Antibacterial resistance in *Helicobacter pylori* strains isolated from Bulgarian children and adult patients over 9 years

Lyudmila Boyanova,1 Rossen Nikolov,2 Elena Lazarova,3 Galina Gergova,1 Nikolai Katsarov,4 Victor Kamburov,5 Zoya Spassova,2 Sirigan Derejian,2 Christo Jelev,3 Ivan Mitov1 and Zacharii Krastev2

Correspondence
Lyudmila Boyanova
lboyanova@hotmail.com or lboyanova@lycos.com

1Department of Microbiology, Medical University of Sofia, Zdrave Street 2, 1431 Sofia, Bulgaria
2Department of Gastroenterology, University Hospital St Ivan Rilski, Sofia, Bulgaria
3Department of Gastroenterology, University Paediatric Hospital, Sofia, Bulgaria
4Second Surgery Department of Alexander Hospital, Sofia, Bulgaria
5Urgent Endoscopy Unit, Emergency Hospital Pirogov, Sofia, Bulgaria

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The aim of this study was to evaluate the primary, combined and post-treatment antibacterial resistance rates in 1205 *Helicobacter pylori* strains from non-treated (786 adults, 282 children) and treated (109 adults, 28 children) patients in Bulgaria. Susceptibility was tested by the limited agar dilution method. Overall primary resistance rates to metronidazole, clarithromycin, amoxicillin, tetracycline and both metronidazole and clarithromycin were respectively 15±0, 12±5, 1±5, 3±4 and 4-7% in children and 25±6, 12±6, 0±8, 5±2 and 4-9% in adults. Primary metronidazole resistance in adults was more common than in children, but the differences for other agents tested were not significant. Primary resistance rates were in the range of those reported worldwide. There was no significant increase in primary resistance rates from 1996/1999 to 2003/2004; however, clarithromycin resistance rates exhibited a slight tendency to increase. Post-treatment resistance to amoxicillin was detected in 1-6% of 63 strains. Post-treatment resistance to metronidazole was common (81±6%) and that to clarithromycin was considerable (36%). Alarming emergence of strains with triple resistance to amoxicillin, metronidazole and clarithromycin was found in two non-treated and three treated patients. The results motivate a larger and continuing surveillance of *H. pylori* resistance in Bulgaria and worldwide.

**INTRODUCTION**

Triple therapy of amoxicillin, clarithromycin and a proton pump inhibitor (PPI) is the treatment of choice for eradicating *Helicobacter pylori* infection (Megraud, 2004). Eradication of the infection leads to clinical cure but is successful in only 65–90% of cases (Cantón et al., 2001; Megraud, 2004). Antibacterial resistance is the major cause for treatment failure (Meyer et al., 2002). The prevalences of primary *H. pylori* resistance to metronidazole and clarithromycin have oscillated around 30–40% and 2–15% in most European countries and those of post-treatment and secondary resistance have attained 66–90% and 11-4–71%, respectively (Megraud, 2004; Peitz et al., 2002).

Resistance rates in *H. pylori* can vary between groups of patients according to the age, sex, disease and place of residence and an evolution of resistance, often reflecting the previous or the national consumption of a given antibacterial agent, has been observed in some countries (Meyer et al., 2002; Megraud, 2004). The aim of this study was to assess the rates of the primary, combined, post-treatment and secondary antibacterial resistance in *H. pylori* clinical isolates from 1205 patients with gastroduodenal diseases over 9 years in Bulgaria.

**METHODS**

Consecutive *H. pylori* strains isolated from non-treated patients (282 children and 786 adults) were evaluated in 1996–2004 and consecutive *H. pylori* strains from treated patients (109 adults and 28 children) were evaluated in 1998–2004. A single antral biopsy specimen was taken from the children and two antral biopsy specimens per patient were taken from the adults. Specimens from the treated patients were taken 4–8 weeks after the end of therapy for *H. pylori* eradication. Most common eradication regimens involved amoxicillin, clarithromycin and PPI (14 patients), amoxicillin, azithromycin and PPI (27), amoxicillin, metronidazole and PPI (26), amoxicillin, metronidazole and bismuth compound (6) and clarithromycin,
metronidazole and PPI (5). No detailed data for the previous treatment of the other patients were available. The PPI used were omeprazole (the most common PPI used), rabeprazole and esomeprazole.

