MODELS OF INFECTION

An ascending non-obstructive model for chronic pyelonephritis in BALB/c mice

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Summary. Chronic pyelonephritis was successfully produced in female BALB/c mice with *Escherichia coli* after introduction of the bacterial inoculum by the ascending route. *E. coli* strain 31-B, a nalidixic acid-resistant derivative of strain TN675 expressing type-1 pili, and *E. coli* strain BH-5, a mutant of 31-B, not expressing type-1 pili, were used in the model. Both the strains were able to induce chronic renal inflammation in the experimental animals. Whereas the initial colonisation was greater with strain 31-B, its non-pilate mutant BH-5 caused a significantly greater inflammatory response and also caused renal scarring observable 5 months after the infection.

Introduction

The most common form of extra-intestinal infection caused by *Escherichia coli* is urinary tract infection, c. 80% of which is caused by this micro-organism.1 Human infections occur most commonly in patients with anatomically and functionally normal urinary tracts and involve spontaneous ascent of bacteria from the urethra to the kidney through the bladder, or occasionally infection arises via the blood stream.2–4 Epidemiological studies can identify an association between certain bacterial properties and virulence but understanding the pathogenesis of the disease requires the use of animal models. These animal models preferably should not involve manipulation of the urinary tract.5

The animal species used must share with man those aspects of the urinary tract that are important in the pathogenesis of the disease. Experimental pyelonephritis is relatively easy to establish in rats because of spontaneous vesico-ureteral reflux (VUR).5 However, rat uro-epithelial cells do not match human cells in the expression of receptors for p-fimbriae.5,7 Furthermore, the globoside isolated from the rat kidney contains a Galz (1–3) Galβ linkage instead of the critical Galz (1–4) Galβ linkage present in human globoside.8 Thus the rat is of questionable value for use in evaluating the pathogenesis of pyelonephritis. The rat model available for chronic pyelonephritis employs direct inoculation of organisms in the kidney.9

In contrast to rat tissue, mouse renal tissue contains an appreciable concentration of Galz (1–4) Galβ-containing glycolipid.10 An ascending model for acute pyelonephritis is available both in the rat and the mouse.11–14 A model employing the ascending route of infection for developing chronic pyelonephritis has not yet been standardised in either animal species. In the present study, a model for chronic pyelonephritis in BALB/c mice was devised in which an ascending route of infection was employed.

Materials and methods

Bacterial strains

Two strains of *Escherichia coli* were used for the study. (i) Strain 31-B, a nalidixic acid-resistant derivative of strain TN 675. This strain produces mannose-sensitive type-1 pili and is uropathogenic. (ii) Strain BH-5, a mutant defective in the ability to produce type-1 pili. Both these strains were kindly supplied by Takeda Chemical Industries Ltd, Japan.

Bacterial cultivation

*E. coli* strains were maintained on nutrient agar slants. The strains were grown in static culture in Brain Heart Infusion Broth (BHI; Hi-media, Bombay, India) at 37°C for 18 h. The bacteria were harvested by centrifugation at 500 g for 30 min, washed three times and suspended in phosphate-buffered saline (PBS, pH 7.3) to the desired concentration of 2 × 10⁹ cfu/ml.
**Animals**

Female BALB/c mice (Central Animal House, Punjab University, Chandigarh, India) weighing 20–30 g were used for the study. They were fed on standard antibiotic-free synthetic feed (JDB Agencies Private Ltd, India) Mice without bacteriuria, as confirmed by culture, were selected for the study.

**Induction of chronic pyelonephritis**

The original method for inducing acute pyelonephritis of Hagberg et al. and Schaeffer et al. was used with the slight modifications outlined by Sinha et al. for the introduction of bacterial inoculum by the ascending route. A soft intermediate non-radio-opaque polyethylene tubing with an outer diameter of 0·61 mm (Clay Adams, USA) was inserted into the urethral meatus and advanced into the bladder. The inoculum (0·1 ml) was injected slowly into the bladder to avoid leakage and the catheter was withdrawn carefully. To achieve persistent infection, these animals were re-infected 15 days after the first infection. To achieve renal scarring, animals were again infected 16 weeks later.

Groups of six mice, infected with either strain of *E. coli*, were killed by cervical dislocation 4 weeks, 8 weeks, 12 weeks, 16 weeks and 20 weeks later. The viscera of the mice were exposed after thoroughly cleaning the abdomen with alcohol. The kidneys were removed aseptically and were examined bacteriologically and histopathologically. Blood cultures were done to exclude bacteraemia.

**Bacteriological examination**

One-half of each kidney was placed in a sterile tube and weighed. The kidney tissue was homogenised in a Corning glass homogeniser with 1·0 ml of sterile PBS. Serial dilutions of the homogenised mass in PBS were plated on MacConkey agar plates. The plates were incubated at 37°C for 24 h and the viable counts were determined.

