ANTIMICROBIAL AGENTS

Effects of silver sulphadiazine on the production of exoproteins by Staphylococcus aureus

VALERIE EDWARDS-JONES and H. A. FOSTER*

Department of Biological Sciences, Manchester Metropolitan University, Chester Street, Manchester M1 5GD and *Division of Biological Sciences, School of Environment and Life Sciences, University of Salford, The Crescent, Salford, Lancs M5 4WT

The effects of subinhibitory concentrations of silver sulphadiazine (AgSD) on exoprotein production in Staphylococcus aureus strains T1, T4, RN4282 and RN 4282agr were studied. AgSD markedly increased levels of toxic shock syndrome toxin (TSST)-1 in strains T4 and RN4282. This effect was independent of agr and AgSD restored TSST-1 production to the wild-type level in RN 4282agr. AgSD had no effect on enterotoxin A or coagulase activity in strains T1 or T4. Strain T4 produced enterotoxin C at high levels and no effect was observed with AgSD. AgSD repressed metalloprotease production in strain T4 but the overall protease activity remained the same. No change in proteolytic activities was seen in strain T1 with AgSD. Molecular mechanisms for these observations are discussed.

Introduction

Staphylococcus aureus is the most common cause of nosocomial infection and is of increasing concern because of the spread of methicillin-resistant strains and the occurrence of strains additionally showing resistance to vancomycin [1]. It is a pathogen that is capable of causing various diseases from mild skin infections through to life-threatening diseases such as toxic shock syndrome (TSS) [2]. The pathogenicity has been attributed to the ability of the organism to elaborate a large number of surface bound proteins, e.g., protein A, adhesins and secreted toxins, e.g., enterotoxins, exfoliative toxins, toxic shock syndrome toxin (TSST)-1. Other exoproteins are involved in the infection process, e.g., proteolytic enzymes, haemolysins, coagulase, lipase and deoxyribonuclease [3, 4]. The production of these virulence determinants is tightly controlled and is dependent on the phase of growth, cell density, metal ions and physiological conditions [5–7]. They are regulated by a number of global regulators including agr and sur [8, 9]. These contribute to the upregulation of exoproteins and the down-regulation or modification of surface-bound proteins in response to cell density and environmental stress, e.g., microaerobic conditions [6, 9].

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Corresponding author: Dr H. A. Foster (e-mail: h.a.foster@salford.ac.uk).
thus slowing the release of Ag into the medium [18].
Although AgSD interacts irreversibly with DNA to form a complex in vitro [23], this does not appear to be its mode of action in vivo and Ag\(^{2+}\) ion is the active agent [24]. Silver ions react with DNA in vitro, firstly in a weak interaction then strongly with the silver ions becoming incorporated by bonding between the base pairs [25]. Although it has been suggested that binding to DNA is the mode of action in vivo [18], other workers suggest that Ag\(^{2+}\) binds preferentially to external cell structures and that only small amounts are bound to DNA [24]. Cells of *S. aureus* treated with AgSD were enlarged, some retraction of the cytoplasmic membrane from the cell wall was seen and division planes between bacteria became indistinct, supporting the view that the effect is on external cell structures [26]. Studies have shown that subinhibitory levels of sulphadiazine affect membrane permeability [27]. This may enhance the entry of silver ions into cells. This study determined which component of Flammazine™ preparations was responsible for the effects on TSST-1 production and the effects of AgSD on other virulence factors were investigated.

Materials and methods

Bacterial strains and culture conditions

*S. aureus* strains T1 and T4 were as described previously [14]. Strain T1 (FR111895) was isolated from a case of menstrual TSS (MTSS) and originally supplied by M. S. Bergdoll. Strain T4 was originally isolated from a burned patient with confirmed TSS (Bury and Rochdale Healthcare Trust). Strain T1 showed no response to Flammazine™ whereas strain T4 produced at least eight-fold more TSST-1 in subinhibitory concentrations. Strains RN4282 and RN4282agpr were supplied by T. J. Foster, University of Dublin, and were originally from R. P. Novick. Cultures were grown in 50 ml of Brain-Heart Infusion Broth (BHI, Oxoid) in 100-ml conical flasks in an orbital incubator at 37°C and 200 rpm for 24 h. Culture supernates were obtained by centrifugation at 1500 \(g\) for 20 min and the supernatant fluids were removed and stored at \(-70°C\).

