies revealed elongated cells as well as mycelial structures (Fig. 2c). The colonies developed aerial mycelial structures and darkened coloration after further incubation at room temperature (Fig. 2d) and microscopy revealed a predominance of septated mycelium (Fig. 2e). Subcultures on potato glucose agar (BD) at room temperature and 30 °C confirmed the growth characteristics observed on SDA (Fig. 2f); however, production of mycelium at 30 °C was reduced on this agar. For a more detailed investigation of temperature tolerance, the strain was point-inoculated on oatmeal agar culture plates at room temperature in duplicate, and subjected to constant temperatures of 27, 30, 33 and 36 °C in darkness, with or without pre-incubation for 2 days at room temperature. Growth expansion was measured every second day over 1 week. Consistent growth was obtained at 27 °C. Without pre-incubation, no growth was observed at 30 °C and above. With pre-incubation, colonies reached 30 mm in 7 days at 27 °C, down to 1.5 mm at 36 °C. Colonies at 27–33 °C were cottony, while those at 36 °C were smooth. The latter colonies showed low-frequency conversion of conidia into chlamydospore-like cells, and hyphae were very thin, but no yeast phase was observed. Growth on Mycosel agar (BD) at 27 °C was not inhibited by cycloheximide. Although the morphology was not really typical, infection by a fungus closely related to *Sporothrix schenckii* was initially suspected. Apart from this fungus, no other fungi or bacteria were cultured from the biopsy. Susceptibility testing of the isolate by E-test (Viva Diagnostics) on RPMI agar (72 h incubation at room temperature) revealed the following MICs: fluconazole 0.19 μg ml⁻¹, posaconazole <0.002 μg ml⁻¹, flucytosine 0.047 μg ml⁻¹, caspofungin 0.25 μg ml⁻¹, amphotericin B <0.002 μg ml⁻¹. For final identification, sequencing of the internal transcribed spacer (ITS) regions 1 and 2, including the 5.8S gene, the rRNA operon, as well as the partial β-tubulin gene, was performed using primers and protocols of De Hoog et al. (2007). The sequences were deposited in GenBank with the accession numbers FJ230885 (ITS sequence) and FJ230886 (β-tubulin gene). Comparisons with previously published sequences from GenBank and phylogenetic analyses (maximum-likelihood, maximum-parsimony and Bayesian) of the data confirmed the identity of the isolate as *Ophiostoma piceae* (Fig. 3). The identification was verified at the Centraalbureau voor Schimmelcultures (CBS) and the strain was deposited in the CBS culture collection (CBS 123358).

Following the guidelines for management of sporotrichosis (Kauffman et al., 2007), antifungal treatment was changed from caspofungin to itraconazole (400 mg daily per os) on day 36, resulting in a significant reduction of the pulmonary lesion but rapid progression of cerebral symptoms. Neurosurgical resection of the abscess was performed on day 46, and in the biopsy specimens fungal structures were observed by microscopy with calcofluor white stain that were morphologically equal to those seen microscopically in the lung biopsy (Fig. 2a). However, the fungus was no longer detectable by culture. A Rickham reservoir was implanted, and, using this access, liposomal amphotericin B (0.01–0.3 mg daily) was instilled into the cerebral lesion, followed by significant clinical improvement. However, treatment of lymphoma was not completed due to the severe fungal infection and 6 months after first complete remission of the lymphoma a systemic relapse with bone marrow involvement was diagnosed. The patient rejected reinduction therapy and died a few weeks later.

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**Fig. 1.** (a) CT scan of the chest on day 2; (b) MRI scan of the brain on day 5.
To our knowledge, this is the first case of invasive fungal infection caused by an *Ophiostoma* species. The genus *Ophiostoma* is the largest of the four teleomorph (sexual) genera of the Ophiostomatales (Ascomycetes). However, for several species in the genus, only anamorph (asexual) states are known. These asexually reproducing species, although phylogenetically placed in the *Ophiostoma*, are classified in one of three anamorph genera: *Sporothrix*, *Hyalorhinocladiella* or *Pesotum* (Zipfel et al., 2006). The majority of the approximately 160 species of *Ophiostoma* (including the anamorph species) are wood-inhabiting saprophytes associated with bark beetle vectors. A few of the species are serious tree pathogens. One subgroup of *Ophiostoma*, the so-called *Ophiostoma stenoceras–S. schenckii* complex (Supplementary Fig. S1 in JMM Online), produces *Sporothrix* anamorphs and contains several soil-inhabiting species, including the human pathogen *S. schenckii* (De Beer et al., 2003a; De Meyer et al., 2008). *O. piceae*, however, belongs to another subgroup of the genus, the *O. piceae* complex, and causes blue-stain in the sapwood of trees and logs (Harrington et al., 2001). The complex also includes the causal agents of the notorious Dutch elm disease, *Ophiostoma ulmi* and *Ophiostoma novo-ulmi*, that wiped out millions of elm trees during the previous century in Eurasia and North America (Brasier, 1990). All the species in the *O. piceae* complex have hitherto been regarded as non-pathogenic to humans. *O. piceae* has a global distribution, non-specific associations with many bark beetle vectors, a wide host range and occurs commonly in areas where freshly felled

