**Thermococcus eurythermalis** sp. nov., a conditional piezophilic, hyperthermophilic archaeon with a wide temperature range for growth, isolated from an oil-immersed chimney in the Guaymas Basin

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A conditional piezophilic, hyperthermophilic archaeon showing growth over a wide range of temperature, pH and pressure was isolated from an oil-immersed hydrothermal chimney at a depth of 2006.9 m in the Guaymas Basin. Enrichment and isolation of strain A501T were performed at 80 °C at 0.1 MPa. Cells of isolate A501T were irregular motile cocci with a polar tuft of flagella and generally 0.6–2.6 μm in diameter. Growth was detected over the range 50–100 °C (optimal growth at 85 °C) at atmospheric pressure and was observed at 102 °C at a pressure of 10 MPa. At 85 °C, growth was observed at a pressure of 0.1–70 MPa (optimum pressure 0.1 MPa–30 MPa), while at 95 °C, the pressure allowing growth ranged from 0.1 MPa to 50 MPa (optimum pressure 10 MPa). Cells of strain A501T grew at pH 4–9 (optimum pH 7.0) and a NaCl concentration of 1.0–5.0 % (w/v) (optimum concentration 2.5 % NaCl). This isolate was an anaerobic chemo-organoheterotroph and was able to utilize yeast extract, peptone, tryptone and starch as the single carbon source for growth. Elemental sulfur and cysteine stimulated growth; however, these molecules were not necessary. The DNA G+C content of the complete genome was 53.47 mol%. The results of 16S rRNA gene sequence analysis indicated that strain A501T belongs to the genus **Thermococcus**. There was no significant similarity between strain A501T and the phylogenetically related species of the genus **Thermococcus** based on complete genome sequence alignments and calculation of the average nucleotide identity and the tetranucleotide signature frequency correlation coefficient. These results indicate that strain A501T represents a novel species, **Thermococcus eurythermalis** sp. nov. The type strain is A501T (≡CGMCC 7834T=JCM 30233T).

A deep-sea hydrothermal vent is a likely environment for the discovery of extremophiles (Jørgensen & Boetius, 2007) and is ideal for studying multiple extreme adaptations of micro-organisms because of the unique physical conditions, including the combination of high temperature, high hydrostatic pressure, extreme pH, reducing power, toxic chemistry and steep physical-chemical gradients (Gorlas et al., 2013). Many studies on the microbial diversity of the deep-sea hydrothermal vent ecosystem have shown that the euryarchaea group is abundant, including the oftencultured members of the order **Thermococcales** (Takai et al., 2006). Moreover, species of the order **Thermococcales** are relatively abundant and a great diversity of species including thermophiles and hyperthermophiles have been isolated from various geothermally heated systems (Prieur, 2002). The order **Thermococcales** represents an archaeal order that contains only one family, **Thermococcaceae**. Three genera, **Thermococcus** (Zillig et al., 1983), **Pyrococcus** (Fiala & Stetter, 1986) and **Palaeococcus** (Takai et al., 2000), belong to this family. Members of the genus **Thermococcus** are organotrophic and anaerobic hyperthermophiles. Elemental sulfur is required for the growth of some species, and elemental sulfur stimulates growth significantly in other species. Twenty-nine species of the genus **Thermococcus** have been characterized to date according to the List of Prokaryotic Names with Standing in Nomenclature (LPSN; http://www.bacterio.net), and can be divided into two groups based on their respective DNA G+C content (Jolivet et al., 2004); one group has a higher DNA G+C content of 50–60 %, and the other group has a lower DNA G+C content of 38–47 %. The temperature

The GenBank/EMBL/DDBJ accession number for the 16S rRNA gene sequence of strain A501T is KJ616741. The accession numbers for the complete genome sequence of strain A501T are CP008887 and CP008888.

Two supplementary figures and one supplementary table are available with the online Supplementary Material.
supporting growth of species of the genus *Thermococcus* ranges from 48 °C to greater than 100 °C, and the majority of species have a wide temperature range for growth of more than 30 °C. Compared with species of the genus *Pyrococcus*, the response to pressure in species of the genus *Thermococcus* is not well reported. *Thermococcus barophilus* was the first true piezophilic, hyperthermophilic archaeon described within the order *Thermococcales* (Marteinsson et al., 1999).

In this study, strain A501T was isolated from an oil-immersed chimney of a deep-sea hydrothermal vent in the Guaymas Basin and was characterized as a representative of a novel species of the genus *Thermococcus*. Strain A501T was a conditional piezophilic, hyperthermophilic archaeon that could grow over wide ranges of temperature, pressure and pH.

