Emergence of carp edema virus in cultured ornamental koi carp, Cyprinus carpio koi, in India

T. Raja Swaminathan,1 Raj Kumar,1 Arathi Dharmaratnam,1 V. S. Basheer,1 Neeraj Sood,2 P. K. Pradhan,2 N. K. Sanil,3 P. Vijayagopal3 and J. K. Jena4

1Peninsular and Marine Fish Genetic Resources Centre of National Bureau of Fish Genetic Resources, Indian Council of Agricultural Research, Central Marine Fisheries Research Institute Campus, Kochi 682018, India
2National Bureau of Fish Genetic Resources, Indian Council of Agricultural Research, Lucknow 226002, India
3Central Marine Fisheries Research Institute, Indian Council of Agricultural Research, Kochi 682018, India
4Division of Fisheries Science, Indian Council of Agricultural Research, Krishi Anusandhan Bhawan-II, Pusa, New Delhi 110012, India

A disease outbreak was reported in adult koi, Cyprinus carpio koi, from a fish farm in Kerala, India, during June 2015. The clinical signs were observed only in recently introduced adult koi, and an existing population of fish did not show any clinical signs or mortality. Microscopic examination of wet mounts from the gills of affected koi revealed minor infestation of Dactylogyrus sp. in a few koi. In bacteriological studies, only opportunistic bacteria were isolated from the gills of affected fish. The histopathological examination of the affected fish revealed necrotic changes in gills and, importantly, virus particles were demonstrated in cytoplasm of gill epithelial cells in transmission electron microscopy. The tissue samples from affected koi were negative for common viruses reported from koi viz. cyprinid herpesvirus 3, spring viraemia of carp virus, koi ranavirus and red sea bream iridovirus in PCR screening. However, gill tissue from affected koi carp was positive for carp edema virus (CEV) in the first step of nested PCR, and sequencing of PCR amplicons confirmed infection with CEV. No cytopathic effect was observed in six fish cell lines following inoculation of filtered tissue homogenate prepared from gills of affected fish. In bioassay, the symptoms could be reproduced by inoculation of naive koi with filtrate from gill tissue homogenate of CEV-positive fish. Subsequently, screening of koi showing clinical signs similar to koi sleepy disease from different locations revealed that CEV infection was widespread. To our knowledge, this is the first report of infection with CEV in koi from India.

INTRODUCTION

In the international ornamental fish trade, more than 90% of the freshwater fishes (by volume) are farm bred, and about 4000 species of freshwater ornamental fishes are traded every year (Whittington & Chong, 2007). Goldfish (Carassius auratus) and koi carp (Cyprinus carpio koi) are the most extensively traded fishes species (Ariel, 2005). Importantly, the global ornamental fish trade relocates large quantities of live fish species between countries, and therefore can be a potential source for spread of exotic pathogens, particularly viral pathogens, which have been associated with high morbidity and mortality in fishes, in the importing countries (Oyamatsu et al., 1997a; Amita et al., 2002).

Some of the important viral diseases affecting koi carp, Cyprinus carpio koi, are caused by infection with cyprinid herpesvirus 3 (CyHV-3, also known as koi herpesvirus), spring viraemia of carp virus (SVCV), ranavirus (George et al., 2015) and carp edema virus (CEV) (Oyamatsu et al., 1997a). CEV is an unclassified virus belonging to the family Poxviridae and has two main lineages: lineage 1 (CEV) from koi carp which is more similar to original Japanese CEV and lineage 2 (CEV-like) from common carp (Way & Stone, 2013).

The GenBank/EMBL/DDBJ accession numbers for the new sequences reported in this paper are KX503806 and KX503807 (will be released to public database after July 2017).
Infection with CEV, also known as koi sleepy disease (KSD), was originally described from koi carp in Japan in 1974 (Murakami et al., 1976) and has recently been reported from many European countries including UK, France (Way & Stone, 2013), The Netherlands (Haenen et al., 2013), Germany (Jung-Schroers et al., 2015), Italy (Pretto et al., 2015), Czech Republic (Vesely et al., 2015) and Austria (Lewisch et al., 2015). Recently, a highly specific and sensitive quantitative PCR protocol was developed to improve CEV detection in koi by Adamek et al. (2016). Importantly, there is no report of KSD from India, till date.

