IMMUNOLOGICAL RESPONSES OF GUINEA-PIGS TO BERYLLIUM SALTS

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Chronic pulmonary disease (Weber and Engelhardt, 1933) and contact dermatitis (DeNardi, Van Ordstrand and Carmody, 1949) have been observed in many beryllium workers. An immunological mechanism was first suggested (Sterner and Eisenbud, 1951) because only a small percentage of exposed workers developed chronic lesions, with an insidious onset several months after exposure. The histological picture of the granulomatous lesions was characteristic. Sterner and Eisenbud's hypothesis was supported by the fact that the cutaneous reactions to soluble beryllium salts of the delayed hypersensitivity type (Curtis, 1951) were seen in patients suffering from berylliosis, and granuloma formation in people who were cut by beryllium phosphor-coated glass from broken lamps (Dutra, 1951). Furthermore, beryllium fluoride is an excellent sensitiser in healthy man (Curtis; McCord, 1951).

Similar responses have been produced experimentally in animals. Chiappino, Cirla and Vigliani (1969) demonstrated that after endotracheal injection of beryllium oxide (BeO) a lung reaction characteristic of hypersensitivity developed in some guinea-pigs, and that these animals also had dermal hypersensitivity to soluble salts. Furthermore, the application of the sulphate (BeSO₄) to the ear induced the same histological changes in the regional lymph-node as known sensitisers. A state of delayed hypersensitivity has also been induced with various salts of beryllium given by different routes (Belman, 1957, 1969; Alekseeva, 1966; Polák, Barnes and Turk, 1968; Chiappino et al., 1969). These findings suggest that the tissue reactions induced by beryllium are immunological ones of the delayed hypersensitivity type. This was confirmed in guinea-pigs by passive transfer by means of lymphoid cells (Cirla, Barbiano di Belgiojoso and Chiappino, 1968), and by the inhibition of the intradermal reaction to beryllium by antilymphocytic serum (Chiappino, Barbiano di Belgiojoso and Cirla, 1968).

The work reported here shows that immunological paralysis (or tolerance) to beryllium can be readily induced in adult guinea-pigs, and can be recognised by inhibition of the delayed skin-hypersensitivity reactions to the topical application of beryllium in pre-treated animals. Beryllium compounds may induce either sensitisation or tolerance. Beryllium as the citrate is tolerogenic, whereas a compound more easily phagocytosed or one that reacts with proteins is immunogenic. This supports the conclusions previously drawn by Macher and Chase (1969a) from the results of their work on the elimination from the skin of

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various sensitisers (dinitrochlorobenzene and picryl chloride) injected intradermally. It is also shown that the tolerant state induced by beryllium can fit the model described by Mitchison (1964, 1968) for protein antigen.

**MATERIALS AND METHODS**

**Animals.** Male guinea-pigs of the Dunkin Hartley (Porton) strain from M. Mitchard, Dewbrook, Vines Cross Road, Horam, Sussex (source 1), and A. Tuck & Son Ltd, Laboratory Animal Breeding Station, Rayleigh, Essex (source 2) were used. The guinea-pigs weighed $577 \pm 13$ g (SE for $n = 94$), except in the comparison of the responses in the animals from the two sources, when smaller ones were used (table I); they were fed on diet S.G.1 (Short and Gammage, 1959) with 1.5 kg of added vitamin C per ton.

**Preparations of beryllium used.** BeF$_2$ (stock solution 20 per cent. in water) and BeSO$_4$.4H$_2$O were dissolved in de-ionised water or in MCWT (methyl-cellosolve : water : Tween “80” :: 45 : 45 : 10). 1-064m-$\text{BeSO}_4$ (aqueous solution) was incorporated in an equal volume of complete Freund adjuvant (Difco). Be phosphate was prepared by mixing m-$\text{BeSO}_4$$_4$, and 3m-Na$_2$HPO$_4$; the pH of the final suspension was adjusted with NaOH until a slight turbidity appeared ($\text{pH}$ approximately 7-4). Be citrate was prepared by mixing equimolar solutions (unless otherwise indicated) of BeSO$_4$.4H$_2$O and trisodium citrate. Some solutions were made radioactive by addition of carrier-free $^7\text{BeCl}_2$ (Radiochemical Centre, Amersham, Bucks.).

**Administration of the beryllium solutions.** Except for percutaneous application when BeF$_2$ and BeSO$_4$.4H$_2$O were used dissolved in MCWT, beryllium salts were injected as an aqueous solution of different concentrations in the following volumes: 0.1 ml intradermally (under ether anaesthesia, with 0.5-ml syringes sealed with warmed wax to avoid any leakage); 4 ml per kg intraperitoneally and 1 ml per kg intravenously (through the penis vein, under ether anaesthesia).

