Morphological and genomic characterization of the polydnavirus associated with the parasitoid wasp *Glyptapanteles indiensis* (Hymenoptera: Braconidae)

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*Glyptapanteles indiensis* polydnavirus (GiPDV) is essential for successful parasitization of the larval stage of the lepidopteran *Lymantria dispar* (gypsy moth) by the endoparasitic wasp *Glyptapanteles indiensis*. This virus has not been characterized previously. Ultrastructural studies of GiPDV showed that virions had a rod-like or rectangular form and each contained as many as ten nucleocapsids enclosed by a single unit membrane envelope. Field inversion gel electrophoresis (FIGE) analysis of the virus genomic DNA revealed that GiPDV had a segmented genome composed of 13 dsDNA segments, ranging in size from approximately 11 kb to more than 30 kb. Four genomic segments were present in higher molar concentration than the others. Further characterization of the GiPDV genome yielded several cDNA clones which derived from GiPDV-specific mRNAs, and Northern blot analysis confirmed expression of isolated cDNA clones in the parasitized host. Each was present on more than one GiPDV genomic DNA segment, suggesting the existence of related sequences among DNA segments. It has been proposed previously that in polydnavirus systems, genome segmentation, hypermolar ratio segments and segment nesting may function to increase the copy number of essential genes and to increase the levels of gene expression in the absence of virus replication. The present data support this notion and suggest that GiPDV morphology and genomic organization may be intrinsically linked to the function and evolutionary strategies of the virus.

**INTRODUCTION**

Polydnaviruses (PDVs) are complex insect viruses characterized by segmented double-stranded circular DNA genomes that vary in size and molar ratio (Stoltz *et al.*, 1984). Viral DNA is integrated into the wasp genome as provirus and transmitted vertically through the germ-line (Stoltz *et al.*, 1984, 1986; Fleming & Summers, 1986, 1991; Fleming, 1992; Fleming & Krell, 1993; Stoltz, 1993). PDVs replicate only in the nucleus of calyx epithelial cells of the wasp ovaries during the pupal stage, and are stored in the lumen of the oviduct (Norton & Vinson, 1983; Theilmann & Summers, 1986). During oviposition, PDVs are injected with eggs into the host, and enter different host tissues. Within the host tissues, PDVs do not replicate, but host-specific viral genes are expressed. PDV gene products function to disrupt the host immune system as well as host development, which appear to be essential for successful parasitoid development (Stoltz, 1993; Lawrence & Lanzrein, 1993, 1994; Lavine & Beckage, 1995; Strand & Pech, 1995).

The PDVs are recognized as two genera: the ichnoviruses found exclusively in parasitoid wasps of the Ichneumonidae, and the Bracoviruses found only in parasitoid wasps of the Braconidae. *Ichnovirus* and *Bracovirus* differ from each other in morphology, host range and molecular characteristics (Stoltz & Whitfield, 1992). Ichnoviruses characteristically contain one lenticular nucleocapsid per virion surrounded by two unit membranes, whereas bracoviruses characteristically contain one or more rod-shaped nucleocapsids per virion surrounded by a single unit membrane envelope. It is thought that Ichnoviruses are released into the ovary lumen by budding from calyx cells, while bracoviruses, in contrast, are believed to be released through lysis of the calyx cells.

The PDV genome structure is complex and poorly understood. Only a few PDV genomes have been described in detail. Estimates of PDV genome sizes have been complicated by the characteristic presence of unequal molar ratios of genomic segments, comigrating DNA segments of equal size, and direct terminal repeats (Fleming, 1992; Fleming & Krell, 1993; Stoltz, 1993). Comparisons of various PDV genomes showed that the DNA segment number and molar ratio and the total genome size appeared to be specific to...
the PDVs of each individual PDV-containing wasp species (Krell, 1991; Stoltz, 1993; Fleming & Krell, 1993). Descriptions of ichnovirus genomes have been largely based on studies of PDVs isolated from the parasitoid wasps *Campoplexis sonorensis* (Krell et al., 1982), *Hyposoter exiguae* (Krell & Stoltz, 1980) and *Diadegma terebrans* (Krell, 1987). Of these ichnoviruses, the *C. sonorensis* polydnavirus (CsPDV) genome has been by far the most systematically characterized, and appears to contain 28 DNA segments ranging in size from 5-5 to 21 kbp. Genome analyses of bracoviruses have been less complete than those described for Ichnoviruses. The available information on bracovirus genomes comes mainly from descriptions of the PDVs associated with the parasitoids *Cardiochiles nigriceps* (Varriechio et al., 1999), *Chelonus inanitus* (Albrecht et al., 1994; Johner & Lanzrein, 2002; Wyder et al., 2002) and *Microplitis demolitor* (Strand et al., 1992). In general, the genome segments of bracoviruses tend to be fewer in number but larger in size than those of the ichnoviruses, and exhibit a relaxed open circular topology (Albrecht et al., 1994; Webb, 1998). For example, characterization of the bracovirus of *C. inanitus* showed that its genome consisted of 10 different segments ranging in size from 7 to 31 kbp, and that individual DNA segments appeared to be singly encapsidated (Albrecht et al., 1994).