The disease is characterized by typical sleepy behaviour, enophthalmia, generalized oedematous condition and gill necrosis (Lewisch et al., 2015). The affected juvenile carp usually congregate near the surface of pond or water inlet, whereas the older fish tend to lie on the bottom of the pond and eventually die of anoxia (Miyazaki et al., 2005) and the mortality may reach 80–100 %. The koi carp culture in India is growing vigorously and it is localized in selected states viz. Kerala, Tamil Nadu, West Bengal and Mahara-shtra. In the present study, we report an outbreak in koi carp in Ernakulam District, Kerala, India, resulting in 100 % mortality and, to our knowledge, this is the first report of infection with CEV from India. In subsequent studies, the infection appears to be quite widespread in koi in the country.

RESULTS AND DISCUSSION

The disease outbreak was reported in an ornamental fish farm in the month of June 2015 which is known to be a rainy season in Kerala, India. The affected fish exhibited lethargic behaviour and were seen lying at the bottom of the pond. The other clinical signs included haemorrhages on the surface, sunken eyes and swelling of body and gills (Fig. 1). Importantly, these clinical signs were observed only in introduced adult koi, 3 days after stocking, and the number of fish displaying these signs increased daily. The mortality started on the fifth day after stocking and 100 % mortality (a total of 800 fish) was reported within 8 days of stocking. The water temperature of the affected pond was 18–20 °C.

The clinical signs viz. anorexia, lethargy, skin erosions, enophthalmia, and gill necrosis, as observed in the present outbreak, have been reported earlier as well (Miyazaki et al., 2005; Haenen et al., 2013; Way & Stone, 2013; Lewisch et al., 2015). Interestingly, the disease was not observed in existing population of koi (250 fish), goldfish (300 fish) and Indian major carps (IMCs; Labeo rohita and Catla catla) (500 fish) reared in the same pond. Previously, Jung-Schroers et al. (2015) observed that an existing population of koi in a German farm was affected by the disease, but the disease was not observed in goldfish and sturgeon kept in the same farm. In conformity with the findings of the present study, very high mortality ranging from 80 to 100 % has also been reported previously in affected koi (Oyamatsu et al., 1997a; Miyazaki et al., 2005). Similar to the observation of the present study, CEV outbreak in koi ponds has been reported at 15–25 °C (Oyamatsu et al., 1997a; Amita et al., 2002). It has been reported that water temperature and acute stress conditions (restocking, transport, etc.) play a crucial role in the onset of disease outbreaks in fish populations (Miyazaki et al., 2005; Magnadottir, 2010). Therefore, it can be assumed that stress of transportation could have precipitated KSD outbreak in adult koi in the present outbreak. In accordance with our findings, Lewisch et al. (2015) reported that restocking is an important predisposing factor responsible for KSD outbreaks (Lewisch et al., 2015). Similarly, Jung-Schroers et al. (2015) also reported that KSD outbreaks are generally observed when koi are moved from earthen ponds to cemented tanks. In the outbreak reported here, the lack of transmission of disease to the previously resident fish might be due to lack of handling stress in those fish or, possibly, immunity due to previous sublethal exposure to the virus.

Microscopic examination of wet mounts from the gills revealed minor infestation of Dactylogyrus sp. in a few koi. Furthermore, bacteria viz. Aeromonas sobria, Klebsiella pneumoniae, Proteus penneri, Shewanella decolorationis and Escherichia coli were isolated from the gills of affected fish. However, bacteria could not be isolated from internal organs viz. spleen and kidney of affected fish. Since the parasites and bacteria were not observed consistently in the diseased koi, therefore, it was presumed that the aetiological agent of disease might be other than parasites and bacteria.

The histopathological examination of the affected fish revealed severe necrosis of gills, proliferation of the epithelial cells and adhesion of gill lamellae. In addition, infiltration of eosinophilic granular cells was observed between the secondary gill lamellae along with large number of monocellular cells and a few erythrocytes (Fig. 2a). In the kidney,
Degenerative changes were observed in tubular epithelial cells (Fig. 2b). The histopathological alterations, as observed in the present case, are consistent with previous reports of KSD outbreaks (Ono et al., 1986; Miyazaki et al., 2005; Jung-Schroers et al., 2015; Lewisch et al., 2015). As the virus is reported to replicate in gill epithelial cells (Ono et al., 1986; Miyazaki et al., 2005), this would have resulted in severe histopathological alterations in the gills. However, atrophy of splenic pulp as reported by Miyazaki et al. (2005) was not observed in affected koi. In a previous study, multifocal necrotic areas in hepatopancreas and myocarditis due to infection with CEV have been reported (Lewisch et al., 2015). However, in the present study, these tissues were not examined for histopathological alterations. It has been suggested previously that cellular changes in organs other than gills are due to effect of hypoxia caused by severe damage to gills (Lewisch et al., 2015).