Samples from the spire of a chimney, white-covered with oily areas, were collected by the submersible *Alvin* from a hydrothermal vent site at a depth of 2006.9 m in the Guaymas Basin [Gulf of California (27° 0.405′ N 110° 24.567′ W)] during a cruise on 14 November 2009. The preliminary temperature inside the samples was 229 °C. The samples were immediately transferred into sterile sample tubes, sealed with a lid and covered with aluminium foil; the samples were preserved at 4 °C. Members of the order *Thermococcales* are among the dominant micro-organisms in these samples, as shown by meta-genomic data (He et al., 2013).

In the laboratory, the samples were used for enrichment cultures. These samples were incubated anaerobically at 80 °C for 7 days in YPS medium (Jolivet et al., 2003). Positive enrichments were obtained. A coccoid bacterial strain was purified by the dilution-to-extinction method in YPS medium containing strain A501T with 5 % (v/v) DMSO (Sigma) as the gas phase, and 5 ml syringes with TRM were used at high hydrostatic pressures. The medium was inoculated with late exponential-phase cells at approximately 10⁶ cells ml⁻¹. All of the samples were tested in duplicate, and uninoculated medium was used as a negative control. Growth was measured by determining the cell number, which was counted by microscopy or by flow cytometry (Marteinsson et al., 1999).

Strain A501T grew over the temperature range 50 °C to 100 °C at atmospheric pressure (Fig. S1a, available in the online Supplementary Material). Optimal growth occurred at 85 °C; no growth was observed at 102 °C. Cells lost the ability to divide after 12 h of cultivation at 102 °C, possibly because no growth occurred when the culture was inoculated back to fresh medium at 85 °C at atmospheric pressure. However, growth at 102 °C did occur at 10 MPa, and no growth was observed at 100 °C at the same pressure. At 10 MPa or 40 MPa, strain A501T maintained the ability to divide, even at 112 °C, for at least 12 h; significant growth up to 10⁸ cells ml⁻¹ was observed after inoculating the culture back to fresh medium at 85 °C at atmospheric pressure. However, no growth occurred after inoculating a culture incubated at 115 °C for only 4 h back to fresh TRM at 85 °C and 0.1 MPa.

At 85 °C, growth was recorded over a pressure range from 0.1 MPa to 70 MPa (Fig. 2). The optimum pressure was 0.1–30 MPa because no significant difference was detected in the specific growth rate at these pressures. However, the growth yields at 10 MPa and 20 MPa were larger than those at 0.1 MPa and 30 MPa. No growth occurred at 80 MPa. At 95 °C, the pressure range narrowed to 0.1–50 MPa, and the optimum pressure was 10 MPa (Fig. 2). No growth occurred at 60 MPa. These results indicate that the isolate is a conditional piezophile.

The range of pH and NaCl concentration for growth were determined in TRM medium at 85 °C at atmospheric pressure. Positive enrichments were obtained. A coccoid bacterial strain was purified by the dilution-to-extinction method in YPS medium at 80 °C. A single colony was isolated with a scraping from a single colony and named A501T. For long-term storage, cryotubes were repeated three times. The purified strain was obtained by inoculating a culture incubated at 115 °C to 85 °C at atmospheric pressure. However, no growth occurred after 12 h of cultivation at 102 °C, possibly because no growth occurred when the culture was inoculated back to fresh medium at 85 °C at atmospheric pressure. However, growth at 102 °C did occur at 10 MPa, and no growth was observed at 100 °C at the same pressure. At 10 MPa or 40 MPa, strain A501T maintained the ability to divide, even at 112 °C, for at least 12 h; significant growth up to 10⁸ cells ml⁻¹ was observed after inoculating the culture back to fresh medium at 85 °C at atmospheric pressure. However, no growth occurred after inoculating a culture incubated at 115 °C for only 4 h back to fresh TRM at 85 °C and 0.1 MPa.

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Fig. 2. Effects of the hydrostatic pressure and temperature on the specific growth rate of strain A501\textsuperscript{T}. Cells were grown in TRM medium at 85 °C (■) and 95 °C (●) at different hydrostatic pressures. The growth rates at and below 50 MPa were calculated by linear regression analysis along the exponential portion of the growth curves. For the operation of equipment used to test growth under higher pressures (>50 MPa), the growth rates were estimated as $\mu = (\ln N_2 - \ln N_1) / (t_2 - t_1)$, where $N_2$ and $N_1$ were the numbers of cells at the times $(in \ h)$ $t_2$ and $t_1$, respectively. Error bars present the standard deviation shown when the results of more than two separate experiments were available.