The braconid endoparasitic wasp *Glyptapanteles indiensis* parasitizes the larval stage of the gypsy moth, *Lymantria dispar*. Previous studies on *G. indiensis* polydnavirus (GiPDV) in our laboratory demonstrated that GiPDV could be integrated not only as a provirus within the parasitoid wasp genome, but also *in vitro* within the chromosomal DNA of cells derived from the natural host *L. dispar* (Gundersen-Rindal & Dougherty, 2000). More recently, a GiPDV putative protein tyrosine phosphatase gene believed to be associated with gene regulation during immune response was shown to be differentially expressed in various tissues of the parasitized host (Chen et al., 2003). Although information on the *in vitro* and *in vivo* properties of GiPDV has been gathered, fundamental knowledge pertaining to GiPDV virion morphology and genomic organization remains undescribed. To fill this gap, the morphological features, genomic organization and molecular features of GiPDV were investigated and are reported here. The isolation of cDNA clones representing GiPDV sequences from mRNA isolated from parasitized host and confirmation of the expression of cDNA clones in the parasitized host are described. Further, mapping of these cDNA fragments to the GiPDV genome is shown.

### METHODS

**Insect rearing.** The parasitoid wasp was reared from larvae of the host insect, *L. dispar*. Adult wasps were fed with 30 % honey water. Both *G. indiensis* and its host were maintained at 26 °C, 50 % relative humidity and a 16 h light:8 h dark photoperiod according to the methodology established by Bell et al. (1981). To obtain parasitized larvae, individual *L. dispar* larvae were exposed to a single *G. indiensis* female within a 35 × 10 mm Petri dish to ensure 100 % parasitization. After a single oviposition was observed, each parasitized larva was removed from the Petri dish and reared in a plastic cup containing high wheat-germ diet for 24 h prior to RNA extraction.

**Virus purification and viral DNA isolation.** Virus was purified from the calyx fluid of female wasps. Female wasps were anesthetized in 75 % ethanol for a few seconds and then rinsed in sterile H2O. Each pair of ovaries was dissected in a 15 μl drop of cold PBS and punctured individually with forceps, causing the calyx fluid to diffuse into the PBS drop. The calyx fluid was filtered through a 0-45 μm filter to remove eggs and cellular debris by the methods of Beckage et al. (1994). The resulting viral fluid was transferred into a 1-5 ml microcentrifuge tube and immediately processed for viral DNA isolation and electron microscopy of viral ultrastructure.

**Electron microscopy of viral particles.** For negative staining, GiPDV virus particles in filtered calyx fluid were absorbed onto a 400 mesh Formvar-coated nickel grid for 2 s, quickly washed in distilled water, stained with 4 % uranyl acetate for 1 min, and then viewed in an H-7000 Hitachi electron microscope at 75 kV. For thin sections, embedded ovaries dissected from female *G. indiensis* were fixed in 3 % glutaraldehyde/0-05 M sodium cacodylate buffer (pH 7-0) for 2 h at room temperature and placed in a 4 °C refrigerator overnight. After washing in sodium cacodylate buffer six times within 1 h, ovaries were post-fixed in 2 % buffered osmium tetroxide for 2 h, dehydrated in graded ethanol solutions and infiltrated with Spurr’s low-viscosity embedding resin. Ovaries were sectioned on a Riechert/AO Ultracut microtome with a Diatome diamond knife. Silver sections of the ovaries were mounted on 200-mesh Ni grids, stained with 4 % uranyl acetate and 3 % lead citrate, and observed in an H-7000 Hitachi electron microscope at 75 kV.