**Skin test for the determination of sensitivity.** The hair on the back was removed with electric clippers the day before the application of 0.03 ml of 0.02, 0.1 or 0.4 m-BeF$_2$ solution with MCWT per cm$^2$. The solutions were spread in the direction of the hair growth and allowed to dry, sometimes assisted by an electric hair-drier. The skin reactions were examined after 48 hr and scored as follows: (0) no reaction or doubtful; (1) mild but appreciable reactions; (2) obvious definite redness; (3) strong reaction; (4) very strong reaction with oedema of the skin and some haemorrhagic spots. The results are expressed as the number of animals in each group that responded and as the mean score of all the animals in a group. The maximum theoretical score that any animal could contribute was 12.

**Sensitisation.** For sensitisation, skin tests as described above were repeated at 7-day intervals for 2-4 wk. The results are expressed as previously and also as the mean score for a series of skin tests. Attempts were also made to sensitise the animals by intradermal, intraperitoneal or intravenous injections. In some experiments after intradermal injection, the injection sites were removed 2 hr later by excising them under ether anaesthesia.

Statistical examinations were done by analysis of variance, with a two-way classification to allow comparison of different groups of animals versus the different repeated skin tests. Where there was no statistical difference within the repeated skin tests, the results were pooled and analysis repeated by the previous method. When the ratio (F) was significant, the means were compared by Duncan’s multiple Range Test (Duncan, 1955) or Least Significant Difference; the level of significance was taken as $P \leq 0.01$.

**Elimination of beryllium from the skin.** Radioactive solutions of Be fluoride, sulphate and citrate were injected intradermally in a volume of 0-1 ml into the skin of the back under ether anaesthesia. Two or twenty hours later, the animals were killed and the injection sites excised. The radioactivity due to $^7\text{Be}$ was measured with a Packard Autogamma Scintillation Counter. The results are expressed as the percentages of radioactivity remaining in the skin.
RESULTS

Sensitisation of guinea-pigs by beryllium compounds

Different factors were investigated, the results of which are partly summarised in table I.

Salts of beryllium. Fluoride and sulphate given to guinea-pigs from source 1 showed that BeF₂ was a much more potent sensitiser than BeSO₄ as demonstrated by the greater number (60 per cent.) of animals sensitised after three applications compared with 30 per cent. for sulphate, and by the rapidity of the development as well as the intensity of the sensitisation. Some non-specific skin reactions were also observed after the first application of Be fluoride.

Source of guinea-pigs. Guinea-pigs of the same strain supplied by two breeders were not equally susceptible to the percutaneous application of BeF₂. Analysis of variance showed that greater numbers of animals from source 2 (100 as against 60 per cent.) were sensitised and the reactivity was more pronounced than in those from source 1. Guinea-pigs from source 2 were used in further experiments.

Reproducibility. Analysis of variance showed that groups of animals from source 2 did not differ in their reactivity to beryllium, so that they were pooled.
Comparison between the different skin tests showed that only the cutaneous reactions following the first application of beryllium differed significantly from those observed during subsequent applications. These were, in fact, non-specific inflammatory reactions; the mean score for a series of skin tests was therefore determined for the period of active sensitisation itself, and the first skin test was excluded.

**Influence of the route of administration.** The dose of beryllium administered by topical application of BeF₂ for each skin test was 140 μg (i.e., 350 μg Be per kg), part of which remained on the surface of the skin. Attempts to elicit skin sensitisation were also made by injecting solutions of beryllium compounds. Sensitisation was judged 15 days later by skin testing with BeF₂ in MCWT and by active sensitisation elicited by subsequent repeated skin paintings.

1. **Intradermal (ID) injection** (table VII, groups 2, 3, 5). Analysis of variance on the intensity of the skin reactions did not demonstrate any difference between the various groups given injections of different salts of beryllium. However, on the basis of the number of animals that reacted to beryllium during the first and second skin test, there was some indication that fluoride and sulphate were better sensitisers than citrate.

In a comparison between animals in which beryllium was first administered either percutaneously or intradermally (table VII, group 1 versus groups 2, 3, 5), it appeared that the percutaneous route was much more efficient in producing sensitisation. However, the dose given percutaneously was much greater and the least amount of beryllium that would produce sensitisation is unknown.

2. **Intravenous (IV) injection.** Injection of beryllium (19.15 μg per kg) by the IV route had no sensitising effect as shown by skin testing 15 days later, and did not modify significantly further active sensitisation (see table IV, fluoride; table VII, group 8, citrate).