Importantly, virus particles were demonstrated in cytoplasm of gill epithelial cells of affected koi carp and these were spherical and electron dense in transmission electron microscopy (TEM) analysis (Fig. 3). However, no virus particles were observed in spleen and kidney tissue sections of the infected koi carp. These findings are consistent with previous reports (Ono et al., 1986; Oyamatsu et al., 1997b; Miyazaki et al., 2005; Jung-Schroers et al., 2015). In contrast, Way & Stone (2013) failed to demonstrate virus particles in gill tissue from diseased carp. In accordance with our results, the virus particles have not been demonstrated in tissues other than gills (Oyamatsu et al., 1997a; Miyazaki et al., 2005).

PCR screening of the tissue samples (gills, spleen and kidney) from affected koi revealed that these were negative for CyHV-3, SVCV, koi ranavirus and red sea bream iridovirus. However, gill tissue (37 samples) from affected koi carp was positive for CEV in nested PCR developed by Oyamatsu et al. (1997b). Interestingly, other co-habitating fishes viz. resident koi carp, goldfish and IMCs collected from the affected farm were not positive even in the second-step PCR (five fish tested for each species). The sequence analysis of 548 bp fragment of randomly selected 10 PCR amplicons from CEV-positive fish revealed greater than 96 and 98 % sequence identity with the original sequences submitted by Oyamatsu et al. (1997b) and Jung-Schroers et al. (2015) (accession number KM283182), respectively. This confirmed the identity of CEV as the probable cause of the mortality in affected koi. The sequences have been submitted to NCBI GenBank (accession numbers KX503806 and KX503807). Way & Stone (2013) also reported that amplification products from the majority of CEV-positive cases of koi in UK share 97.5–99.5 % nucleotide sequence identity with the original Japanese CEV sequence. Further, all 37 fish samples were re-tested with a second nested PCR assay (recent assay better at detecting CEV in carp) targeting 478 bp partial 4a gene sequence from CEV. The sequence analysis of 10 partial 4a gene sequences (analysed in the previous PCR) revealed 95 and 99 % identity with sequence of CEV isolated from common carp (KX254010, KX254011, KX254019 and KX254020) and koi carp (KX254000, KX254001, KX254021, KX254022 and KX254024).

**Fig. 2.** Histopathological studies of gills from CEV-affected koi carp. (a) Extensive infiltration of mononuclear cells (arrows) with a few eosinophilic granular cells (arrowheads) and erythrocytes in the interlamellar region of affected gill tissue. Bar, 20 µm. (b) Section of kidney showing degenerative changes (arrows) in tubular epithelial cells. Bar, 20 µm.

**Fig. 3.** Transmission electron photomicrograph of a koi gill epithelial cell containing large intracytoplasmic spheroid CEV virions (arrows). Bar, 500 nm.
respectively. No substitutions of the nucleotide were found among all 12 Indian CEV sequences and showed 100% sequence identity. The sequences have been submitted to NCBI GenBank. Phylogenetic analysis of 478 bp fragment of 4a gene revealed two distinct clusters viz. CEV sequences from koi carp and common carp and suggested that there was a high relatedness between the Indian koi carp CEV sequence and koi carp CEV sequences reported in UK, Poland and Japan (Fig. 4). Thus, the koi carp CEV identified in India had highest identity with koi carp CEV detected in UK, Poland and Japan while no amplification was derived from liver and kidney.