**Viral DNA isolation and field inversion gel electrophoresis (FIGE).** Filter-purified viral fluids were incubated in an equal volume of extraction buffer (4 % sarcosyl, 1 % SDS, 50 mM EDTA, 10 mM Tris, 0-2 M NaCl, 20 μg RNase ml⁻¹, 50 μg proteinase K ml⁻¹) at 50 °C for 2 h, followed by two extractions with ultrapure buffer-saturated phenol (Invitrogen). DNA was precipitated in 2 vols of cold ethanol and centrifuged at 15 000 r.p.m. for 30 min. The viral DNA pellet was resuspended in TE buffer and stored at 4 °C. The purified viral DNA was analysed by FIGE, an electro- phoretic method used to resolve DNAs in the 10–150 kb range. Undigested viral DNA (2 μg) and a molecular size standard (DIG-labelled linear λ DNA digested with HindIII) were loaded onto a 0-8 % (w/v) pulse field certified agarose gel (Bio-Rad) in 0-5 x TBE buffer (50 mM Tris/borate, 1 mM EDTA, pH 8-3). Gel electrophoresis was performed at 4 °C for 2 × 24 h using a 0-1–0-4 s switch time ramp with 180 V forward voltage and 120 V reverse voltage for each 24 h run. The gels were stained with ethidium bromide for 15 min, destained with sterile H2O for 30 min, and visualized using a UV transilluminator.

**cDNA library construction and screening.** Total RNA was isolated from parasitized *L. dispar* larvae 24 h post-parasitization. Insects were frozen in liquid nitrogen and ground to a fine powder. RNA was extracted with TRizol reagent (Invitrogen) according to the manufacturer’s instructions. RNA samples were resuspended in nuclease-free water in the presence of ribonuclease inhibitor (Invitrogen). The RNA quality was verified by formaldehyde gel electrophoresis, and mRNA was isolated from total RNA using the Messagemaker Reagent Assembly kit (Life Technologies) following the manufacturer’s instructions. The concentration of mRNA was measured using a spectrophotometer. cDNA libraries were prepared using the Superscript Choice System for cDNA Synthesis (Invitrogen) following the manufacturer’s instructions. mRNA was converted into size-fractionated, EcoRI-adapted cDNA. The cDNA was ligated into the EcoRI-digested ZIPLOX expression vector (Life Technologies). The ligated vector-cDNAs were packaged in Gigapack III Gold
Packaging Extract (Stratagene). The packaged cDNA-containing phage were introduced into Y1090 (ZL) E. coli that had been grown overnight in LB medium with 10 mM MgSO₄ and 0·2 % maltose. The transformants were plated on LB-agar and incubated overnight at 37 °C. The cDNA libraries were titrated and amplified, and approximately 5 × 10⁶ p.f.u. of library were plated onto ten large LB plates. The plaques were transferred to nitrocellulose membrane (Roche) and the phage DNAs were denatured, neutralized and UV cross-linked to membrane according to standard procedures. The GiPDV genomic DNA used as a probe was radiolabelled to high specific activity with the Random Primers DNA Labelling System (Invitrogen) following the manufacturer’s instructions. Hybridization screening of cDNA libraries using the 32P-labelled GiPDV probe was carried out in the hybridization solution (6 × SSPE, 0·5 % SDS, 5 × Denhardt’s solution, 100 μg denatured fragment salmon sperm DNA ml⁻¹) at 65 °C for 16 h. Membranes were washed once in lower stringency solution (1 × SSPE, 0·1 % SDS) for 15 min at room temperature and twice in medium stringency solution (0·5 × SSPE, 0·1 % SDS) at 48 °C for 15 min. After air-drying, membranes were exposed to Hyperfilm (Amersham Pharmacia) at −80 °C over-night. Positive plaques were identified and subjected to a second round of screening. Two positive clones confirmed by secondary screening were picked with a Pasteur pipette and resuspended in 500 μl of SM buffer (50 mM Tris/HCl, pH 7·9, 100 mM NaCl, 10 mM MgSO₄, 0·01 % gelatin) and stored at 4 °C with 20 μl of chloroform. Conversion of recombinant phage into plasmid (in vivo excision) was done by infecting E. coli DH10B(ZIP), according to the manufacturer’s instructions (Life Technologies). Plasmid DNA containing the cDNA insert was purified for each using a Plasmid Miniprep kit (Bio-Rad).