Stomach biopsy specimens were transported in Stuart transport medium (Merck) for <5 h. A smear was prepared from part of the specimen and was used for modified Gram staining with carbol fuchsin. Part of the specimen was placed in 10% urea agar medium and the colour change was read after incubation at 37°C for 30 min and 3 h. The remaining part of the specimen was homogenized in sterile saline with sterile needles and inoculated onto blood agar (Columbia agar base; Becton Dickinson) with 10 μg vancomycin, 5 μg trimethoprim, 5 μg cefsludin and 5 μg amphotericin B ml⁻¹ and/or 10% defibrinated sheep blood, and 1% IsovitaleX (Becton Dickinson). Both selective and non-selective media were used for primary culture. Plates were incubated microaerophilically for 12 days. Identification was made by Gram staining of the colonies, lack of aerobic growth on blood agar plates and testing for urease, oxidase and catalase.

Susceptibility was tested by a limited agar dilution method. Two drops (about 60 μl) of *H. pylori* suspensions, prepared in Mueller–Hinton broth [National Centre of Infectious and Parasitic Diseases (NCIPD)] to obtain McFarland turbidity standard 3–4, were inoculated on Mueller–Hinton blood agar plates (NCIPD) containing 1% IsovitaleX and one of the following drug concentrations: 8, 16 or 32 μg metronidazole ml⁻¹, 0.25, 0.5, 1 or 2 μg clarithromycin ml⁻¹, 0.5, 1 or 2 μg amoxicillin ml⁻¹ and 4 μg tetracycline ml⁻¹. Antimicrobial agents were obtained from Sigma (amoxicillin, metronidazole and tetracycline) and Abbott Laboratories (clarithromycin). The plates were incubated microaerophilically at 37°C for 2–3 days. If *H. pylori* growth appeared on the plate, the isolate was considered as resistant to the corresponding drug concentration. Non-selective medium plates were used for a control of strain viability. The cut-off concentrations for resistance were: >8 μg metronidazole ml⁻¹, >1 μg clarithromycin ml⁻¹, >0.5 μg amoxicillin ml⁻¹ and >4 μg tetracycline ml⁻¹ (Megraud et al., 1999; NCCLS, 2000). Primary resistance was defined as resistance of an *H. pylori* strain isolated from a patient with no previous therapy for *H. pylori* eradication. Post-treatment resistance was determined as resistance to a given antibacterial agent after therapy involving the agent, regardless of the strain sensitivity before treatment. Secondary resistance was defined as resistance acquired during treatment by a strain that was susceptible to the agent before the treatment. Secondary resistance to amoxicillin and macrolides was assessed in 18 and 13 strains, respectively.

Differences between patients with susceptible and resistant strains were assessed with a chi-squared test with or without Yates’ correction. Yates’ correction factor for continuity was added in the calculation of chi-squared values for 2 × 2 tables when the expected frequency was <10 in one or more cells.

**RESULTS**

Overall primary resistance rates to metronidazole, clarithromycin, amoxicillin, tetracycline and both metronidazole and clarithromycin were respectively 15–0, 12–5, 1–5, 3–4 and 4–7% in children and 25–6, 12–6, 0–8, 5–2 and 4–9% in adults. Primary metronidazole resistance in adults was more common than in children (*P<0.001*), but the differences between the age groups for the other antibacterial agents were not significant (*P>0.20*). There were no significant differences between primary resistance rates in adults aged >60 years and those aged 19–59 years (*P>0.20*).

There was no significant increase in antimicrobial resistance rates in children and adults from 1996/1999 to 2003/2004 (*P>0.20*) (Table 1); however, the rates of clarithromycin resistance exhibited a slight tendency to increase in children (from 10–4 to 13–9%; *P>0.20*) and in adults (from 9–8 to 14–3%; *P>0.20*).

**Table 1. Evolution of primary *H. pylori* resistance to antibacterial agents in Bulgaria**

% R is the percentage of resistant strains; 95% CI is the 95% confidence interval for % R.