3. **Intraperitoneal (IP) injection.** A dose of 5 μg Be per kg given 25 and 18 days before skin testing reduced the non-specific reactions observed after the first skin test and decreased the active sensitisation elicited by repeated skin paintings. This partial immunological paralysis was demonstrated (standard controls, table I) by the fact that only 25 per cent. of the animals were sensitised after three topical applications of BeF₂ compared with 50 per cent. of untreated guinea-pigs, and there was a reduction in the intensity of the skin reactions.

**Immunological paralysis induced in adult guinea-pigs**

The treatment needed to induce tolerance was investigated in more detail. Guinea-pigs from source 2 were given an intraperitoneal injection of various beryllium compounds 7 or 15 days or on both days before their immunological state was checked by skin testing with BeF₂. Active sensitisation was performed by repeated applications for 2–4 weeks and comparisons made with untreated controls. Doses of beryllium compounds are expressed in μg Be per kg. Different factors were investigated, and the results are given in tables II–V.
Influence of the dose (table II). The degree of immunological paralysis judged by the number of animals responding and by the intensity of the sensitisation obtained after further repeated skin paintings, was determined in guinea-pigs that had been given an injection of BeF₂ intraperitoneally.

**Table II**
Effect of previous intraperitoneal injection of BeF₂ on the sensitisation of male guinea-pigs by percutaneous application of BeF₂ in MCWT

<table>
<thead>
<tr>
<th>Pre-treatment with Be (as BeF₂)</th>
<th>number of animals giving positive responses/number tested, and (below and to right) mean score in skin test no.</th>
</tr>
</thead>
<tbody>
<tr>
<td>(μg per kg)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>on days before first skin test 1 2 3 4</td>
</tr>
<tr>
<td></td>
<td>15 7</td>
</tr>
<tr>
<td>0 (controls)</td>
<td>5/10 0.70 10/10 4.30 5/15 3.20 10/10 3.90</td>
</tr>
<tr>
<td>4.78</td>
<td>2/5 0.4 4/5 2.6 4/5 2.0 4/5 2.4</td>
</tr>
<tr>
<td>9.57</td>
<td>1/5 0.2 1/5 0.2 4/5 1.6 4/5 2.4</td>
</tr>
<tr>
<td>19.15</td>
<td>0/5 0.2 3/5 0.6 3/5 1.6 3/5 1.4</td>
</tr>
<tr>
<td>9.57</td>
<td>0/5 0 3/5 1.8 2/5 1.8 3/5 2.0</td>
</tr>
<tr>
<td>19.15</td>
<td>0/5 0 1/5 0.6 3/5 1.2 3/5 1.0</td>
</tr>
<tr>
<td>38.3</td>
<td>2/5 1.2 3/5 0.8 1/5 0.2 1/5 0.6</td>
</tr>
<tr>
<td>76.6</td>
<td>1/5 0.2 2/5 0.8 ... ... ... ...</td>
</tr>
<tr>
<td>383</td>
<td>2/5 0.4 4/5 3.2 4/5 4.4 4/5 3.8</td>
</tr>
</tbody>
</table>
FIG. 1a.—Percentage of responders derived from two series of skin tests, excluding the first skin test of each series so as to exclude non-specific responses.

FIG. 1b.—Mean scores for the intensity of the skin reactions in two series of skin tests, excluding the first skin test of each series so as to exclude non-specific responses.

FIG. 1.—Immune paralysis induced in male guinea-pigs by pre-treatment with various doses of BeF₂ IP. ☒ No pre-treatment. ■ Pre-treatment at −15 and −7 days. □ One pre-treatment at −15 days.
The response of the pre-treated animals differed significantly from the controls except in those given the highest dose (383 µg Be per kg).

Although no difference can be demonstrated at the 1 per cent. level between the groups pre-treated with various doses of beryllium, there was an indication that the degree of immune tolerance was approximately proportional to dose in the range 4.78–38.3 µg per kg (fig. 1a and b). At high doses no immune paralysis was produced.

**Duration of immunological paralysis.** This was studied by repeating the skin tests at 7-day intervals for 76 days after the injection of beryllium. The duration of the tolerant state is, to a certain extent, also dose-related, as is illustrated by the number of animals that remained non-responders in comparison with those not pre-treated with beryllium. Thus, of guinea-pigs given 9.57, 19.15 and 38.3 µg Be per kg, 20, 40, and 60 per cent. respectively were still non-responders after 76 days (table III).