DNA sequence analysis and nucleotide sequence accession numbers. Sequencing reaction of isolated cDNA clones was carried out with the ABI PRISM Big Dye Terminator Cycle Sequence kit (Applied Biosystems) using T7 and SP6 promoter-specific primers, followed by primer walking. The nucleotide sequences of the cDNA clones were determined using an ABI PRISM 310 Genetic Analyser (Applied Biosystems). Sequence fragments were edited and assembled into contiguous regions using the SeqManII sequence assembling software of the DNASTAR package. Sequence homology searches were done using BLAST (Altschul et al., 1990) and searched against the GenBank database. Sequences have been deposited in GenBank with accession numbers AF414845 and AF414846, respectively.

Northern blot analysis. Expression of isolated cDNA clones in the parasitized host was confirmed by Northern blot analysis. The RNA samples isolated from parasitized L. dispar larvae 24 h post-parasitization and nonparasitized L. dispar larvae were resolved on denaturing formaldehyde/1 % agarose gel and blotted onto nylon membrane. DIG-labelled RNA probes complementary to two isolated cDNA clones were synthesized using the DIG RNA labelling kit (Roche). The membrane was prehybridized in prehybridization solution (50 % formamide, 5 % blocking reagent, 5 × SSC, 0·1 % sarcosyl, 0·1 % SDS) at 52 °C for 2 h, followed by hybridization with DIG-labelled RNA probes overnight. After hybridization, the membrane was washed twice in low stringency wash solution (2 × SSC, 0·1 % SDS) at room temperature for 5 min and washed twice in high stringency wash solution (0·1 × SSC, 0·5 % SDS) at 48 °C for 15 min. The hybridization signals were detected on X-ray film (Amersham) after a 30 min exposure. The same membrane was probed with two different cDNA probes individually. For repeated hybridization of the same blot, the nylon membranes were washed in stripping solution (0·2 M NaOH, 0·1 % SDS) for 30 min at 37 °C to remove the probe.

RESULTS

Electron micrographs of GiPDV

Thin section analysis of the calyx region in the lumen of the G. indiensis ovipuct revealed the presence of large quantities of viral particles (Fig. 1a). Variable densities of viral particles were seen among different calyx cells (Fig. 1b). Each viral particle contained, at most, ten nucleocapsids enveloped by a single unit membrane, with an average of six nucleocapsids per virion (Fig. 1c). The size and shape of virions were variable, with a mean length of 273 ± 34·24 nm, ranging from 175 to 350 nm, and a mean width of 162 ± 19·32 nm, ranging from 140 to 190 nm (n = 80) (Fig. 2a).

A negative stain of GiPDV particles present in calyx fluid showed the nucleocapsids were rod-like or rectangular in shape. The electron-dense cores were probably DNA genome with their associated proteins (Fig. 1d). The sizes of nucleocapsids were slightly variable, with a mean length of 58 ± 5·21 nm, ranging from 46 to 66 nm, and a mean width of 38 ± 6·37 nm, ranging from 34 to 52 nm (n = 60) (Fig. 2b).

Electrophoretic profiles of GiPDV genomic DNA

GiPDV FIGE hybridization patterns showed that GiPDV comprised multiple DNA segments, similar to other previously described polydnaviruses, and 13 visible bands were recognized ranging in size from approximately 11 kb to more than 30 kb (Fig. 3). The various DNA segments were present in non-equimolar ratios, as indicated by differing band intensities. Four segments (D, F, K and M) were present in higher molar concentration than the other segments (Fig. 3). Assuming that each DNA segment was unique, and that there were no co-migrating DNA segments, GiPDV genomic size was estimated to be 250 kb.
Screening of parasitized host cDNA libraries and isolation of expressed GiPDV-specific cDNA clones

To further characterize the complexity of the GiPDV genome, cDNA was synthesized and cloned from mRNA isolated from parasitized *L. dispar* larvae 24 h after *G. indiensis* parasitization. After screening the library with GiPDV, two positive cDNA clones, which derived from GiPDV-specific mRNA expressed in parasitized hosts, were identified, isolated and sequenced. The first cDNA clone, cDNA 1-1, contained a 1·1 kb insert, whereas the second cDNA clone, cDNA 1-8, contained a 1·8 kb insert. The sequences of cDNA 1-1 and cDNA 1-8 are presented in Fig. 4 and Fig. 5. Clone 1-1 contained a single large open reading frame (ORF) encoding 325 amino acids. Clone 1-8 contained one large ORF comprising 494 amino acids. BLAST searches of these sequences did not yield any significant homology to nucleotide or amino acid sequences in GenBank.