Viable counts

Samples were serially diluted in saline and samples were inoculated on to Nutrient Agar plates (Oxoid) by the method of Miles and Misra [28].

Determination of TSST-1 and enterotoxins

TSST-1 and enterotoxins A and C (SEA and SEC) in culture supernatant fluids were determined with reverse passive latex agglutination kits supplied by Unipath (Oxoid TD 40 and TD900) used according to the manufacturer’s instructions. Briefly, the supernates were double diluted in PBS supplied with the kit. This was mixed with an equal volume (25 \(\mu\)l) of sensitised latex particles and allowed to react at room temperature. The titre was determined as the last dilution showing agglutination. Hence the results were semi-quantitative and assumed that this titre represented a concentration of 2 ng/ml. Kts from the same batch were used within each experiment to ensure comparability. TSST-1 was also expressed as ng/ml/10^9 cells (specific toxin production).

Determination of the active components of Flammazine™

Subinhibitory concentrations were chosen that allowed comparison of growth and toxin production at 24 h. This had been determined previously and was 25 \(\mu\)g of Flammazine™/ml [14]. This was one-quarter of the MIC for strains T1 and T4. Details of the concentrations of each component within the preparation were obtained from the manufacturers and were subject to a confidentiality agreement. The respective concentrations of the individual ingredients were calculated and added to 9 ml of BHI in 20-ml universal bottles. The experiments were set up in triplicate. The ingredients were AgSD, polysorbate 60, polysorbate 80, glycerol monooleate, cetyl alcohol, propylene glycol and liquid paraffin. These were incubated at 37°C and 200 rpm for 24 h in an orbital incubator. The viable count and levels of TSST-1 were determined.

Effects of different concentrations of AgSD on TSST-1 production and the effects of agr

Cultures of strains T1, T4, RN4282 and RN4282agpr were grown in BHI supplemented with AgSD 0, 17, 25, 33 and 50 \(\mu\)g/ml for 24 h in an orbital incubator (37°C and 200 rpm) and viable count and TSST-1 production were determined.

Exoprotein production in subinhibitory concentrations of AgSD

Overnight broth cultures of *S. aureus* T1 and T4 in BHI were washed three times by centrifugation and resuspension in saline and then after the final wash the cells were resuspended in the same volume of fresh BHI; 0.5 ml of a 1 in 100 dilution of this broth was added to 49.5 ml of fresh BHI containing AgSD 25 \(\mu\)g/ml and incubated at 37°C and 200 rpm for 24 h in an orbital incubator. Samples (1 ml) were removed at 0, 3, 6, 24 and 48 h, the viable count was determined; the remainder was centrifuged at 1500 \(g\) and 4°C for 20 min and the supernatant fluids were stored at \(-70°C\).

Determination of coagulase activity

BHI (100 \(\mu\)l) was added to each well of a flat-bottomed microtitration plate. Supernatant fluid (100 \(\mu\)l) was added to the first well and doubling dilution was performed by transferring 100 \(\mu\)l into the next well and mixing thoroughly. This was repeated for eight wells.
The final 100 μl were discarded. Rabbit plasma (Prolab PL 850; 100 μl of a 20% v/v dilution in distilled water) was added to each well. The plate was shaken gently to mix the contents, covered and incubated at 37°C for 4 h. The titres were recorded as the final well showing a visible clot. Positive and negative controls from previously determined supernates were included in each batch.

**Determination of proteolytic activity**

A modification of the method of Lincoln and Leigh [29] was used. Samples of supernatant fluid (100 μl) were added to 1 ml of azocasein (Sigma; 1 mg/ml in 100 mM Tris-HCl, pH 7.2) and incubated at 37°C for 1 h. The reaction was stopped by addition of 1 ml of 5% w/v trichloroacetic acid and mixing; undigested azocasein was allowed to precipitate for 30 min. The mixture was then centrifuged at 10,000 g for 10 min and the absorbance of the supernate read at 328 nm. One unit of protease activity was defined as giving an absorbance of 0.001 after incubation for 1 h at 37°C.

