Short Communication

Protective humoral responses to severe acute respiratory syndrome-associated coronavirus: implications for the design of an effective protein-based vaccine

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Some of the structural proteins of severe acute respiratory syndrome-associated coronavirus (SARS-CoV) carry major epitopes involved in virus neutralization and are essential for the induction of protective humoral responses and the development of an effective vaccine. Rabbit antisera were prepared using full-length N and M proteins and eight expressed fragments covering the S protein. Antisera to S and M proteins were found to have different neutralizing titres towards SARS-CoV infection in vivo, ranging from 1:35 to 1:128. Antiserum to the N protein did not contain neutralizing antibodies. Epitopes inducing protective humoral responses to virus infection were located mainly in the M protein and a region spanning residues 13–877 of the S protein. The neutralizing ability of antisera directed against the expressed structural proteins was greater than that of convalescent patient antisera, confirming that, as immunogens, the former induce strong, SARS-CoV-specific neutralizing antibody responses. The in vitro neutralization assay has important implications for the design of an effective, protein-based vaccine preventing SARS-CoV infection.

Severe acute respiratory syndrome-associated coronavirus (SARS-CoV) is the causative agent of an atypical, highly transmissible respiratory disease that struck in 32 countries in 2003 resulting in approximately 800 deaths (Drosten et al., 2003; Lee et al., 2003; Poutanen et al., 2003; Rota et al., 2003). Development of a safe and effective vaccine to prevent SARS-CoV infection should be pursued with urgency as SARS is highly lethal and, if the virus does re-emerge, is likely to become endemic in regions of the world. SARS-CoV belongs to a diverse family of large, enveloped, single-stranded, positive-sense RNA viruses (Rota et al., 2003). The full-length viral genome, which also serves as an mRNA, may initiate a SARS-CoV infection in transfected cells (Yount et al., 2003). Therefore, the use of killed, recombinant or live-attenuated virus as a vaccine to immunize humans against SARS-CoV infection may pose serious risks (Martin et al., 1999). To evaluate the potential of expressed structural proteins of SARS-CoV for the development of a safe and effective vaccine against virus infection, we investigated the neutralizing ability of antibodies against SARS-CoV derived from sera of animals that had been immunized with the expressed structural proteins.

Eight truncated fragments covering the S protein (Fig. 1a) and the full-length N protein were expressed in Escherichia coli, using genes of the BJ01 strain (GenBank accession no. AY278488) of SARS-CoV isolated in China, while the M protein was expressed in Pichia pastoris as it could not be easily expressed in E. coli. All fragments of the S protein containing a six-histidine tag were expressed in E. coli as inclusion bodies. The proteins were purified by sonication and repeated washing, and then dissolved in Tris/HCl buffer with 8 M urea and further purified using Ni-NTA agarose (Invitrogen). Finally, the denatured proteins were refolded by dialysis in Tris/HCl buffer (pH 7.4). Soluble N and M proteins containing six-histidine tags were also purified using Ni-NTA agarose (Fig. 1b). It should be noted that, unlike our expression products, the native S protein is highly glycosylated and that oligosaccharides may affect the generation of neutralization antibodies. Consequently,
we cannot rule out the possibility that the lack of glycosylation in E. coli has influenced the outcome of our experiments. Nevertheless, expression of the proteins in E. coli could provide a simple and cheap method for large-scale production of vaccines protecting against SARS-CoV infection.

To generate antibodies to the purified proteins, we subcutaneously immunized 10 New Zealand white rabbits (male, 2–3 kg) each with one of the recombinant proteins. On day 0, each rabbit was vaccinated with 0.5 mg purified protein in Freund’s complete adjuvant. Animals received further vaccinations on days 14 and 21 with 0.5 mg of each protein in Freund’s incomplete adjuvant. Finally, the 10 rabbit sera were collected on day 28 after vaccination. Antibody titres were assessed by ELISA using the recombinant proteins. Using end-point dilution, high antibody titres were observed, ranging from 1:60,000 to 1:120,000 (Fig. 2a).