**Table III**

*Duration of immunological paralysis induced by pre-treatment with a range of doses of BeF₂ injected IP before the first percutaneous skin test with BeF₂.*

<table>
<thead>
<tr>
<th>Pre-treatment with Be as BeF₂</th>
<th>Number of animals giving positive response/number tested, in skin tests on day after first pre-treatment injection</th>
</tr>
</thead>
<tbody>
<tr>
<td>µg per kg</td>
<td>15</td>
</tr>
<tr>
<td>-----------------</td>
<td>----</td>
</tr>
<tr>
<td>0 (controls)</td>
<td>5/10</td>
</tr>
<tr>
<td>4.78</td>
<td>2/5</td>
</tr>
<tr>
<td>9.57</td>
<td>1/5</td>
</tr>
<tr>
<td>19.15</td>
<td>1/5</td>
</tr>
<tr>
<td>38.3</td>
<td>0/5</td>
</tr>
</tbody>
</table>

*Influence of beryllium compounds* (table IV). The next question to answer was whether other beryllium salts could induce tolerance. Beryllium citrate, which remains in solution at physiological pH, and beryllium phosphate, an insoluble compound, were injected IP in a single pre-treatment at −15 days, and the degree of tolerance compared on day 0 with that of uninjected controls and with that of guinea-pigs pre-treated with the same tolerogenic dose (19.15 µg Be per kg) of fluoride. Statistical examination shows that at the 1 per cent. level, fluoride and citrate induced a significant degree of tolerance, whereas phosphate did not.

*Influence of the immunological state of the guinea-pigs.* BeF₂ injected IP in immunologically virgin animals induced a dose-related tolerance in a low dose-range (table II); the same doses injected into guinea-pigs already sensitised by
**TABLE IV**

*Effect of beryllium salts injected 7 days before first skin test on the sensitisation of male guinea-pigs by percutaneous application of BeF₂ in MCWT*

<table>
<thead>
<tr>
<th>Pre-treatment with BeF₂</th>
<th>Other pre-treatment</th>
<th>Number of animals giving positive responses/number tested, and (below and to right) mean score in skin test no.</th>
<th>Mean percentage* of responders, and (below and to right) mean score, for skin tests 2, 3, and 4</th>
</tr>
</thead>
<tbody>
<tr>
<td>as</td>
<td>at concentration (µg per kg)</td>
<td>by route</td>
<td>1</td>
</tr>
<tr>
<td>...</td>
<td>0 (controls)</td>
<td>...</td>
<td>5/8</td>
</tr>
<tr>
<td>BeF₂</td>
<td>10.8</td>
<td>IP</td>
<td>0/5</td>
</tr>
<tr>
<td>BeF₂</td>
<td>10.8</td>
<td>IP 15 ml glycogen solution IP 9 and 8 days before 1st skin test</td>
<td>0/5</td>
</tr>
<tr>
<td>...</td>
<td>0 (controls)</td>
<td>...</td>
<td>1/5</td>
</tr>
<tr>
<td>BeF₂</td>
<td>19.15</td>
<td>IP</td>
<td>0/5</td>
</tr>
<tr>
<td>Be phosphate</td>
<td>19.15</td>
<td>IP</td>
<td>1/5</td>
</tr>
<tr>
<td>Be citrate</td>
<td>19.15</td>
<td>IP</td>
<td>0/5</td>
</tr>
<tr>
<td>BeF₂</td>
<td>19.15</td>
<td>IP adjuvant added to BeF₂</td>
<td>2/5</td>
</tr>
<tr>
<td>BeF₂</td>
<td>19.15</td>
<td>IV</td>
<td>0/4</td>
</tr>
</tbody>
</table>

* The reactions in the first skin test, which are mainly non-specific and inflammatory in nature, have been ignored in the calculation of the means in this column.
† Significantly different from the results for guinea-pigs pre-treated with BeF₂ or Be phosphate.
skin tests did not modify their immunological state, i.e., did not desensitise the animals (unpublished observations). However, bigger doses (400 µg per kg) injected intravenously in the form of sulphate gave a transient desensitisation 7 days later, thus confirming the findings of Polák and Turk (1968), who used beryllium lactate.

*Influence of the route of administration.* The induction of immune tolerance by pre-treatment with beryllium can be induced by IP injection only, for the same dose given ID (table VII) or IV (tables IV and VII) does not modify the immunological status significantly, except perhaps when beryllium citrate is used (group 8, table VII).