Total protease activity was determined and the activities of serine, thiol and metallo-proteases were determined on the 24-h samples by addition of the appropriate inhibitor and subtraction from the total activity. Serine protease activity was inhibited by addition of 12.5 mM phenyl methyl sulphonyl fluoride (PM SF; Sigma), thiol protease activity was inhibited by addition of 12.5 mM iodoacetic acid (IAA; Sigma) and metalloprotease activity was inhibited by adding 12.5 mM ethylenediamine tetraacetic acid (EDTA; Sigma). The inhibitors were incubated with the supernates for 10 min at 37°C before assaying for residual protease activity. The results are the means of three determinations on each sample and the experiment was performed in duplicate.

**Statistical analysis**

Where possible, experiments were performed in triplicate and the results were analysed statistically by the unpaired t test with the Minitab program on an IBM-compatible PC.

**Results**

**Determination of the active component of Flammazine™**

There was an apparent decrease in viable counts at 24 h when the strains were grown in subinhibitory concentrations of Flammazine™ or its individual components but these differences were not statistically significant (p > 0.05). The results are shown in Table 1. There was an eight-fold increase in TSST-1 in the presence of Flammazine™ and AgSD and a two-fold increase in the presence of polysorbate 60 or 80 when compared with the levels in BHI alone. These differences were also reflected when TSST-1 production was expressed as specific toxin produced (ng/10^9 cfu/ml; Table 1).

**Table 1. The effects of the individual components of Flammazine™ on TSST-1 production in S. aureus strain T4**

<table>
<thead>
<tr>
<th>Component</th>
<th>Mean TSST-1 toxin (units)</th>
<th>Specific TSST-1 toxin (units/10^9 cfu/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (BHI only)</td>
<td>16</td>
<td>5.1</td>
</tr>
<tr>
<td>Flammazine™</td>
<td>128</td>
<td>76.6*</td>
</tr>
<tr>
<td>Silver sulphadiazine</td>
<td>128</td>
<td>66.7*</td>
</tr>
<tr>
<td>Polysorbate 60</td>
<td>32</td>
<td>21.1*</td>
</tr>
<tr>
<td>Polysorbate 80</td>
<td>32</td>
<td>14.5*</td>
</tr>
<tr>
<td>Glycerol monostearate</td>
<td>16</td>
<td>8.8</td>
</tr>
<tr>
<td>Cetyl alcohol</td>
<td>16</td>
<td>8.8</td>
</tr>
<tr>
<td>Propylene glycol</td>
<td>16</td>
<td>7.7</td>
</tr>
<tr>
<td>Liquid paraffin</td>
<td>8</td>
<td>3.6</td>
</tr>
</tbody>
</table>

Specific toxin production is expressed as units of toxin/10^9 cells/ml after 24 h. *p < 0.05.

The effects of different concentrations of AgSD and the effect of agr

Table 2 shows the response of S. aureus strains T1, T4, RN4282 and RN4282agr to different concentrations of AgSD and the results are expressed as both a titre of TSST-1 and as specific TSST-1, i.e., ng of toxin/10^9 cfu/ml. AgSD stimulated TSST-1 production in strain T4 at concentrations of ≥20 μg/ml. There was no effect on the TSST-1 titre in cultures of strain T1, although specific TSST-1 was slightly raised. AgSD also stimulated TSST-1 production in strain RN4282 and, as expected, TSST-1 was greatly reduced in strain RN4282agr but AgSD restored TSST-1 production to almost wild-type levels.

The effects of AgSD on the production of enterotoxins A and C

Both strains T1 and T4 produced enterotoxin A but only T4 produced enterotoxin C. In strain T1, enterotoxin A was detected at 6 h in the control broth (8 units) whereas in the presence of AgSD, it was detected only at 24 h. There was no difference in the amounts of enterotoxin A produced at 24 h in the presence of AgSD (64 units) but there was a slight increase (two-fold) at 48 h. In strain T4 there was no difference in the timing or final amount of enterotoxin A produced.

Strain T4 also produced enterotoxin C. AgSD did not affect the final amount produced when compared to the control.