For neutralization tests, Vero E6 cells were grown in MEM medium supplemented with 10% fetal bovine serum (FBS) and antibiotics in 96-well plates. To assay each serum, 200 TCID$_{50}$ of virus was added at each step of a serial twofold dilution of heat-inactivated antisera (for 45 min at 56°C) in triplicate wells. After incubation for 1 h at 37°C, these mixtures containing 100 TCID$_{50}$ of virus were added to the rinsed Vero E6 monolayers. Using a microscope, the plates were monitored daily for the presence of cytopathic effect until day 4–5 after incubation. Rabbit pre-immune sera were included in each test as controls. In addition, dilutions of each serum without virus were added to control wells to assay for toxicity of the sera. The rabbit sera were highly toxic to Vero E6 cells, with all cells being killed at dilutions of 1:2 to 1:8 and approximately 20% of cells being killed at a 1:16 dilution. The neutralizing capacity of the sera directed against the SF fragment and the N protein could not be detected at dilutions below 1:16. Neutralizing titres were expressed as the reciprocal of the highest serum.
dilution that gave full protection against the standard virus dose used in these experiments. The neutralizing capacity of the antisera to each protein was characterized and validated in vitro and the results of the neutralization assay are shown in Fig. 2(b) as log_{10} values. The neutralizing capacity of antibodies to SA (at a 1:89 serum dilution), SB (1:79), SC (1:79) and SG1 (1:89) derived from the S1 domain of S protein was higher than that of SG3 (1:35) and SE (1:35) from the S2 region. No neutralization could be detected by antibodies to fragment SF from the S2 region, suggesting that the S1 domain of the S protein may be the major target of neutralizing antibodies preventing SARS-CoV infection. A recent report showed that an adenovirus-based vaccine with the SARS-CoV S1 fragment could induce strong neutralizing antibody responses to SARS-CoV infection in monkeys (Gao et al., 2003). Generally, a fragment with a domain (ligand) in the envelope protein that can bind to viral receptors on target cells may induce a higher neutralizing antibody titre than other fragments that do not contain such a ligand. Our SB fragment (residues 232–508) overlapped a 193 aa fragment of the S protein (residues 318–510) that has been shown to bind to angiotensin-converting enzyme 2, a functional receptor of SARS-CoV (Li et al., 2003; Wong et al., 2003). The neutralization titre of the antiserum to SB was almost equal to that of antisera to some domains of the S protein of SARS-CoV may influence receptor binding by a steric hindrance effect, as described for reoviruses (Nason et al., 2001). In addition to this steric hindrance effect, our observations suggested that SARS-CoV may also utilize more than one receptor for fusion with target cells during virus entry. It is known that human immunodeficiency virus type 1 (HIV-1) and HIV-2 require at least two receptors, CD4 and a member of the CXC-chemokines, to mediate virus entry into target cells (Endres et al., 1996; Oberlin et al., 1996; Trkola et al., 1996; Wu et al., 1996).

Notably, the neutralizing capacity of antiserum to SD (1:128), which overlapped both the SG1 and SG3 fragments, was higher than that of either SG1 or SG3

Fig. 2. Neutralization titres of antisera to the expressed structural proteins of SARS-CoV. (a) The titre of SARS-CoV-specific antibodies in the serum of immunized rabbits was determined and quantified by end-point dilution ELISA assay using the expressed proteins. Briefly, purified proteins (100 μl per well of 1 μg ml^{-1} solution, incubated overnight at 4 °C) were used to capture antibodies in the sera (incubated for 1 h at 37 °C), which were then detected with 100 μl horseradish peroxidase-conjugated goat anti-rabbit IgG (Jingmei Biotech) per well (diluted 1:5000 in PBS containing 0.5% Tween 20 and 10% FBS), followed by 100 μl o-phenylenediamine dihydrochloride solution (Sigma) per well for 30 min at room temperature in the dark. End-point titres were defined as the highest plasma dilution that resulted in an absorbance value (A_{450}) two times higher than that of non-immune plasma with a cut-off value of 0.05. Data are presented as log_{10} values. (b) Detection of virus-specific neutralizing antibodies in each serum by inhibition of Vero E6 cells lysis by SARS-CoV strain BJ01 in a micro-neutralization assay. Data are presented as the mean ± SE for four sets of tests each carried out in triplicate. (c) Evaluation of neutralizing capacity in pooled human convalescent sera. Equal volumes of antisera were mixed with several individual rabbit antisera raised using the fragments of the S protein, and their neutralizing capacity was determined under the same conditions described above. Data are presented as the mean ± SE.
individually (Fig. 1a and 2b). To confirm this synergistic effect of neutralizing capacity between antibodies to epitopes in SG1 and SG3, we mixed the two antisera in equal volumes to measure the neutralizing titre. The neutralizing capacity of the mixed antiserum against SG1 and SG3 was equal to that of SD (Fig. 2b and c). Neutralization of coronaviruses is a specific event that requires the presentation of the epitope involved in neutralization in the appropriate structural context. Work with transmissible gastroenteritis coronavirus (TGEV) has shown that the S glycoprotein domain recognized by the receptor on ST cells is located in the globular domain of the protein close to the antigenic sites A and D (Sune et al., 1990). In TGEV neutralization experiments using binary combinations of mAbs specific for different antigenic sites, a synergistic effect was also observed (Sune et al., 1990).

