Enteroaggregative *Escherichia coli*: epidemiology, virulence and detection

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Enteroaggregative *Escherichia coli* (EAEC) is a subgroup of diarrhoeagenic *E. coli* (DEC) that during the past decade has received increasing attention as a cause of watery diarrhoea, which is often persistent. EAEC have been isolated from children and adults worldwide. As well as sporadic cases, outbreaks of EAEC-caused diarrhoea have been described. The definition of EAEC is the ability of the micro-organism to adhere to epithelial cells such as HEp-2 in a very characteristic ‘stacked-brick’ pattern. Although many studies searching for specific virulence factor(s) unique for this category of DEC have been published it is still unknown why the EAEC cause persistent diarrhoea. In addition, the aggregative property of EAEC causes a lot of problems in serotyping due to the cells auto-agglutinating. The gold standard for identification of EAEC includes isolation of the agent and an adherence assay using tissue culture, viz. HEp-2 cells. This assay is in most cases reliable; however, emergence of ‘atypical’ EAEC has been described in several publications. In addition, the HEp-2 assay is time consuming, demands a tissue culture lab and trained staff. Several molecular biological assays have been described, however, none show 100% specificity.

Introduction

*Escherichia coli* is the type species of the genus *Escherichia* that contains mostly motile Gram-negative bacilli that fall within the family Enterobacteriaceae. It is the predominant facultative anaerobe of the human colonic flora. The organism typically colonizes the infant gastrointestinal tract within hours of life, and thereafter *E. coli* and the host derive mutual benefit for decades (Kaper et al., 2004). However, there are several highly adapted *E. coli* clones that have acquired specific virulence factors, which increase their ability to adapt to new niches and allow them to cause a broad spectrum of diseases. Three general clinical syndromes can result from infection with pathogenic *E. coli* strains: enteric/diarrhoeal disease, urinary tract infection and sepsis/meningitis (Nataro & Kaper, 1998). As long as these bacteria do not acquire genetic elements encoding virulence factors, they remain benign commensals. Strains that acquire ‘foreign’ DNA encoding enterotoxins, adhesins or invasion factors become virulent and can cause either a plain, watery diarrhoea or inflammatory dysentery.

Enteroaggregative *E. coli*

Among the *E. coli* causing intestinal diseases, there are six well-described categories: enteroaggregative *E. coli* (EAEC), diffusely adherent *E. coli* (DAEC), enteroinvasive *E. coli* (EIEC), enteropathogenic *E. coli* (EPEC), enterohaemorrhagic *E. coli* (EHEC) and enterotoxigenic *E. coli* (ETEC) (Nataro & Kaper, 1998). These categories have virulence attributes that help bacteria to cause diseases by different mechanisms. The EIEC, EHEC and ETEC are defined as *E. coli* strains possessing specific virulence attributes, including different toxins, invasion plasmid and colonization factors. Almost half a century ago, Ewing et al. (1963) determined that certain serotypes of *E. coli* were associated with outbreaks of diarrhoea. Strains belonging to these serotypes were referred to as EPEC. In 1979, Cravioto and colleagues described an in vitro assay based on the adhesion of the bacteria to HEp-2 cells showing that EPEC bind to the cells in a localized pattern (Cravioto et al., 1979). A few years later it was shown that adherent non-EPEC strains were associated with diarrhoea. These strains were called ‘enteroadherent *E. coli*’ (Cravioto et al., 1991; Mathewson et al., 1985, 1987). At the same time, Nataro et al. (1987) recognized two different phenotypes among the enteroadherent strains, i.e. diffuse and aggregative. The finding of Nataro was the first description of EAEC. The aggregative adherence is characterized by a ‘stacked-brick’ formation of bacterial cells attached to the HEp-2 cells (Nataro & Kaper, 1998; Nataro et al., 1998). The basic strategy of EAEC seems to comprise colonization of the intestinal mucosa, probably predominantly that of the colon, followed by secretion of enterotoxins and cytotoxins (Nataro et al., 1998). Studies on human intestinal specimens indicate that EAEC induces mild, but significant, mucosal damage (Hicks et al., 1996). EAEC strains characteristically enhance mucus secretion from the mucosa, with trapping of the bacteria in a bacterium-mucus biofilm (Nataro & Kaper, 1998). Two further observations support a role of mucus in EAEC pathogenesis: EAEC bind avidly in the rabbit in vitro.
Epidemiology

Developing countries

A growing number of studies have supported the association of EAEC with diarrhoea in developing countries, most prominently in association with persisting diarrhoea (Bhan et al., 1989a, b, c; Fang et al., 1995; Lima et al., 1992). Several studies in children with diarrhoea have shown a significant difference in the prevalence of EAEC compared to the controls (Bhatnagar et al., 1993; Bouzari et al., 1994; Cravioto et al., 1991; Gonzalez et al., 1997; Nataro et al., 1987). EAEC and persistent diarrhoea syndrome have been consistently associated (Fang et al., 1995; Lima et al., 1992; Wanke et al., 1991). The increasing number of such reports and the rising proportion of diarrhoeal cases in which EAEC are implicated suggest that EAEC are important emerging agents of paediatric diarrhoea.