The effects of AgSD on the production of coagulase

In strain T1, coagulase was detected at 3 h (12 units), increasing to 16 units at 6 h and reaching a maximum of 64 units at 24 h and 48 h. In the presence of AgSD, coagulase was detected only at 24 h and there were no
The effects of AgSD on the production of proteolytic enzymes

Fig. 1 shows the effects of AgSD on total protease production. Protease activity was detected in both strains at 3 h and the levels increased gradually to a maximum at 24 h. In the presence of AgSD, protease production was delayed but there was no significant difference (p >0.05) at 24 h when compared to the control broth. Strain T4 produced twice as much total protease when compared with strain T1. The activities of the individual proteases after 24 h are shown in Fig. 2. The predominant protease produced by strain T1 was thiol protease (72%) of the total activity followed by serine protease (20%) and metalloprotease (8%). In the presence of AgSD there was little difference from the control culture. Strain T4 produced two-fold higher protease and the predominant activity was metalloprotease (74%) followed by serine protease (23%) and thiol protease (3%). In the presence of AgSD the total protease activity was similar to the control but the relative proportions of the different activities were altered with serine and thiol-proteases being dominant (52% and 47%, respectively) and metalloprotease activity was greatly reduced (2%).

Discussion

The results obtained confirm that the marked increase in TSST-1 production in S. aureus strain T4 by subinhibitory concentrations of Flamazine™ was due to the active ingredient AgSD. The increase in TSST-1 production caused by AgSD also occurred in strains RN4282 and RN4282agr and was, therefore, independent of agr. Studies on the effects of AgSD on the other exoproteins produced by strains T1 and T4 showed that the effects of AgSD were dependent on the strain and the exoprotein. Strain T1 was originally isolated from a menstrual case of TSS and it is probable that the array of expressed virulence factors differs from strain T4, which was isolated from a patient with TSS resulting from an infected burn wound. Of the exoproteins studied here, strain T1 differed from T4 in the variety of proteases produced and strain T4 also produced enterotoxin C.

AgSD had no effect on enterotoxin A and C production in strain T4 and delayed enterotoxin A production in strain T1 without a marked effect on levels. AgSD had no effect on coagulase production by either strain. In this study, only one subinhibitory concentration of AgSD was used to determine if it had any global

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Table 2. The effect of different concentrations of AgSD on TSST-1 production in S. aureus T1, T4, RN4282 and RN 4282agr

<table>
<thead>
<tr>
<th>Concentration of AgSD (μg/ml)</th>
<th>T1</th>
<th>T4</th>
<th>RN4282</th>
<th>RN4282agr</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>128 (85.3)</td>
<td>128 (34.1)</td>
<td>256 (341)</td>
<td>16 (17.2)</td>
</tr>
<tr>
<td>17</td>
<td>128 (118.5)</td>
<td>128 (26.9)</td>
<td>2046 (2153)</td>
<td>128 (284.4)</td>
</tr>
<tr>
<td>20</td>
<td>256 (204.8)</td>
<td>1024 (240.9)</td>
<td>2046 (2046)</td>
<td>128 (232)</td>
</tr>
<tr>
<td>25</td>
<td>128 (128)</td>
<td>1024 (195)</td>
<td>1024 (731.4)</td>
<td>192 (240)</td>
</tr>
<tr>
<td>33</td>
<td>256 (205)</td>
<td>1024 (1024)</td>
<td>1024 (929.5)</td>
<td>64 (160)</td>
</tr>
<tr>
<td>50</td>
<td>256 (85.3)</td>
<td>1024 (341)</td>
<td>1024 (1625)</td>
<td>24 (36.9)</td>
</tr>
</tbody>
</table>

Specific toxin (in brackets) is expressed as ng/10^7 cells. The results are expressed as the mean of three experiments. The MIC of AgSD for all strains was 100 μg/ml.
effects on other toxins or just affected TSST-1. AgSD appeared to markedly increase TSST-1 only in certain strains while other toxins were not affected.