*Other experimental conditions modifying the induction of tolerance.* The differences observed in the induction of tolerance by various beryllium compounds suggested a possible role played by their particle size. Citrate seems to be the most tolerogenic salt. To test the effect of phagocytosis by the reticulo-endothelial (RE) system, a tolerogenic dose of BeF$_2$ (19.15 µg Be per kg) was injected IP in animals in which the RES activity was stimulated by incorporating BeF$_2$ in complete Freund adjuvant (table IV). Comparison of skin reactions in these animals with those in animals given the same dose of beryllium in water showed a significantly greater degree of sensitisation, similar to that seen in untreated controls given repeated skin paintings.

The tolerogenic activity of beryllium (10.8 µg per kg) was also found to be abolished by pre-treatment of the guinea-pigs with a solution of glycogen IP, which is known to stimulate macrophage proliferation (table IV).

*Other results*

As shown previously, beryllium can induce either skin sensitisation or immune tolerance depending on the route of administration and perhaps on the nature of the salts.

Therefore, the question arises to what extent these two phenomena take place during sensitisation by skin painting. This could be the result of a concomitant immunisation and tolerance, the former being predominant.

More detailed investigations were also done on the fate of injected beryllium into the skin in order to know more about its binding by skin proteins.

*Elimination from the skin of labelled beryllium injected intradermally (ID).* Solutions of beryllium compounds were made radioactive by addition of $^{7}$Be, and 0.1 ml of various concentrations was injected ID into immunologically virgin guinea-pigs. The skin was removed 2 and 20 hr later and the residual radioactivity measured. Table V shows that the elimination from skin was biphasic; within 2 hr quite a large amount of beryllium disappeared, and then the elimination continued slowly over the next 18 hr. This disappearance was much more striking for beryllium citrate at the higher concentration.

The influence of concentration was studied in more detail with beryllium citrate. The results (fig. 2) showed that the percentage residual radioactivity in the skin 2 hr after ID injection was inversely proportional to the molarity of beryllium from 0.001 to 0.05 M; then the rate of elimination became constant.
TABLE V
Elimination of 7Be-labelled compounds and carrier-free 7BeCl₂ from the intradermal injection sites

<table>
<thead>
<tr>
<th>Compound injected*</th>
<th>Concentration (m)</th>
<th>Mean percentage±SE of initial radioactivity remaining in skin site† at time after injection (hr)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>2</td>
</tr>
<tr>
<td>BeF₂</td>
<td>0.01</td>
<td>74.7±2.40 (11†)</td>
</tr>
<tr>
<td></td>
<td>0.001</td>
<td>69.2±2.56 (6)</td>
</tr>
<tr>
<td>BeSO₄</td>
<td>0.01</td>
<td>78.0±3.46 (6)</td>
</tr>
<tr>
<td></td>
<td>0.001</td>
<td>73.4±4.82 (5)</td>
</tr>
<tr>
<td>Be-citrate</td>
<td>0.01</td>
<td>36.3±0.85 (6)§</td>
</tr>
<tr>
<td></td>
<td>0.001</td>
<td>58.8±2.93 (6)∥</td>
</tr>
<tr>
<td>7BeCl₂ (carrier-free)</td>
<td>0.01</td>
<td>12.4±2.36 (6)</td>
</tr>
</tbody>
</table>

* In a volume of 0.1 ml.
† Numbers of animals in brackets.
‡ No radioactivity was found in skin outside the injection sites.
§ These figures are less than those for BeSO₄ or BeF₂ (P<0.001).
∥ These figures are less than those for BeSO₄ or BeF₂ (0.01<P<0.02).

Fig. 2.—Elimination of 7Be citrate (0.1 ml) of various concentrations from the intradermal injection site. Results expressed as the percentage of initial radioactivity remaining in the skin 2 hr after injection.
IMMUNOLOGICAL RESPONSES OF GUINEA-PIGS TO BERYLLIUM

at a maximum level since 90 per cent. disappeared within the first 2 hr. Any variation in the rate of elimination dependent on the site of injection was checked by injecting 0·01M-BeF₂ into various parts of the skin of the back on one animal. The mean score for six well-separated spots (76±2·75 per cent.) did not differ from that obtained in other experiments done in individual guinea-pigs (74·4±2·40 per cent.). The rate of elimination probably depends on the binding of beryllium by the skin constituents. It has been shown that

**TABLE VI**

Mean percentage radioactivity (±SE) remaining in the skin 2 hr after ID injection of ⁷Be-labelled beryllium salts in guinea-pigs in different physical states