<table>
<thead>
<tr>
<th>Agent</th>
<th>Years</th>
<th>Children</th>
<th>Adults</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Strains</td>
<td>% R</td>
<td>95% CI</td>
</tr>
<tr>
<td>Amoxicillin</td>
<td>1996–1999</td>
<td>38</td>
<td>7–9</td>
</tr>
<tr>
<td></td>
<td>2000–2002</td>
<td>161</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>2003–2004</td>
<td>73</td>
<td>1–4</td>
</tr>
<tr>
<td>Tetracycline</td>
<td>1996–1999</td>
<td>38</td>
<td>2–6</td>
</tr>
<tr>
<td></td>
<td>2000–2002</td>
<td>135</td>
<td>3–7</td>
</tr>
<tr>
<td></td>
<td>2003–2004</td>
<td>64</td>
<td>3–1</td>
</tr>
<tr>
<td>Metronidazole +</td>
<td>1996–1999</td>
<td>47</td>
<td>4–2</td>
</tr>
<tr>
<td>clarithromycin</td>
<td>2000–2002</td>
<td>160</td>
<td>4–4</td>
</tr>
</tbody>
</table>
The overall resistance rates to metronidazole, clarithromycin, amoxicillin, tetracycline and both metronidazole and clarithromycin in treated patients were 30–4, 18–2, 4–3, 0 and 4–5% in children and 59–2, 31–3, 4–9, 4–3 and 18–5% in adults, respectively. Treated adult patients harboured fourfold more strains (18–5%) resistant to both metronidazole and clarithromycin than did the children (4–5%). Post-treatment resistance rates were: amoxicillin, 1–6%; metronidazole, 81–6%; and clarithromycin, 36–0% (Table 2). Post-treatment resistance was more common than primary resistance to metronidazole ($P < 0.001$) and clarithromycin ($P < 0.001$). Secondary resistance to clarithromycin was found in 3 (23–1%) of 13 cases. Secondary resistance to amoxicillin was not detected in 18 cases evaluated.

**DISCUSSION**

Primary resistance rates of *H. pylori* were in the range of those reported in other countries (Megraud, 2004), with relatively higher resistance rates to amoxicillin and tetracycline and a lower resistance rate to metronidazole in Bulgarian children. Primary metronidazole resistance was more common in adults than in children, probably because of the use of nitroimidazoles to treat gynaecological and dental infections in adults.

*H. pylori* resistance to amoxicillin and tetracycline has been reported only in some countries (Meyer et al., 2002; Megraud, 2004). The detection of two strains with triple resistance to amoxicillin, clarithromycin and metronidazole from a non-treated man (in 2003) and a boy aged 14 years (in 2004) is worrying. This focuses attention on possible multidrug resistance in some *H. pylori* strains and related future therapeutic problems. Combined *H. pylori* resistance to beta-lactams, nitroimidazoles, tetracycline and other agents has been reported recently (Kwon et al., 2003).

Primary *H. pylori* resistance rates to clarithromycin or metronidazole have exhibited a significant increase or only slight differences over time according to different authors (Lopez-Brea et al., 2001; Bontems et al., 2001; Kalach et al., 2001). In the present study, no significant increase in primary antibacterial resistance rates was detected in children and adults from 1996/1999 to 2003/2004; however, the clarithromycin resistance rate exhibited a slight tendency to increase. The results could reflect the relatively low macrolide consumption in Bulgaria [0–34 defined daily doses per 1000 inhabitants per day (DID) in 1999], the moderate nitroimidazole consumption (0–41 DID in 1999) and the rare use of metronidazole in children. The primary tetracycline resistance rate (low but constant since 1999) can be associated with the still frequent tetracycline consumption in our country (4–24 DID in 1999). Amoxicillin resistance was rare but was detected in 1999, 2001, 2003 and 2004.

Amoxicillin resistance (MIC 1–2 μg ml$^{-1}$), combined with metronidazole resistance, was found in four strains from treated patients. Three of the strains also exhibited clarithromycin resistance. Amoxicillin resistance has been associated with a mutation in the *pbp-1A* gene and altered uptake of beta-lactams after long exposure of *H. pylori* to amoxicillin (DeLoney & Schiller, 2000). Japanese authors have reported an increase in *H. pylori* resistance rates to amoxicillin from 2000 to 2003 (Watanabe et al., 2005).

In conclusion, the primary resistance rates of *H. pylori* from Bulgarian patients were in the range of those reported worldwide. There was no significant increase in resistance rates to antimicrobial agents in children and adults from 1996/1999 to 2003/2004, although the rates of clarithromycin resistance exhibited a slight tendency to increase. Since 1999, tetracycline-resistant strains have been detected every year, and amoxicillin resistance has been observed less frequently. The alarming appearance of strains with triple resistance to amoxicillin, metronidazole and clarithromycin was detected in non-treated and treated patients. The post-treatment resistance rate in *H. pylori* to metronidazole was high and that to clarithromycin was considerable. The present results motivate the need for a larger and continuing surveillance of *H. pylori* resistance in our country and worldwide.

**REFERENCES**


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