During the initial years after the discovery of EAEC there were a lot of doubts about the pathogenicity of this category of diarrhoeagenic *E. coli* (DEC) (Echeverria et al., 1992; Gomes et al., 1989). However, Nataro et al. (1995) showed that a reference strain of EAEC could cause diarrhoea in a volunteer study. In addition, a number of outbreaks have proven that at least some EAEC strains cause diarrhoea in humans. Furthermore, many case–control and cohort surveys of the past 15 years strongly suggest that EAEC is an important cause of diarrhoea in people of all ages in developing and industrialized countries. In a recent study from Vietnam by Vu Nguyen et al. (2006) it was shown that EAEC is more frequently associated with diarrhoea in children less than 2 years of age. In this study, 587 children of less than 5 years of age with diarrhoea and 249 age-matched healthy controls were examined for, among other pathogens, EAEC. Of all identified EAEC strains in the diarrhoeal group, 87% were isolated from children less than 2 years of age. The corresponding figure for the control group was 39% (Vu Nguyen et al., 2006). Many of the epidemiological surveys that identified EAEC as a diarrhoeal pathogen were done in developing countries. However, EAEC has been found to be associated with diarrhoea in developed countries as well.

Developed countries

In a Scandinavian case–control study the prevalence and the association of EAEC with diarrhoea was greater than for EPEC (Bhatnagar et al., 1993). Another study conducted in east London showed that EAEC could be recovered from children with acute and persistent diarrhoea (Chan et al., 1994). A clear association of EAEC with diarrhoea in children in Germany was shown by Huppertz et al. (1997) who recovered EAEC from 16 (2%) of 798 children with diarrhoea but none from 580 healthy controls. Other European studies in children (Knutton et al., 2001; Presterl et al., 1999) also indicate that EAEC may be a leading cause of diarrhoeal disease in developed as well as developing countries. In a Serbian neonatal ward an outbreak of EAEC diarrhoea was described by Cobeljic et al. (1996) where 19 babies were affected and 3 of these patients got diarrhoea. One EAEC strain resistant to multiple antibiotics was implicated. In the largest reported outbreak so far, 2697 (40.6%) Japanese children who ate infected school lunches had severe diarrhoea and EAEC was found in 10% of cases (Itoh et al., 1997). Several other outbreaks both in children and in adults have been described in the UK (Smith et al., 1997; Spencer et al., 1999), India (Pai et al., 1997) and France (Boudailliez et al., 1997; Morabito et al., 1998).

In a 1 year prospective Swedish study on enteropathogens in adult patients with diarrhoea and healthy control subjects, 105 of 760 patients with diarrhoea were positive for DEC. EAEC was present in 16 cases and was the second most common isolate among DEC surpassed only by ETEC (Svennungsson et al., 2000). Other studies have shown that this pathogen is an important cause of travellers’ diarrhoea (Adachi et al., 2001; Gascon et al., 1998; Schultz et al., 2000). However, documented reports are less common because EAEC is not sought in many studies. Travellers to all developing countries are at risk and if EAEC were sought in all laboratories, infection by this pathogen could explain over 25% of cases for which no pathogen is recovered (Adachi et al., 2001).

Immunocompromised

Diarrhoea is an important cause of morbidity in the immunocompromised. A wide range of pathogens is implicated in AIDS-associated diarrhoea, and in many cases a causative agent is not found. Case reports from Mayer & Wanke (1995) described that EAEC could be recovered from diarrhoeal stools from AIDS patients. There have been other reports after epidemiological surveillance among HIV-positive patients (Mayer & Wanke, 1995).
Virulence factors