<table>
<thead>
<tr>
<th>Immunological state of guinea-pigs</th>
<th>Mean percentage radioactivity (±SE) remaining in injection area 2 hr after ID injection of Be citrate at concentration (m)</th>
<th>BeF₂ at concentration (m)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0·01</td>
<td>0·001</td>
</tr>
<tr>
<td>&quot;Virgin&quot; (V)</td>
<td>36·3±0·85 (6)*</td>
<td>58·8±2·93 (6)†</td>
</tr>
<tr>
<td>Sensitised (S)</td>
<td>44·6±1·15 (7)</td>
<td></td>
</tr>
<tr>
<td>Given 19·15 µg Be per kg IP 2 days before expt (I)</td>
<td>41·0±1·53**</td>
<td>47·2±2·78‡ ††</td>
</tr>
<tr>
<td>Immunologically paralysed by injection of 19·15 µg Be per kg IP 15 days before expt (P)</td>
<td>44·5±4·50 (8)</td>
<td>41·4±3·51 (8)</td>
</tr>
</tbody>
</table>

* Number of guinea-pigs used in brackets.
† Significantly greater (P<0·001) than figure at 0·01M.
‡ Significantly greater (0·02<P<0·05) than figure at 0·01M.
§ Not significantly different from figure at 0·01M.
|| In comparison with S, V figure is significantly less (P<0·001).
§§ In comparison with S, V figure is significantly greater (0·02<P<0·05).
** In comparison with I, V figure is significantly less (P<0·001).
†† In comparison with I, V figure is significantly greater (0·001<P<0·01).

non-specific skin reactions are observed during the first skin test (table I), and that they are suppressed by the previous injection of a tolerogenic dose of beryllium (table II). Therefore, the question arises whether prior contact with beryllium modifies the binding to skin of beryllium given later. The elimination from the skin was therefore studied in animals already sensitised by skin painting, animals made tolerant with BeF₂ given IP, and animals given an IP injection of the same tolerogenic dose of beryllium (19·15 µg per kg), but 2 days earlier (table VI). The rates of elimination of two different molar concentrations of BeF₂ (0·01 and 0·001 M) in the various groups were identical with that found in immunologically virgin animals.
### TABLE VII

*Effect of ID injection of beryllium compounds (0.01μ) 15 days before first skin test and of the excision of injection sites on further sensitisation elicited by percutaneous application of BeF₂ in MCWT*

<table>
<thead>
<tr>
<th>Group no.</th>
<th>Pre-treatment</th>
<th>Injection site</th>
<th>Number giving positive responses/number tested, and (below and to right) mean score, in skin test no.</th>
<th>Mean percentage of responders, and (below and to right) mean score in skin tests 2, 3 and 4</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>none (standard controls)</td>
<td>none</td>
<td>11/23 0.83 21/23 4.35 18/18 5.0 22/23 4.61</td>
<td>95 4.65</td>
</tr>
<tr>
<td>2</td>
<td>BeSO₄, 23.6 μg per kg ID</td>
<td>left in situ</td>
<td>4/5 1.80 5/5 4.00 5/5 2.60 3/5 2.00</td>
<td>87 2.9</td>
</tr>
<tr>
<td>3</td>
<td>BeF₂, 26.8 μg per kg ID</td>
<td>left in situ</td>
<td>4/5 1.60 5/5 3.60 5/5 3.60 4/5 3.00</td>
<td>100 3.4</td>
</tr>
<tr>
<td>4</td>
<td>BeF₂, 10.1 μg per kg ID</td>
<td>excised 2 hr after injection</td>
<td>3/5 0.8 1/5 0.20 2/5 1.80</td>
<td>... ... 30 1.0</td>
</tr>
<tr>
<td>5</td>
<td>Be citrate, 26.3 μg per kg ID</td>
<td>left in situ</td>
<td>1/4 0.5 3/4 3.25 4/4 2.75 3/3 2.67</td>
<td>91 2.9</td>
</tr>
<tr>
<td>6</td>
<td>Be citrate, 24.6 μg per kg ID</td>
<td>excised 2 hr after injection</td>
<td>0/5 0 5/5 4.0 5/5 5.20 5/5 4.20</td>
<td>100 4.47</td>
</tr>
<tr>
<td>7</td>
<td>Be citrate, 19.15 μg per kg ID</td>
<td>excised 2 hr after injection</td>
<td>4/5 2.0</td>
<td>†</td>
</tr>
<tr>
<td>8*</td>
<td>Be citrate, 19.15 μg per kg IV</td>
<td>none to excise</td>
<td>3/5 1.20 3/5 2.00 5/5 2.60</td>
<td>... ...</td>
</tr>
</tbody>
</table>

* Group added to complete experiment given in table IV, and to be compared with the same dose of BeF₂ IV.  † Animal infected; killed.
After beryllium citrate the elimination rate was fairly constant in the various groups that had received beryllium previously, either percutaneously (sensitised group) or parenterally (tolerant and injected groups). In these animals, therefore, unlike the findings in virgin animals, no statistical difference was noticed between the two concentrations used.