**Fig. 2.** (a) Calcofluor white stain of lung biopsy sample (magnification 600-fold). (b) Culture on SDA after 3 days incubation at 30 °C. (c) Gram stain of fungal colonies from SDA after 4 days of incubation at 30 °C (magnification 600-fold). (d) Culture on SDA after 4 days incubation at room temperature. (e) Lactophenol blue preparation from colonies grown on SDA for 4 days at room temperature. (f) Culture on potato dextrose agar after 2 weeks incubation at room temperature.
timber is handled and stored, or on injured or stressed trees in native forests and commercial plantations (De Beer et al., 2003b; Harrington et al., 2001; Kirisits, 2004).

Due to the wide distribution of *O. piceae* in the environment, infection in humans may be acquired by several modes, including direct inoculation through trauma, and/or pulmonary inhalation of the airborne conidia of the *Sporothrix* anamorph. Since biopsies were not done on the skin lesions in our patient, it remains open whether these lesions represented the initial focus of infection. Due to his profession as a gardener living in a rural environment, the patient might have had an increased exposure to the fungus. Since the *Ophiostoma* isolate did grow significantly less well at 36°C than at lower temperatures in the laboratory, the pathogenic potential of the isolate may be lower than that of more heat-tolerant species. However, invasive infections with high mortality in immunocompromised patients by other filamentous fungi that do not grow well or at all at 36°C in the laboratory, like some species of the genera *Chrysosporium*, *Mucor*, *Curvularia* and *Alternaria*, are known (Morrison et al., 1993).

Identification of *O. piceae* in the clinical microbiology laboratory is a challenge. Occurrence of loosely aggregated hyphae (Fig. 2e) has been described for *O. piceae* (Przybyl & De Hoog, 1989) and may help to differentiate this species from *S. schenckii*. In addition, clustered denticles as seen in *S. schenckii* are missing. The *Ophiostoma* strain showed good growth at 27°C but proved to be temperature-sensitive in showing no growth at higher temperatures if pre-incubation at room temperature was omitted. Rudimentary, smooth colonies were obtained after 1 week at 36°C. At this temperature, conidia were still abundant, though supported by fragile hyphae, while hyaline chlamydospore-like cells were also obtained. An unambiguous yeast phase as in *S. schenckii* remained absent. Molecular identification methods, such as ribo-somal gene sequencing, are necessary for accurate identification of this species.

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**Fig. 3.** Neighbour-joining tree obtained by maximum-likelihood (ML) analysis of the partial β-tubulin gene, identifying the patient isolate as *Ophiostoma piceae*. Bootstrap values for ML are indicated above, and those for maximum-parsimony below, branching points. Posterior probabilities >95%, obtained from Bayesian analyses, are indicated by bold lines.
Optimal treatment of patients with *Ophiostoma* infection is unclear, and methods for antifungal susceptibility testing as well as interpretative criteria for test results have not been validated. Due to the close relationship to *S. schenckii* and the comparably low MIC value for itraconazole, high-dose itraconazole therapy for 10 weeks (Gullberg et al., 1987; McGinnis et al., 2001; Van Cutsem, 1992) was chosen, resulting in improvement of the patient and partial regression of the lesions. Nevertheless, progression of the underlying disease led to the fatal outcome in the patient. Since this *Ophiostoma* infection developed under prophylactic treatment with voriconazole (given, however, in a therapeutical dosage of 400 mg daily), it may be discussed whether the first-line and long-term azole therapy was adequate. *In vitro* susceptibility testing did not reveal increased MIC values for itraconazole or voriconazole, and treatment with itraconazole showed some clinical and radiologically documented success regarding the pulmonary lesion. However, for cure of cerebral lesions caused by *Ophiostoma*, amphoterin B or an azole with good penetration into cerebrospinal fluid, like voriconazole, may be required. After intracerebral instillation of amphoterin B, the patient improved significantly. Nevertheless, susceptibility testing of isolates appears to be recommended since the sensitivity of *S. schenckii* to amphoterin B has been reported to be strain-dependent (McGinnis et al., 2001). According to *in vitro* studies, echinocandins have less activity than triazoles against *S. schenckii* and other dimorphic fungi (Espinel-Ingoff, 1998) and, thus, after final identification of the isolate, therapy was changed from caspofungin to an azole. In further cases, posaconazole may also be a promising antymycotic agent since it had the lowest MIC of all azoles.

In conclusion, ubiquitous fungi from the genus *Ophiostoma* may represent a new group of human-pathogenic fungi in immunocompromised patients. This fatal case of *O. piceae* infection emphasizes the significance of invasive diagnostic measures and fungal cultures followed by molecular identification of the isolates in patients with clinical suspicion of fungal infection.

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References