In order to cause diarrhoeal disease, EAEC adheres to intestinal mucosa, forms a mucoid biofilm and induces toxic effects on the intestinal mucosa that result in diarrhoea. The exact mechanism of pathogenesis is not fully understood; however, adhesins, toxins and several other factors have been implicated. Certain strains carry a high molecular weight plasmid associated with the aggregative adherence (Law et al., 1998; Vial et al., 1988), on which a number of virulence genes are located. These are (i) heat stable toxin-1 (EAST-1) (Savarino et al., 1991), (ii) aggregative adherence fimbriae I and II (AAF/I and AAF/II) (Nataro et al., 1992; Rich et al., 1999), as well as the AAF/III (Bernier et al., 2002). Also located on the plasmid are (i) transcriptional activator gene (Nataro et al., 1994, 1998), (ii) anti-aggregation protein gene (Sheikh et al., 2002), and (iii) anti-aggregation protein transporter gene (Baudry et al., 1990). In addition, the plasmid-encoded toxin (Eslava et al., 1998) and a cryptic ORF known as shf (Czezczulin et al., 1999) have been described. Other virulence factors that are believed to be associated with EAEC are 18 and 30 kDa outer-membrane adhesins (Chart et al., 1997; Debroy et al., 1995; Grover et al., 2001). In addition, different pathogenicity islands have been identified within the EAEC group, including Shigella she pathogenicity island, containing enterotoxin and mucinase genes (Henderson et al., 1999), and Yersinia high-pathogenicity island, containing the yersiniabactin siderophore gene (Schubert et al., 1998). Strains that carry the high molecular weight plasmid represent an important subgroup (Cerna et al., 2003; Elias et al., 2002) and may be regarded as ‘typical EAEC’. However, EAEC is a heterogeneous group of E. coli (Czezczulin et al., 1999) and certain strains, although adherent to the HEp-2 cells in the stacked-brick mode, lack the high molecular weight plasmid and can be called ‘atypical EAEC’ (Cobeljic et al., 1996; Elias et al., 2002; Gioppo et al., 2000; Itoh et al., 1997).

Identification – serotyping and molecular biology

However, none of the above-described gene(s) is conserved among all of the EAEC and many are not unique to this category of DEC. Recently, Jenkins et al. (2006a) used several different probes to characterize EAEC strains. The results show that the majority of the HEp-2 positive strains were also positive for the anti-aggregation protein transporter gene described by Baudry et al. (1990) against which Schmidt and colleagues developed a PCR assay in 1995 (Schmidt et al., 1995). Overall, 143 EAEC strains were analysed and 128 (90%) were positive for the anti-aggregation protein transporter gene (Jenkins et al., 2006a). However, 10% of the strains verified by HEp-2 assay were negative in the PCR assay. This makes it difficult to provide a genotypic definition for EAEC and to design specific molecular biological assays for the detection of this pathotype. In addition, serotyping of EAEC is a problem. Due to their aggregative phenotype, many of the strains auto-agglutinate and are often described in the literature as non-typable or as O-rough. It is also well established that the EAEC group is highly heterogeneous. Huppertz et al. (1997) analysed EAEC from German children and found that of 14 typable isolates all belonged to different serotypes. In another study in the UK, 97 EAEC strains were serotyped to 40 different O-types. However, the remaining 121 EAEC isolates were non-typable (Food Standards Agency, 2000). In a study by Jenkins et al. (2006a), 93 out of 143 EAEC strains could be serotyped and belonged to as many as 47 different serotypes. This shows that serotyping, although useful in the characterization of other DEC, is of little value in the diagnosis of EAEC. As mentioned earlier, the HEp-2 adherence test is the gold standard for the identification of EAEC. This test requires specialized facilities and can therefore only be conducted in reference laboratories, and strict adherence to protocol is required (Haider et al., 1992; Vial et al., 1990). Several attempts have been made to develop a molecular biological assay for the identification of EAEC. In 1990, Baudry and colleagues developed a DNA probe, CVD432, that was found to be 89% sensitive and 99% specific for EAEC (Baudry et al., 1990). Based on this probe, a PCR assay was developed by Schmidt et al. in 1995. This assay has been widely used since then. In search for additional virulence factors, several other assays have been described, none of which with has as high sensitivity and specificity as the one described in 1995. The problems in identification of EAEC have been clearly highlighted very recently (Jenkins et al., 2006b). The authors set up a multiplex PCR targeting three different genes: (i) the anti-aggregation protein transporter gene described by Baudry et al. (1990), (ii) the EAST gene (Savarino et al., 1991) and (iii) a chromosomal gene present in the pheU pathogenicity island designated aggR-activated island (Jenkins et al., 2006b). The authors used this multiplex PCR to identify EAEC from patients in the community suffering from diarrhoea. They found that twice as many isolates were positive against at least one of the primers as compared to the HEp-2 adherence assay alone.

Conclusion

The HEp-2-adherence assay and/or the CVD432 standard probe represent the best means at present for detecting EAEC. However, it is clear that more sensitive, specific and practical methods are needed in order to improve the diagnosis of infected cases with EAEC and to understand the disease.

Note from the editor-in-chief

A review on EAEC by Huang and colleagues was published in the Journal of Medical Microbiology October 2006 issue (Huang et al., 2006) that concentrated on the epidemiology and pathogenic mechanisms of the bacterium. More recently a paper by Jenkins and colleagues (Jenkins et al., 2006b) has been published drawing attention to the difficulties in diagnosing EAEC infection by molecular methods. This mini review by Andrej Weintraub discusses the problem further.
References


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