No cytopathic effect was observed in six cell lines following inoculation of filtrate from gill tissue homogenate of CEV-positive koi, after 15 days of inoculation and even after five blind passages. The cell lines tested were derived from koi, goldfish and four other ornamental fish species cultured in India (Swaminathan et al., 2010, 2013, 2015, 2016a, b). The results are in accordance with earlier report of Oyamatsu et al. (1997a) who were not able to isolate virus using 31 cell lines including 9 cyprinid, 8 salmonid, 4 anguillid and 10 other teleostean cell lines inoculated with filtered

Fig. 4. Unrooted neighbour-joining tree based on 478 bp partial 4a gene sequence of CEV. Except the Indian CEV sequences, all other CEV sequences of common carp and koi carp were obtained from NCBI. The scale bar represents substitutions per nucleotide site.
homogenate of gills. Similarly, Jung-Schroers et al. (2015) did not observe any cytopathic effect following inoculation of filtrate from pooled tissues (gills, kidney, gut and brain) in fathead minnow and common carp brain cell lines. Therefore, it is imperative to develop in vitro cultivation methods to better understand host–pathogen interaction.

In bioassay, koi inoculated with filtrate from gill tissue homogenate of CEV-positive fish showed clinical signs 4 days post-inoculation (dpi) onwards and infected fish began to die from 6 dpi and during experimental period of 10 dpi; 5 out of 15 inoculated fish died. PCR of gill tissue of dead fish was positive for CEV in the first step of nested PCR. However, in the control group, there was neither morbidity nor mortality and all the tested fish were negative in PCR.

Survey was carried out to detect presence of CEV in koi exhibiting clinical signs similar to sleepy disease from other locations in India (Fig. 5). The samples were screened for the presence of CEV as per Matras et al. (2016). A total of 53 out of 320 koi carps (16.56 %) were positive for CEV in the first step, whereas additional 9 samples (2.8 %) were positive in the second-step PCR (Table 1). CEV was detected at nine locations during active surveillance of KSD/CEV. All the nine sequences generated from the survey of CEV in India were identical, suggesting high genetic similarity among Indian CEV. Previously, several cases of mortality in koi exhibiting gill necrosis were negative for CyHV-3 in PCR. In the context of present findings, it may be assumed that some of these cases could have been due to infection with CEV and would require testing to confirm in future studies. Importantly, CEV-like virus has been detected in clinically healthy koi carps imported from Israel and Japan to the UK (Way & Stone, 2013). So, trading apparently healthy koi may pose a threat of spreading CEV to unaffected regions, which emphasizes the need of precautionary approach to prevent its spread.

METHODS

Fish samples. During May 2015, a disease outbreak was reported from an ornamental fish farm in Ernakulam District, Kerala, India, with a cumulative mortality up to 100 % in adult koi carp (n=800; 40–42 cm

---

Fig. 5. Collection sites of koi carp (C. carpio koi) in the study for the survey of CEV.
Briefly, swabs from the gill and kidney of affected koi carps were plated from gills and kidney of the affected fish, using standard methods. Bacteriological investigation. Gill, spleen and kidney tissues were fixed in 10 % neutral buffered formalin and in 2.5 % glutaraldehyde (Sigma-Aldrich) respectively for bacterial and virus isolation. The tissues were also fixed in 10 % ethanol for molecular diagnosis. Histopathology. gill, spleen and kidney tissues were collected aseptically for ectoparasites. Thereafter, the fish were euthanized with MS222 on the second day of onset of mortality.

On arrival, the affected fish were observed for clinical signs and gross lesions. Fresh smears of gills and skin were examined under a microscope for ectoparasites. Thereafter, the fish were euthanized with MS222 (Sigma-Aldrich) and gill, spleen and kidney tissues were collected aseptically for bacterial and virus isolation. The tissues were also fixed in 10 % neutral buffered formalin and 2.5 % glutaraldehyde (Sigma-Aldrich) in phosphate buffer for histopathology and TEM, respectively. Gill, spleen and kidney tissue samples were also preserved separately in 99 % ethanol for molecular diagnosis.

Bacteriological investigation. Bacterial isolation was attempted from gills and kidney of the affected fish, using standard methods. Briefly, swabs from the gill and kidney of affected koi carps were plated onto brain–heart infusion agar (HiMedia) and the plates were incubated at 28 °C for 48 h. Pure bacterial colonies were biochemically characterized as per Bergey’s Manual of Determinative Bacteriology and also by 16S rRNA gene sequencing.