The medium used in the carbon source utilization tests was basal TRM (pH 7.0, no yeast extract or tryptone) with a single carbon source at a final concentration of 0.2 % (w/v) supplemented with 0.002 % (w/v) yeast extract and elemental sulfur (10 g l\textsuperscript{-1}). The headspace gas in the tests was 20 % air and 80 % N\textsubscript{2}. Yeast extract, tryptone, Casamino acids, glycerogen, starch, citrate, lactate, acetate, formate, butyrate, propionate, pyruvate, succinate, cellobiose, trehalose, galactose, glucose, sucrose, fructose, cysteine, methionine, asparagine, histidine, lysine, tryptophan, tyrosine, phenylalanine and arginine could not stimulate the growth of strain A501\textsuperscript{T} under test conditions.

Elemental sulfur and cysteine stimulated the growth of strain A501\textsuperscript{T} by three- to fourfold, although these molecules were not necessary for growth. Other sulfur substitutes, such as thiosulfate, polysulfide or methionine, could not stimulate growth.

Genomic DNA was prepared by the alkaline-SDS method. The entire genome of strain A501\textsuperscript{T} (unpublished) was sequenced by the paired-end strategy at Shenzhen Huada Genomics Institute (BGI; Shenzhen, China). Nearly 1340 Mb of reads was produced from a 500 bp paired-end library, and approximately 1013 Mb of reads was produced from a 5 kb paired-end library. The total of the reads covered the genome approximately 1105-fold. All of these reads were assembled by BGI, and 11 contigs (ranging from 3.5 kb to 543.3 kb) were generated. Gaps were closed and verified by PCR. The chromosome of strain A501\textsuperscript{T} was 2 122 535 bp and circular; the DNA G+C content was 53.47 mol\%. A501\textsuperscript{T} possessed one 3.6 kb plasmid with 46.71 mol\% DNA G+C content, which was considered as extra-chromosomal DNA.

The 16S rRNA genes (1488 bp) were predicted by the RNAmmer 1.2 Server on the basis of the entire genome sequence. BLAST results indicated that strain A501\textsuperscript{T} was a member of the genus Thermococcus. Similarity between A501\textsuperscript{T} and closely related species was greater than 98 %. The phylogenetic relationship based on 16S rRNA gene sequences of strain A501\textsuperscript{T} is shown in Fig. 3. The most closely related species are Thermococcus kodaikarensis (99.1 % 16S rRNA gene sequence similarity) and Thermococcus gammatolerans (99.0 % similarity). Another phylogenetic tree based on the whole genome amino acid sequences was also reconstructed by using the CVTree tool (Xu & Hao, 2009), which is shown in Fig. S2.

Because of the high levels of similarity between species of the genus Thermococcus, quantitative DNA–DNA hybridizations were necessary. Because the two species most closely related to the species A501\textsuperscript{T} have complete genome sequences, in this study, quantitative DNA–DNA hybridizations were simulated by similarity analysis based on whole genome sequence alignments with the BLAST tool. The results of the alignments showed that the proportion of similar sequences between the genome sequence of strain A501\textsuperscript{T} and the complete genome sequences of species of the genus Thermococcus available in GenBank was less than 50%. The most similar species is Thermococcus sp. 4557, with 46 % coverage. T. kodaikarensis KOD1 and Thermococcus gammatolerans EJ3, which represent the

control, and inoculated basal TRM was used as a negative control. No growth was observed in the inoculated basal TRM. All of the tests were performed in duplicate.

In tests of substrate utilization, yeast extract, tryptone and peptone supported rapid and significant growth of strain A501\textsuperscript{T}. Starch supported poor growth, with a maximum yield of only approximately $10^7$ cells ml\textsuperscript{-1}. No obvious growth was observed with the other carbon sources under the test conditions.