To investigate synergistic effects in more detail, we measured the neutralizing capacity of a mixture of antiserum against SA, SB and SD, which covered residues 13–877 of the S protein. As expected, the mixed sera showed a higher neutralizing titre of 1:342 (Fig. 2c). Similarly, the mixed sera to SA, SB and SC, covering residues 13–730 of the S protein, had a titre of 1:297. The mixture of antisera against SA, SB, SC and SG3, covering residues 13–882 of the S protein, had a titre of 1:342, which was the same as for the mixture of sera to SA, SB and SD (Fig. 2c). Generally, the humoral response against a virus is directed towards many different envelope protein epitopes. The results suggest that the region from residues 13–877 of the S protein, including an important epitope in fragment SG3 (residues 735–882), is essential for the development of an effective protein-based vaccine preventing SARS-CoV infection.

It has been reported that the M protein of TGEV is required for virus assembly and budding, that it is immunodominant and that M-specific antibodies both neutralize TGEV and mediate the complement-dependent lysis of TGEV-infected cells (Delmas et al., 1986; Risco et al., 1995; Woods et al., 1987). To investigate the neutralizing capacity of antiserum to the M protein of SARS-CoV, we examined its neutralizing titre using serum of heat-inactivated complement. The neutralization titre of the M protein antiserum was higher than 1:128 (Fig. 2b), suggesting that the M protein is important in development of an effective protein-based vaccine preventing SARS-CoV infection. To evaluate the synergistic effect between antisera to different epitopes, the M antiserum was mixed with antisera against the SA, SB and SD fragments (covering residues 13–877 of the S protein) and a neutralization assay was performed. The neutralizing capacity of the mixed SA + SB + SD + M antiserum showed a considerable synergistic effect, with a neutralizing titre of 1:396 (Fig. 2c). This suggested that sera to the M and S proteins can effectively enhance each other’s activity.

We also assayed the neutralizing capacity of antiserum to the N protein, but no neutralizing ability was detected in vitro (Fig. 2c). However, recent evidence indicates that the N protein can induce a T-cell response to SARS-CoV infection (Gao et al., 2003). An effective vaccine may also aim to elicit a vigorous cellular immunity. Therefore, the N protein is also important in development of an effective vaccine preventing SARS-CoV infection.

Neutralizing antibodies that bind efficiently to the envelope spikes of SARS-CoV can offer protection or may be beneficial if present at appropriate concentrations before exposure to the virus. To define the neutralizing capacity of convalescent antisera from SARS-CoV patients, we determined the neutralization titres for sera from three convalescent patients under the same experimental conditions, by pooling together equal volumes of antisera from each patient. The resulting neutralization titre was found to be 1:342. Clearly, the neutralization titre of mixed antisera to fragments SA, SB and SD (covering residues 13–877 of the S protein) and the M protein was higher than that of antisera from convalescent patients (Fig. 2c).

Our results have confirmed that, as major antigens, the expressed unglycosylated structural proteins can induce a strong, SARS-CoV-specific, neutralizing antibody response in rabbits. These results provide crucial information for a rational approach towards maximizing antibody responses elicited by potential vaccines. It will also be essential to determine whether humans and animals generate similar responses to SARS-CoV (Cyranoski, 2004; Fouchier et al., 2003; Guan et al., 2003; Martina et al., 2003). Virus-challenge tests in animal models will be required for different SARS-CoV isolates to assess the protective capacity of the expressed proteins as vaccines. Furthermore, before any potential vaccine can be tested in humans, a non-human primate challenge model must be used to evaluate its effectiveness against SARS-CoV infection and to exclude any potential side effects (Fouchier et al., 2003), such as those observed for candidate vaccines against feline infectious peritonitis virus (Vennema et al., 1990). In addition, the development of a vaccine against SARS-CoV infection in animals also needs to prevent the virus being carried and transmitted to humans by animals, such as masked palm civet, ferret and cat, and causing infection in humans (Cyranoski, 2004; Guan et al., 2003; Martina et al., 2003).

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References