**Influence of excision of the sensitising injection sites.** After an ID injection of beryllium, one part escapes very quickly, the other remains *in situ*; the immunological role played by these two parts of beryllium was examined. Guinea-pigs were given an intradermal injection of 0.1 ml of 0.01M BeF₂ or citrate. The injection sites were excised 2 hr later and the immunological state was checked 15 days later by skin testing with BeF₂. Comparisons were made with animals given an injection without excision of the sites of injection. The results (table VII) showed that after excision the degree of sensitisation was reduced in the group given an injection of fluoride (P = 0.05) (comparison groups 3, 4) when this was compared with the group in which the skin was left intact. With citrate the excision of the injection sites did not modify the immunological response (groups 5, 6). The conclusion is that the rate of disappearance depends on the chemical form of the injected beryllium.

**DISCUSSION**

It has been shown that in guinea-pigs beryllium induces either skin sensitisation or immunological paralysis (tolerance). The state of tolerance is recognised

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**Fig. 3.**—Factors affecting the immunological response to beryllium salts, and a suggestion on the methods by which they do so.
by inhibition of the specific delayed skin hypersensitivity reactions elicited by topical application of the hapten, i.e., beryllium. The factors that may be involved are summarised in fig. 3.

A specific delayed hypersensitivity results only from the contact of beryllium with skin; parenteral injection fails to sensitise or may induce an immunological tolerance. The particular role of the skin seems to depend upon the combination of beryllium with one of its constituents, leading to an allergenic depot with subsequent slow release of antigenic material. The difference observed in the sensitising effect of fluoride and sulphate applied percutaneously (table I) may depend on their diffusion properties, which enable them to pass through the skin; indeed, injected intradermally both bind to the skin to the same extent (table V) and have the same immunological action (table VII). Beryllium reacts strongly both in vitro and in vivo with serum proteins (Aldridge, Barnes and Denz, 1949; Belman, 1957; Vacher and Stoner, 1968a), and in vivo beryllium can react with the skin (Belman, 1969). Beryllium also inhibits some enzymes (Thomas and Aldridge, 1966) and might also react with cell membranes (as suggested by Belman, 1969). The percutaneous application of beryllium gives rise to non-specific skin reactions, and after ID injection most of it becomes fixed to the tissues and escapes slowly from the injection site (table V). In rabbits, beryllium has been shown to remain in situ after subcutaneous injection for as long as 3 mth (Aldridge et al.). Whether or not this is due to a lack in some animals of a specific protein to act as carrier in the antigenic combination with beryllium, or to differences in the lymphocyte population is not known. The role of the combination of beryllium with skin constituents in the induction of contact sensitisation is further demonstrated by the experiments on the excision of the intradermal injection sites. The excision of the allergenic depots of beryllium (as BeF$_2$) 2 hr after injection, is followed by a partial immune paralysis (table VII), presumably due to a diffusible form of beryllium escaping very rapidly from the injection site (table V).

Therefore, it seems likely that the sensitisation elicited either by ID injection or percutaneous application is the resultant of concomitant immunisation (by skin-bound beryllium) and tolerance (by diffusible beryllium). The same conclusion has been drawn from recent experiments on the immune paralysis that develops after the excision of allergenic depots of other haptens injected ID (DNCB-picrylchloride, Macher and Chase, 1969a) or applied percutaneously (DNCB, Lowney, 1965).

It was noticed that the diffusible fraction of beryllium increased with the molarity of the solutions injected (fig. 3). An increase of this fraction, as for instance with citrate (Van Cleave and Kaylor, 1953), could explain why the sensitisation obtained by intradermal injection of high doses of beryllium was less than that after smaller doses (Alekseeva, 1966) or why the degree of sensitisation after intradermal injection was lower than after topical application (Belman, 1957). These results support the concept of an interaction between allergen and a skin constituent as being essential for the transformation of the
simple chemical into the sensitising antigen. Moreover, when this interaction
does not take place, a state of immune tolerance can be readily induced, as for
instance the one described above after excision of the allergenic depots or even
more after intravenous injection (Macher and Chase, 1969b) or feeding (Chase,
1946).