W. Zhao, X. Zeng and X. Xiao

All of the carbon source utilization tests were performed at 85 °C at atmospheric pressure and included inoculations with late exponential-phase cells ($10^6$ cells ml\textsuperscript{-1}). Growth was determined after 12 h and 24 h of incubation. For these tests, inoculated normal TRM was used as a positive control. No growth was observed in the inoculated basal TRM. All of the tests were performed in duplicate.
most closely related species based on 16S rRNA gene sequences, both have 43% similar sequences with an identity of 87% and 90%, respectively. For 'Thermococcus onnurineus', Thermococcus litoralis and T. barophilus, the proportion of similar sequence in the genomes of their type strains was 33%, 7% and 5%, respectively. All of the coverage values were below the threshold of 70% DNA–DNA hybridization, which is generally accepted for delineation of a novel genomic species. Moreover, the average nucleotide identity (ANI) and the tetranucleotide signature frequency correlation coefficient (TETRA) between two genomes were calculated by JSpecies as a substitute for DNA–DNA hybridization (Richter & Rossello-Móra, 2009). The results showed that all of the ANI and TETRA values between strain A501T and related species were below the threshold values for delineating species, which are 95–96% and 0.99, respectively (Table S1). Based on this method, the most similar species to strain A501T was T. gammatolerans, with an ANI value of 79.87% and a TETRA value of 0.97284. While the ANI and TETRA values when strain A501T was compared with T. kodakarensis were 75.97% and 0.95686. Thus, it is reasonable to suggest that the isolate A501T represents a novel species.

Compared with phylogenetically related species of the genus Thermococcus, strain A501T could be distinguished on the basis of different taxonomic parameters (Table 1). Strain A501T was distinguishable from all of the related species based on its wide temperature range for growth at 0.1 MPa. The variation in the temperature range was up to 50°C. Moreover, this strain differed from most of the other species in its pH range for growth, with Thermococcus barossii as the exception. However, the DNA G+C contents of T. barossii and strain A501T are different. Strain A501T was distinguishable from T. kodakarensis and T. gammatolerans, the two most closely related species based on the 16S rRNA sequencing analysis, by the optimum NaCl concentration for growth, substrate utilization and the function of elemental sulfur for growth. On the basis of the phenotypic and genetic characteristics of strain A501T, we report a novel species within the genus Thermococcus. We propose to name this species Thermococcus eurythermalis sp. nov. according to its wide temperature range for growth.

**Description of Thermococcus eurythermalis sp. nov.**

Thermococcus eurythermalis (eu.ry.ther.ma’lis. Gr. adj. eurus wide, broad; L. adj. thermalis thermal; N.L. masc. adj. eurythermalis with a broad temperature range).

Cells are irregular cocci (diameter 0.6–2.6 μm) and motile with a polar tuft of flagella. Growth occurs over the
Table 1. Characteristics of species of the genus *Thermococcus* that distinguish strain A501<sup>T</sup> from its closest phylogenetic relatives