PCR and sequence analysis. Extraction of nucleic acid (RNA and DNA) was done by GeneJET RNA purification kit (Thermo Scientific) and DNAeasy Blood and Tissue kit (Qiagen), respectively, following the manufacturer’s instructions. The tissue samples viz. gills, spleen and kidney were pooled from individual fish for the isolation of nucleic acids. The cDNA synthesis was done using Verso cDNA kit (Thermo Scientific). Samples were analysed for the presence of nucleic acid sequences specific for SVCV (Stone et al., 2003), CyHV-3 (Bercovier et al., 2005), koi ranavirus (George et al., 2015) and red sea bream iridovirus (Kurita et al., 1998). Further, DNA samples were examined for the presence of CEV-specific DNA sequences (an undefined segment of the CEV genome) by PCR (548 bp in the first step and 181 bp in the second step) as described by Oyamatsu et al. (1997b) and another PCR protocol recently developed based on the partial 4a gene of CEV (528 bp in the first step and 478 bp in the second step) in the Centre for Environment, Fisheries and Aquaculture Science, Weymouth laboratory, UK (Matras et al., 2016), for further confirmation.

Isolation of CEV on different fish cell lines. The tissue homogenate for infecting the cell culture monolayer was prepared from pooled gill, spleen and kidney of CEV-positive koi carps, using the standard protocol. Briefly, pooled tissues were homogenized using sterile Leibovitz’s L-15 medium supplemented with 1000 IU ml⁻¹ penicillin, 1000 µg ml⁻¹ streptomycin and 2.5 µg ml⁻¹ amphotericin B (Life Technologies) without FBS. In order to release the virus particles from the infected cells, the homogenized samples were frozen and thawed alternately, thrice. The homogenate was centrifuged at 300 × g for 30 min at 4 °C and the supernatant was filtered through a 0.22 µm filter (Millipore). The filtrate (500 µl) was inoculated onto a confluent monolayer of the C. carpio koi fin cell line (Swaminathan et al., 2015) in a 25 cm² flask (Nunc) which was incubated at 28 °C. After adsorption for 1 h at room temperature, 5 ml of a maintenance medium (L-15 medium with 2 % FBS) was added. In control flasks, 500 µl of maintenance medium was used. Cells were monitored for any cytopathic effect daily under an inverted microscope (Nikon) for 15 dpi. Similarly, the filtrate was inoculated in fish cell lines that have been developed in our laboratory viz. pearl spot fin (Swaminathan et al., 2010), catopra fish fin (Swaminathan et al., 2013), Horabagrus brachysoma fin (Swaminathan et al., 2016a), C. carpio koi fin (Swaminathan et al., 2015), angelfish fin (Swaminathan et al., 2016b) and goldfish fin (unpublished data) for viral isolation. Blind passage of inoculated cell lines was carried out five times on all cell lines at an interval of 2 weeks. In addition, DNA was isolated from supernatant as well as cells of the inoculated cell lines and checked for CEV-specific sequences using PCR.

Histopathology. For histology, gill, spleen and kidney tissues fixed in 10 % neutral buffered formalin were dehydrated in ascending grades of ethanol, cleared in chloroform and embedded in paraffin wax. Tissue

Table 1. Detection of CEV in koi collected from different locations and exhibiting clinical signs similar to sleepy disease

<table>
<thead>
<tr>
<th>Sample no.</th>
<th>Sampling location</th>
<th>No. of samples tested</th>
<th>No. of samples positive in first-step PCR</th>
<th>Additional number of samples positive in second-step PCR</th>
<th>Remarks</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Thrissur</td>
<td>30</td>
<td>2</td>
<td>0</td>
<td>Samples were collected from a farm</td>
</tr>
<tr>
<td>2</td>
<td>Ernakulam</td>
<td>33</td>
<td>3</td>
<td>0</td>
<td>Samples were collected from a retail shop</td>
</tr>
<tr>
<td>3</td>
<td>Kottayam</td>
<td>28</td>
<td>2</td>
<td>0</td>
<td>Samples were collected from a farm</td>
</tr>
<tr>
<td>4</td>
<td>Kozhikode</td>
<td>24</td>
<td>4</td>
<td>1</td>
<td>Adult koi with symptoms were purchased and transported recently from Madurai, Tamil Nadu</td>
</tr>
<tr>
<td>5</td>
<td>Paravoor</td>
<td>26</td>
<td>0</td>
<td>0</td>
<td>Samples were collected from a farm</td>
</tr>
<tr>
<td>6</td>
<td>Chalakkudy</td>
<td>34</td>
<td>7</td>
<td>2</td>
<td>Adult koi with symptoms were purchased and transported recently from Chennai</td>
</tr>
<tr>
<td>7</td>
<td>Vaikom</td>
<td>36</td>
<td>3</td>
<td>0</td>
<td>Samples were collected from a retail shop</td>
</tr>
<tr>
<td>8</td>
<td>Chennai</td>
<td>38</td>
<td>12</td>
<td>2</td>
<td>Samples were collected when the adult koi were shifted from cemented tank to earthen ponds</td>
</tr>
<tr>
<td>9</td>
<td>Madurai</td>
<td>36</td>
<td>10</td>
<td>2</td>
<td>Samples were collected when the adult koi were shifted from cemented tank to earthen ponds</td>
</tr>
<tr>
<td>10</td>
<td>Kolkata</td>
<td>35</td>
<td>10</td>
<td>2</td>
<td>Samples were collected when the adult koi were shifted from cemented tank to earthen ponds</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td>320</td>
<td>53</td>
<td>9</td>
<td></td>
</tr>
</tbody>
</table>
sections of 5 μm thickness were cut using a rotary microtome, stained with haematoxylin and eosin and examined under the light microscope.