**Northern blot analysis of cDNA clones**

The expression of cDNA 1-1 and cDNA 1-8 in parasitized host was confirmed by Northern hybridization. Using

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**Fig. 1.** Electron micrographs of *G. indiensis* polydnavirus. (a) Thin section showing calyx region in the lumen of the *G. indiensis* oviduct. Large numbers of viral particles were seen in calyx epithelial cells. (b) Thin section showing separate desmosomes joining two calyx cells. Different densities of viral particles between the calyx cells may show different levels or stages of replication occurring in the two cells. (c) Thin section showing virus particles within the calyx fluid. Each virion consists of 5 to 10 nucleocapsids enclosed by a single membrane envelope. Bar marker represents 0·5 μm. (d) Negative staining of liberated GiPDV nucleocapsids (35 nm wide × 46 nm long). Electron micrograph: Charlie Murphy and Christopher Pooley.
DIG-labelled RNA complementary to cDNA 1·1 as a hybridization probe, one transcript of 1·1 kb was identified. Using DIG-labelled RNA complementary to cDNA 1·8 as a hybridization probe, one major transcript of 1·8 kb and two minor transcripts of 1·7 kb and 2·6 kb were identified. No hybridization signal was detected with RNA extracted from non-parasitized gypsy moth host (Fig. 6).

**Mapping of isolated cDNA clones to the GiPDV genome**

In order to determine which GiPDV genomic DNA segments encoded these two expressed GiPDV-specific mRNAs, cDNA 1·1 and cDNA1·8 were DIG-labelled and used as probes to hybridize individually to an identical Southern blot containing size-fractionated genomic DNA. Mapping studies indicated that both cDNAs hybridized to more than one genomic DNA segment and different cDNAs were present on the same genome segment. cDNA 1·1 probe hybridized to four viral DNA segments (C, D, E, K) and cDNA 1·8 probe hybridized to two viral DNA segments (J, K), as shown in Fig. 7.

**DISCUSSION**

**Characteristics of GiPDV morphology**

Polydnaviruses have been found to be mutualistically associated with some hymenopteran endoparasites in the families Braconidae and Ichneumonidae and have been characterized in approximately 40 species of wasps (Fleming, 1992). The morphology of the braconid polydnavirus GiPDV was observed by transmission electron microscopy and the morphological characteristics were similar to those of other described bracoviruses. The calyx fluid of *G. indiensis* females contained large quantities of GiPDV viral particles. The GiPDV virions were ovoid in form and contained electron-dense nucleocapsids enclosed by a single unit membrane envelope. Compared with previously characterized bracoviruses, virions of GiPDV seemed to contain more numerous nucleocapsids; as many as six to ten nucleocapsids in a single virion were frequently observed. Variation in the number of nucleocapsids that appeared in the virions may have been due to the different angles at which the virions were observed under electron microscopy. Compared with those of other bracoviruses, GiPDV nucleocapsids were fairly large, with a mean length of $58 \pm 5\:21$ nm, ranging from 46 to 66 nm, and a mean width of $38 \pm 6\:37$ nm, ranging from 34 to 52 nm. The largest nucleocapsids reported to date were found in the PDV associated with *Chelonus texanus*, with each PDV having a
diameter of 35 to 40 nm and a length of 50 to 100 nm (Stoltz & Vinson, 1979). The smallest nucleocapsids were seen in the PDV associated with *Cardiochiles nigriceps*, with a diameter of 25 to 50 nm and a length of 10 nm (Stoltz & Vinson, 1979). As a general rule, the number and size of nucleocapsids were species-specific among the bracoviruses and were positively related to the size of DNA segments enclosed. It was reported that nucleocapsids of bracovirus contained only one DNA segment and DNA molecules were singly encapsidated, as indicated by the release of nucleic acid particles from the virions. The nucleotide sequence and deduced translated amino acids of GiPDV cDNA 1·1 are shown in Fig. 4. The putative translation initiation site is in bold and indicated by an arrow. Stop codon is bold and indicated by an asterisk.