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
<th>9</th>
<th>10</th>
</tr>
</thead>
<tbody>
<tr>
<td>Taxa:</td>
<td>1, <em>Thermococcus eurythermalis</em> sp. nov. A501&lt;sup&gt;T&lt;/sup&gt; (this study); 2, <em>T. gammatolerans</em> (Jolivet et al., 2003); 3, <em>T. kodakarensis</em> (Atomi et al., 2004); 4, <em>T. peptonophilus</em> (González et al., 1995); 5, <em>T. stetteri</em> (Miroshnichenko et al., 1998); 6, <em>T. coalescens</em> (Kuwabara et al., 2005); 7, <em>T. barossii</em> (Duffaud et al., 1998); 8, <em>T. hydrothermalis</em> (Godfroy et al., 1997); 9, <em>T. profundus</em> (Kobayashi et al., 1994); 10, <em>T. gorgonarius</em> (Miroshnichenko et al., 1998).</td>
<td>+, Positive; −, negative; ND, no data; R, required; S, stimulatory.</td>
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<tr>
<td>Geographical origin</td>
<td>Guaymas Basin</td>
<td>Guaymas Basin</td>
<td>Kodakara Island, Kagoshima, Japan</td>
<td>Izu-Bonin forearc</td>
<td>Kraternaya cove, Ushishir archipelago, Northern Kurils</td>
<td>Suiyo Seamount, Izu-Bonin Arc</td>
<td>Juan de Fuca Ridge</td>
<td>East Pacific Rise</td>
<td>Middle Okinawa Trough</td>
<td>Whale Island, New Zealand</td>
</tr>
<tr>
<td>Sample type</td>
<td>Chimney, hydrothermal vent</td>
<td>Chimney, hydrothermal vent</td>
<td>Solfataric field</td>
<td>Hydrothermal vent</td>
<td>Marine solfataric field</td>
<td>Hydrothermal fluid</td>
<td>Rock fragment, hydrothermal vent</td>
<td>Rock suspensions, hydrothermal vent</td>
<td>Hydrothermal vent</td>
<td>Geothermal vent, tidal zone</td>
</tr>
<tr>
<td>Depth (m)</td>
<td>2007</td>
<td>2616</td>
<td>0</td>
<td>ND</td>
<td>ND</td>
<td>1380</td>
<td>ND</td>
<td>ND</td>
<td>1395</td>
<td>40</td>
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<tr>
<td>Motility</td>
<td>++</td>
<td>+</td>
<td>++</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
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<tr>
<td>Size (μm)</td>
<td>0.6–2.6</td>
<td>0.6–1.4</td>
<td>ND</td>
<td>0.7–2</td>
<td>1–2</td>
<td>0.3–2</td>
<td>0.7–3.7</td>
<td>0.8–2</td>
<td>1–2</td>
<td>0.3–1.2</td>
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<tr>
<td>Temperature range for growth (°C)</td>
<td>50–100 (0.1 MPa)</td>
<td>55–95</td>
<td>60–100</td>
<td>60–100</td>
<td>55–94</td>
<td>57–90</td>
<td>60–92</td>
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<td>68–95</td>
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<td>Optimum temperature for growth (°C)</td>
<td>85</td>
<td>88</td>
<td>85</td>
<td>85–90</td>
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<td>87</td>
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<td>4–8</td>
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<td>3.5–9.5</td>
<td>4.5–8.5</td>
<td>5.8–8.5</td>
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<td>6.5</td>
<td>6</td>
<td>6.5</td>
<td>6</td>
<td>6.5–7.5</td>
<td>6</td>
<td>ND</td>
<td>6.5–7.2</td>
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<td>[NaCl] range for growth (% w/v)</td>
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<td>1–5</td>
<td>1–5</td>
<td>1–5</td>
<td>ND</td>
<td>1.5–4.5</td>
<td>1–4</td>
<td>1–6</td>
<td>1–6</td>
<td>2–3.5</td>
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<tr>
<td>Optimum [NaCl] for growth (% w/v)</td>
<td>2.5</td>
<td>2</td>
<td>3</td>
<td>3</td>
<td>2.5</td>
<td>2.5</td>
<td>ND</td>
<td>3–4</td>
<td>ND</td>
<td>2–3.5</td>
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<td>Piezophilic response</td>
<td>+</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>+</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
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<tr>
<td>Substrates utilized for growth</td>
<td>Starch</td>
<td>Weakly</td>
<td>−</td>
<td>+</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>+</td>
<td>ND</td>
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<tr>
<td>Sucrose</td>
<td>−</td>
<td>ND</td>
<td>+</td>
<td>−</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>+</td>
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<tr>
<td>Maltose</td>
<td>−</td>
<td>−</td>
<td>−</td>
<td>−</td>
<td>−</td>
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<td>−</td>
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<td>Pyruvate</td>
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<td>−</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
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<tr>
<td>DNA G+C content (mol%)</td>
<td>53.5</td>
<td>51.3</td>
<td>52</td>
<td>52</td>
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<td>53.9</td>
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<td>58</td>
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<td>50.6</td>
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<td>Sulfur requirement</td>
<td>s</td>
<td>R</td>
<td>s/R</td>
<td>s</td>
<td>R</td>
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<td>Doubling time under optimum conditions (min)</td>
<td>48</td>
<td>95</td>
<td>86</td>
<td>25</td>
<td>ND</td>
<td>24</td>
<td>35</td>
<td>90</td>
<td>50</td>
<td>ND</td>
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</table>

*Data from Canganella et al. (1997).*
temperature range 50–100 °C (optimal growth at 85 °C) at 0.1 MPa and extends to 102 °C at 10 MPa. Hydrostatic pressure range for growth is 0.1–70 MPa (optimal growth at 0.1–30 MPa) at 85 °C and narrows to 0.1–50 MPa (optimal growth at 10 MPa) at 95 °C. Grows at pH 4–9 (optimal growth at pH 7.0) and with 1.0–5.0% (w/v) NaCl (optimal growth with 2.5% NaCl). Doubling time under optimal growth conditions is 48 min. Strictly anaerobe. Obligately chemo-organotroph. Yeast extract, tryptone, peptone and starch as single carbon sources can support growth, but starch supports very poor growth. Growth is optimal at 100 °C and requires 2.5% NaCl. Doubling time under optimal growth conditions is 48 min. Strictly anaerobe.

The type strain, A501T (=CGMCC 7834T = JCM 30233T), was isolated from the chimney sample of a deep-sea hydrothermal vent site at a depth of 2006.9 m; the vent was located in the Guaymas Basin (27° 0.405’ N 110° 24.567’ W).

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References