AgSD increased TSST-1 production significantly in strain T4 and the reason for this is unknown and requires further investigation. Transcription of tsf is under the control of agr [9]. The effector molecule, RNAIII, is synthesised in mid-to-late exponential growth phase as a result of induction by an autocrine pheromone octapeptide [6]. RNAIII induces transcription of tsf by an unknown mechanism but tsf mRNA does not appear until one or two generations after RNAIII synthesis [30]. Expression of agr and tsf is also affected by sarA, another regulatory molecule, which is produced in response to microaerobic conditions [31]. Production of TSST-1 toxin has been shown previously to occur earlier in the growth cycle in the presence of Flamazine™ [14] and the same effect has been observed with AgSD (unpublished data). Why this earlier expression of TSST-1 should occur is unknown but these observations, alongside the increased TSST-1 levels observed in the agr mutant, suggest that AgSD does not affect agr regulation. Production of enterotoxin A, which is also co-ordinately regulated by agr, was not affected, again suggesting that the effect was independent of agr. It could be postulated that the earlier expression of tsf might account for the increased levels of TSST-1. However, comparison of TSST-1 production at 48 h and 72 h (data not shown) still showed a four-fold increase in the presence of AgSD compared with the control broth.

It is known that silver ions react with both proteins and nucleic acids. It is possible that Ag⁺⁺ may cause distortion to DNA and allow activation of genes such as tsf by binding to the DNA and facilitating RNA–polymerase binding. Studies have shown that when S. aureus is grown in subinhibitory concentrations of novobiocin, known to cause conformational changes to DNA, increased levels of eta are produced [32]. In attempts to isolate DNA from AgSD-treated cells and, with increasing AgSD concentrations, the DNA becomes increasingly yellow in colour and its solubility is also affected (unpublished observations). The inhibition of TSST-1 production caused by the agr mutation in strain RN4220agr was effectively reversed in the presence of AgSD, again suggesting an interaction with the DNA.

Other possibilities for TSST-1 being increased in the presence of AgSD include enhanced stability of tsf mRNA, although it would not explain the earlier appearance of TSST-1 in the growth cycle. Previous workers have shown that metals, including Ag⁺⁺, increased the resistance of tobacco mosaic virus RNA to ribonuclease without loss of infectivity [33]. The effects of AgSD on tsf mRNA and RNAIII are currently being investigated. Another possibility is at the level of translation and more detailed studies need to be undertaken to unravel the possible effect(s). It also may be possible that AgSD enhances secretion of TSST-1 although there is no evidence that TSST-1 is stored intracellularly before excretion. The protein is translated from the mRNA as a pro-toxin with a 40 amino acid N-terminal signal peptide that is removed during transport across the membrane [34]. Finally, inhibition of the degradation of toxin by proteolytic enzymes could enhance the amount of toxin present in the supernate by inhibiting turnover. Other workers have shown that S. aureus isolates from cases of MTSS produced thiol protease (type II) as their major class of protease [35]. Strains that produced predominantly serine protease (type I) were not associated with cases of TSS [36]. Serine protease was later shown to digest TSST-1 [37] and it was postulated that strains producing this as a predominant protease produced little or no TSST-1. In this study, strain T1 produced predominantly thiol protease and strain T4 produced predominantly metalloprotease. AgSD did not affect total protease but inhibited metalloprotease and increased thiol and serine proteases. It has been shown recently that different proteases are responsible for cleavage of different exoproteins and Chan and Foster [31] suggest a major role for proteases in modulation of virulence determinants. For example, metalloprotease activates serine protease by proteolytic cleavage [38] and the serine protease modifies surface-bound proteins [39]. Furthermore, a role has been suggested for proteases in the activation of exocellular lipases in S. hyicus [40, 41] and S. aureus [42]. The results showed that production of metalloprotease was greatly reduced by AgSD in strain T4 but not T1. This might suggest a role for metalloprotease in the turnover of TSST-1.

Chan and Foster [8] showed that incorporation of EDTA enhanced TSST-1 production by stimulating tsf expression. Whether or not this was due to inhibition of metalloprotease by EDTA was not fully investigated. Further investigations are needed to determine whether other metalloprotease inhibitors have a
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similar effect, such as 1,10-phenanthroline, would be worth trying. It has been shown that there are at least five proteases in S. aureus 8325–4 [31] and further work is required to determine whether AgSD specifically affects any or all these.

Overall, the results show that the effects of AgSD are strain dependent and the differences between responder and non-responder strains are being investigated.

We thank B. P. Novick and T. J. Foster for gifts of strains and Oxoid for supply of toxin assay kits at reduced cost.

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