**Fig. 4.** Nucleotide sequence and deduced translated amino acids of GiPDV cDNA 1·1. The putative translation initiation site is in bold and indicated by an arrow. Stop codon is bold and indicated by an asterisk.
**cDNA 1.8:**

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1  gqc gcc ggt cga cct atg cta atc taa ttc att tat aag acc aag aag gaa acc qtg gga
61  tgt ttc cat gat cta cac ata gac ttt ttc aag aat ctt ctc acc cca aag cag tag
121  agt tgg aga atc gaa aca [atg] cgt ggt gct ggc tgc ttt gtt tta ttg tgt atc gcc acc
181  gtc gtc tca aat aag aqa aag aag aag aag gaa tat gga gat gcc atg acc ttc aaa
241  gaa gaa gtc gaa aqa aag gta att gtc ttc cag ggt aqc aag aac gac qtg act att gca
301  gag cca act aat gqg gat ggt tgg aga ttc cag att ttc gaa aga aac agg gga aag
361  tta gga gat acc cqa acq act gag agt aga aag gaa gtt ggg atg acc ttc aaa gac
421  gaa ccc gga aqa ata att atc acc ccg ggt ggg aag tac aag tta gaa gag tca att
481  aca gac aat tat gat gat gga tgt aga ttc acc atc ttt aag aag gaa aag cag cga
541  aaa tta gaa aat aqa aca aac ctg atc gtc gtt gcc gaa gcg gaa cag ttt ctc aag gag att
601  tat cct cag gga gcc ttc aqa ggt att cqa acq gta gtc aca gga agt gtc gac gat aaa gat
661  cct cga aaa aat gct ctt tta gcc gag cag tgc gcg gaa act tat ccg gat ttc gaa tca tgg
721  pra cca atg ttt cct cga cca atc gtc ttc gca aqa aqa aac act aat cag cca gaa aac
781  rmp frf prf pfv ilv slk ktt nnn qen
841  ggt cct cca ccc aga tgt gaa ata tta gca act cct cct ccc cag cat gcc cct gaa tgc
901  ggc gct gat cgc gaa gta cta gcc cag aca gtt ggt ggg aag aca gga ggt taa cca cag tat
gek vsa tvg vrt ggt gty ypf poy
961  gga cct gga tta gga cat aac ccc cag cat cct tct gaa gtc aat gaa gcg cct cct ctc
1021  qca gca cat aat ggc gag aqa gta cca gcc cag aca gtt ggt ggt gaa atg gca aqa ggt taa
1081  cag cct cag tat gga cct gga cct gga tat aac ccc cag gat ctt cct ata gtc aat cca
1141  tca ggt cct cag gcc ctc cag aat gga gaa aag gta cta gcc cag aca aat gga atg
1201  gga cct cgc cta cgc ctc cag tta aag aca gcc ctc cct cca gat cct gct
1261  ata gtc aac gag cgc cct cct ccc gaa gat tat agc aag gga gta cca ccc cag cca
1321  gtt gat gga aag cca gca gct cag aat ggt ggt ggg aag aca gga aat tac ttt
gvd rgt tga pty qyg gpf gys gny fl
1381  cag ggt ccc tct gta gtc aac gag cac cct cct tcc aca gct cat aat agc gag aga cta
1441  tac ggc cca aca cgg ggt gaa aag cca gca ggt tat cca cct cag tat gga cct cta
1501  gga cat aac ccc cag gat cct tct gtc aat gaa cga cct cct ctc aca gct gat tac
1561  gtt caa gta tac ggg cca gtt ctt ttc aag cag ttc cca ccc cct ttc ttt ggg acc ctc aat
gqv yvg pfr qsp fs gtl
1621  tag att att tgt gga aqa atc gtt tgg tct tta aag tca ccc gac aat aat aca tcc
1681  gtc gtt tca tta atg cac aac cta cgg aac aca tca tgg tta gtt ttc
1741  gga gat ttt ttt cta ctc gta cgg ttt ccc gta gta aag tta tga tca cct aac gtc
1801  gac ggc g
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**Fig. 5.** Nucleotide sequence and deduced translated amino acids of GIPDV cDNA 1.8. The putative translation initiation site is in bold and indicated by an arrow. Stop codon is bold and indicated by an asterisk.
individual closed circular DNA segments from the nucleo-
capsids after osmotic shock (Albrecht et al., 1994).