Electron microscopy. The fixed tissues were processed as per the standard methods for ultra-thin sections for TEM analysis with modifications. Briefly, the infected gill, spleen and kidney tissues fixed with 2.5% glutaraldehyde were post-fixed in 2.0% osmium tetroxide prepared in 0.1 M phosphate buffer for 1 h at 4°C. The tissues were dehydrated in ascending grades of ethanol and were embedded in Araldite CY212. Ultra-thin sections (60–70 nm) of tissues were cut using a microtome (Leica Ultracut UCT), mounted onto copper grids and stained with uranyl acetate and alkaline lead citrate. Sections were examined and documented under a Tecnai T12 Spirit transmission electron microscope at 60 kV.

Experimental infection. Apparently healthy koi (n=30, 20–26 g) were procured from local ornamental fish shops and acclimatized in glass aquaria for 10 days, and subsequently, 15 koi were injected intra-peritoneally with 0.5 ml filtrate of pooled gill homogenate from CEV-positive koi and kept in three different aquaria with five fish each as the infected group. Another group of 15 carp was similarly injected with 0.5 ml of 1:15 medium and constituted the control group. The fish were observed daily for development of clinical signs as well as mortality for a period of 10 days. During the experiment, temperature, dissolved oxygen, pH, nitrite and ammonia were 19.0±1.4°C, 6.2±0.81 mg l⁻¹, 7.9±0.56, 0.012±0.009 mg l⁻¹ and 0.11±0.27 mg l⁻¹, respectively. Moribund or dead fish in the infected group and healthy fish in the control group were randomly collected for the PCR in order to detect CEV.

Survey of CEV. A total of 320 koi carps (12–15 cm total length with 25–30 g total body weight) were collected from 12 different locations, from either farms or retail shops in India during the last 1 year. The samples of gills from these fish were screened for CEV using second PCR assay described by Matras et al. (2016).

Phylogenetic analysis of the Indian CEV sequences. The PCR products amplified from gill samples collected from the disease outbreak cases as well as from different locations during survey were cleaned and sequenced in Applied Biosystems 3730 XL capillary sequencer at a private sequencing facility in Kochi, Kerala. The raw DNA sequences were aligned and edited using BioEdit sequence alignment editor version 7.0.5.2. A neighbour-joining tree was also constructed using the partial 4a gene of Indian CEV sequences generated from the disease outbreak (three sequences) and from survey (nine sequences) together with corresponding sequences of CEV sequences obtained from both koi and common carp from the UK, Poland and Japan by the Kimura two-parameter method at 1000 bootstrap replications using MEGA 4.0 (Tamura et al., 2007).

ACKNOWLEDGEMENTS

The authors are thankful to the Secretary, DARE, and Director General, Indian Council of Agricultural Research, New Delhi, India, for their support, encouragement and guidance. We thank Dr B. Gorgoglione, University of Veterinary Medicine, Austria, for providing the positive DNA of CEV to be used as a positive control in the PCR detection of CEV in our study. We are indebted to Dr David Stone, Centre for Environment, Fisheries and Aquaculture Science, UK, for sharing the primer details developed by him to detect the CEV in common carp and for his critical suggestions to improve the manuscript. Authors thank anonymous reviewers for their valued observations, comments and suggestions, which improved the quality of the paper.

REFERENCES