**Histopathological examination**

The kidney tissue, preserved in formalin, was dehydrated in an ethanol gradient of 70–100%. The tissue was embedded in paraffin, sectioned and stained with haematoxylin and eosin.

**Histopathological grading**

Grading of the severity of pathological lesion was done according to the method of Garg et al.

**Results**

Chronic pyelonephritis was induced in BALB/c mice with both *E. coli* strains 31-B and BH-5. The bacteriological studies (table, fig. 1) revealed that the pilate *E. coli* strain 31-B was able to colonise BALB/c mouse renal tissue much better than its non-pilate mutant, BH-5, (p < 0·05) when assessed 4 weeks after infection.

Eight weeks after infection a significant decrease in the colonisation by both strains was observed (p < 0·05) when compared with colonisation at 4 weeks, but the pilate strain still showed higher bacterial counts than the non-pilate mutant. However, this difference was not significant (p > 0·01). However, at 12 weeks the non-pilate mutant showed a significant rise in

![Graph](https://via.placeholder.com/150)

**Fig. 1.** Renal bacterial load at time intervals after infection. *, time of intra-urethral inoculation; – , *E. coli* 31-B; – ● , *E. coli* BH-5.

**Table.** Comparison of mouse kidney bacterial load of *E. coli* 31-B and of *E. coli* BH-5 at different time intervals after the first infection

<table>
<thead>
<tr>
<th>Group of mice</th>
<th>Viable count of bacteria in kidney after</th>
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<tr>
<td></td>
<td>4 weeks</td>
</tr>
<tr>
<td>Group I</td>
<td></td>
</tr>
<tr>
<td>(E. coli 31-B)</td>
<td>7·918 (0·639)</td>
</tr>
<tr>
<td>Group II</td>
<td>6·235 (0·235)**</td>
</tr>
</tbody>
</table>

Values are expressed as mean (SD) log_{10} cfu/g of kidney tissue. p values are derived from Student’s *t* test. Group I versus Group II *p* < 0·05, **p < 0·01; value compared to that of immediate previous time interval † † p < 0·05, † † † p < 0·001.
numbers, whereas the pilate strain did not. At 16 weeks after infection, the kidneys were all found to be sterile, irrespective of the strain involved.

Histopathological studies (fig. 2) showed that, despite the fact that the non-pilate strain was less effective at colonisation, it was still able to cause more renal parenchymal damage and ultimately induced scarring 5 months after infection (fig. 3).

Discussion

A pre-requisite for a valid animal model of human pyelonephritis is that the uro-epithelial surface should exhibit, qualitatively and quantitatively, the same adhesive receptors for the invading organism as does that of man. In both man and BALB/c mice, receptors for type-1 pili have been detected on the epithelial surface of vagina, bladder, ureter, renal pelvis, proximal tubules and collecting ducts, but they have not been detected in the glomeruli. The carbohydrates of type-1 pili have been detected in the urine of man and BALB/c mice. The urine of both species also contains a titratable factor that inhibits binding of type-1 pili to their receptors. This factor was determined to be physically, chemically and immunologically identical to the Tamm-Horsfall (T-H) glycoprotein and is produced by the ascending limb of the loop of Henle and distal convoluted tubules. It is secreted into the tubular urine and represents urinary slime. This material may entrap type-1 pilate E. coli strains and nullify their capacity to bind to cell-surface receptors. This study revealed that the pilate E. coli strain (31-B) was able to colonise BALB/c mouse renal tissue much better than its non-pilate mutant at 4 weeks after infection (p < 0.01). Iwahi and Imada also observed that the non-pilate strain (BH-5) did not readily colonise the urinary tract of mice. This may be an example of the principle that colonisation of a mucosal surface is a function of the relative molar
concentration in situ of cell-bound and free receptors and components like T-H proteins which are produced in the kidney and excreted in urine. Furthermore, the non-piluate mutant is more prone to the lysosomal vacuoles as long as they are non-opsonised, possibly because of the low levels of the myeloperoxidase activity in lysosomes containing non-opsonised bacteria.

A study of the pathological changes reveals that in spite of the fact that the non-pilate strain was less effective at colonisation, it was still able to cause more renal parenchymal damage at all stages of infection and ultimately was able to induce scarring after 5 months in 60% of the animals. The contribution of excessive release of reactive oxygen metabolites (ROS) from PMNLs bringing about adventitious damage of surrounding tissue needs to be studied. The suggestion has been made that if the bacteria remain extra-cellularly attached and continue triggering the release of ROS and lysosomal enzymes without being ingested, this may potentiate the inflammatory process.  

Thus, chronic pyelonephritis can be induced in BALB/c mice by employing an ascending route of infection and this model can be used to study the pathogenesis of the disease process.

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References