Characteristics of the GiPDV genome

Bracoviruses appear to comprise fewer but larger DNA
molecules than ichnoviruses, and exhibit a relaxed open
Electrophoretic profiles of genomic DNA showed that the
GiPDV genome contained 13 segments ranging in size
from 11 kb to more than 30 kb. The aggregated GiPDV
genome size was approximately 250 kb. The visible GiPDV
segments may represent distinctive genomic molecules or
the same genomic molecules in different conformations.
Since supercoiled, relaxed circular and linear forms of
viral DNA were very difficult to differentiate, the poten-
tial increase in sequence complexity associated with co-
migration of DNA segments of the same size or the same
DNA segment present in more than one conformation was
not included in the size estimate for the GiPDV genome in
this study. The ongoing GiPDV sequencing project in our
laboratory will provide a better and more precise character-
zation of genome size and organization in the future.
Compared with the most systematically characterized
ichnovirus, C. sonorensis polydnenvirus (CsPDV), contain-
ing 28 DNA segments ranging in size from 5.5 to 21 kbp,
GiPDV comprised fewer but larger DNA segments. Con-
sistent with previous findings for other ichno- and braco-
viruses, GiPDV consisted of multiple DNA segments with
variable sizes and molar ratios (Theilmann & Summers,
1987, 1988; Summers & Dib-Hajj, 1995; Cui & Webb, 1997;
Strand et al., 1997; Webb & Cui, 1998, Varricchio et al.,
1999). Four segments (D, F, K and M) were present in higher
molar concentration than other segments, as indicated by
a higher band intensity. The higher molar concentration
of DNA segments indicated the presence of multiple
DNA segments of similar size with either homologous or
polymorphic sequences.

Identification of cDNA clones encoding GiPDV-
specific mRNA and expression of cDNA clones
in the parasitized host

Construction and screening of a cDNA library allowed
identification of cDNA clones deriving from GiPDV-
specific mRNA expressed in parasitized host 24 h post-
parasitization. Both cDNA clones contained a single ORF.
Analysis of amino acid sequences indicated that isolated
transcripts have no significant homology to other ichno-
virus or bracovirus gene families (Webb & Cui, 1998).Northern blot analysis confirmed the expression of cDNA
1·1 and cDNA 1·8 in the parasitized host, with cDNA 1·1 expressed as a single 1·1 kb transcript and cDNA 1·8 expressed as one major 1·8 kb transcript, with minor 1·7 and 2·6 kb transcripts also detected in the parasitized host. Whether the minor transcripts resulted from alternative splicing of the same precursor transcript or were derived from independent genes is unknown.

Southern hybridization showed that cDNA 1·1 resided on four separate viral DNA segments, while cDNA 1·8 apparently resided on two viral DNA segments, indicating multiple gene loci within GiPDV. The multiple gene loci suggested the probability of homologous sequences or related genes among different GiPDV DNA segments, as is characteristic of other described polydnaviruses (Theilmann & Summers, 1988; Cui & Webb, 1997, 1998; Strand et al., 1997; Volkoff et al., 1999). These are the first internal sequence homologies noted for GiPDV. The features of genome segmentation, hypermolar segment ratios and sequence homology have been suggested to be involved in increasing the copy number of essential genes and the levels of gene expression in the absence of virus replication (Xu & Stoltz, 1993; Cui & Webb, 1997). Homologous viral genes residing on different DNA segments could be transcribed simultaneously or separately to exert additive or complementary functions in host regulation. Webb & Cui (1998) reported that abundantly expressed genes are often associated with nested DNA segments, while genes that are expressed at a lower level are not generally present on nested DNA segments. The multiple loci among GiPDV genome segments suggested that cDNA 1·1- and 1·8-encoded GiPDV-specific genes represent related genes of importance in the parasitized host, members of gene families, and are probably not associated with nested segments.

The morphological features and genomic organization of the bracovirus of GiPDV are unique, although characteristic of braconid PDVs in general. Isolation of cDNAs encoding GiPDV-specific mRNA expressed in the parasitized host, as well as mapping of these cDNAs to multiple GiPDV genomic DNA segments, supported the notion that GiPDV morphology and genomic organization are intrinsically linked to the function and evolutionary strategies of the virus. This study enhances knowledge about this virus and provides a basis for future functional characterization of GiPDV.

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