A central failure, i.e., at the level of the lymphocytes, has been shown (by
adoptive immunisation by cell transfer) in work on tolerance to allergens in-
duced by feeding (Battisto and Chase, 1955, 1963). The hypothesis that direct
access of antigen to lymphoid tissue induces tolerance, whereas indirect access
via macrophages induces immunity, would explain the present results. How-
ever, one must bear in mind that tolerance has been elicited with a simple hapten.
According to this hypothesis, the immune paralysis observed after the IP
injection of beryllium (table II) should result from the direct contact of beryllium
with the lymphoid system. This is supported by the observation that the tolero-
genic action of beryllium is suppressed by its incorporation in complete Freund
adjuvant or by its injection in animals pre-treated with glycogen, which is
known to stimulate a macrophage proliferation (table IV). The suppression of the
tolerogenic action is due to the phagocytosis of the beryllium particles, which
cannot now reach the lymphocytes. The protective effect of phagocytosis is
also suggested when stimulation of the RE system was produced incidentally
by infection following excision of the injection sites (table VII). In this experi-
ment the possibility of a binding between skin constituents and circulating
beryllium with the subsequent formation of an antigenic material is probable,
because the hapten cannot by itself induce an immunisation even after phago-
cytosis.

These results suggest that after administration beryllium is taken up by
various organs, the uptake depending on its physical (particle size) and chemical
form. These events can lead to different immunological situations.

Beryllium can be bound to proteins to form an antigenic material. As far
as the delayed type of hypersensitivity is concerned, this antigenic complex can
be formed only with skin proteins. Some protein binding will occur after IV
injection (Vacher and Stoner, 1968a) and probably after IP injection these com-
plexes of beryllium do not induce hypersensitivity; no humoral form of immunity
(in terms of antibodies) has been found (unpublished results) by the use of Farr’s
method (Farr, 1958), though this, of course, cannot be regarded as proved.

Beryllium can also be, in part, freely diffusible. After IV injection of BeSO₄ in
rats, this small-particle-size form has been shown to be phagocytosed much more
slowly than the big aggregates that are also formed (Vacher and Stoner, 1968b);
the diffusibility of the salt has been thought to be determined by the associated
organic acid, such as citrate (Feldman, Havill and Neuman, 1953). This is con-
firmed in our experiment on elimination from skin, in which it has been shown that
the diffusible part is much more evident after the ID injection of the citrate than
of the fluoride or sulphate. This diffusible fraction was associated with the tolero-
genic action, which could be explained by a direct interaction of the hapten
with the virgin lymphoid cells. As soon as the process of sensitisation is induced
by the antigenic material resulting from the interaction of beryllium with a
specific skin protein, tolerogenic activity can be elicited only by large toxic doses.

The final immune response, therefore, depends on the relative proportions of these fractions of beryllium, which vary according to the route of administration.

Some other factors must also be involved, such as a difference in the distribution and the rate of elimination of beryllium. It is indeed surprising not to induce the same degree of immune paralysis by pre-treatment with IV injection as with IP, and not to demonstrate tolerance after excision of injection sites after ID injection of the citrate. This could possibly be explained by rapid excretion of the diffusible, tolerogenic, fraction of beryllium. The role of slow release from the deposition sites in organs in the maintenance of the tolerance initiated by IP injection has also to be considered.

For the reasons discussed, it is difficult to calculate the actual amount of beryllium responsible for the production of immune paralysis. However, two dose ranges have been described; one very low, since as little as 5 μg Be per kg IP induces a partial paralysis, which becomes almost complete 56 days after 38.3 μg (table II); the other very high, as when toxic doses of BeSO₄ (this paper) or beryllium lactate (Polák and Turk, 1968) are injected IV. This seems to fit the model described by Mitchison (1964, 1968) for the tolerance induced in adult animals with a protein antigen. In this study, it was not possible to demonstrate the intermediate zone of immunisation since beryllium is administered in a non-antigenic form, and the intimate mechanism of the immune paralysis remains unknown. It could perhaps be linked with the adjuvant property of beryllium recently described (Unanue, Askonas and Allison, 1969), and possibly explained by a cell division of the immunologically competent cells, which would then become insensitive to a further antigenic stimulus.

SUMMARY

In adult guinea-pigs, beryllium induces either a skin sensitisation or an immunological paralysis.

Skin delayed hypersensitivity results only from the contact of beryllium with the skin. Parenteral administration fails to sensitise, but may induce tolerance. These immune responses are associated with different forms of beryllium: immunogenic (bound to skin constituents) and tolerogenic (freely diffusible). The relative proportion of these fractions depends on the route of administration and on the salt of beryllium. The tolerant state is achieved either by intraperitoneal injection of a very low dose of beryllium (4.78 μg Be per kg intraperitoneally) or by intravenous injection of high toxic doses (400 μg Be per kg); this fits with the model described by Mitchison (1968) for protein antigens.

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