Candida halmiae sp. nov., Geotrichum ghanense sp. nov. and Candida awuaii sp. nov., isolated from Ghanaian cocoa fermentations

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During an investigation of the microbiology of Ghanaian cocoa fermentations, a number of yeast isolates with unusual pheno- and genotypic properties representing three possible novel species were isolated. Members of Group A divided by multilateral budding and ascospores were not produced. Group B strains produced true hyphae and ascospores were not produced. Group C representatives divided by budding and formed chains and star-like aggregates. Ascospores were not produced. Sequence analysis of the 26S rRNA gene (D1/D2 region) revealed that the Group A isolates were phylogenetically most closely related to Saturnispora mendoncae (gene sequence similarity 92.4 %), Saturnispora besseyi (88.8 %), Saturnispora saitoi (88.8 %) and Saturnispora ahearnii (88.3 %). Members of Group B were most closely related to representatives of the genera Dipodascus and Galactomyces and the asporogenous genus Geotrichum, but in all cases with 26S rRNA gene (D1/D2 region) similarities below 87 %. For Group C, the most closely related species were Candida rugopelliculosa (92.4 %), Pichia occidentalis (91.6 %) and Pichia exiguza (91.9 %). The very low gene sequence similarities obtained for the three groups of isolates clearly indicated that they represented novel species. Repetitive Palindromic PCR (Rep-PCR) of the isolates and their closest phylogenetic relatives confirmed that the new isolates belonged to previously undescribed species. In conclusion, based on the genetic and phenotypic results, the new isolates were considered to represent three novel species, for which the names Candida halmiae (group A, type strain G3T = CBS 11009T = CCUG 56721T); Geotrichum ghanense (group B, type strain GB = CBS 11010T = CCUG 56722T) and Candida awuaii (group C, type strain G15T = CBS 11011T = CCUG 56723T) are proposed.

Cocoa beans, the principal raw material of chocolate, have to be fermented, dried and roasted to obtain the characteristic cocoa flavour and taste. The fermentation of cocoa is a microbiologically complex process involving the activities of yeasts, lactic acid bacteria and acetic acid bacteria with yeasts being particularly important during the initial phases of the fermentation process (Schwan & Wheals, 2004; Jespersen et al., 2005; Nielsen et al., 2007). During an investigation of the micro-organisms involved in the fermentation of cocoa beans, a number of yeast isolates with unusual pheno- and genotypic properties were isolated from malt extract agar (Nielsen et al., 2007). Preliminary pheno- and genotypic results showed that the isolates clustered in three distinct groups, all representing putatively novel yeast species. This study presents the morphological, biochemical and molecular characterization of these isolates. Group A consisted of a single isolate (G3T = CBS 11009T = CCUG 56721T), group B comprised two isolates [G6T (= CBS 11010T = CCUG 56722T) and G17] and group C contained four isolates [G15T (= CBS 11011T = CCUG 56723T), G68, G71 and G174]. All isolates were isolated during the first 12 h of cocoa fermentation and represented between 3 and 14 % of the total yeast biota at the time of sampling (Nielsen et al., 2007).

All isolates and reference strains were grown in YPG broth [yeast extract 5 g l⁻¹ (Merck), glucose 10 g l⁻¹, peptone 10 g l⁻¹, pH 5.6] at 25 °C for 2–14 days. For long-term storage, 20 % glycerol was added to the medium and cultures were stored at −80 °C.

The micro morphology of the isolates grown for 1–5 days in YM (yeast extract 3 g l⁻¹, malt extract 3 g l⁻¹, peptone

Abbreviation: Rep-PCR, repetitive palindromic polymerase chain reaction.

The GenBank/EMBL/DDBJ accession numbers for the 26S rRNA gene sequences of Candida halmiae sp. nov. G3T, Geotrichum ghanense sp. nov. G6T and Candida awuaii sp. nov. G15T are DQ466525, DQ466527 and EU876854, respectively.

Supplementary figures showing (GTG)₅-PCR fingerprints and corresponding dendrograms, derived from UPGMA linkage of correlation coefficients, for the three novel species are available with the online version of this paper.
5 g l⁻¹, glucose 10 g l⁻¹, pH 5.6) and YPG broth (25 °C) was determined using phase-contrast microscopy (Fig. 1). Colony morphology was determined after 4–14 days of growth on YGP and MYGP agar (25 °C). Spore formation was examined on YM agar, V8 agar, 5% malt agar and acetate agar (Robert et al., 2008) incubated for 4–28 days (20 and 25 °C). Spore formation was determined using bright-field microscopy. None of the isolates investigated produced ascospores.

The carbohydrate assimilation and fermentation patterns of the new isolates were determined following the protocol of Yarrow (1998) aided by use of the API ID32C identification system (bioMérieux). The results of these analyses are presented in Tables 1, 2 and 3 and the species descriptions.

For 26S rRNA gene sequencing, DNA was extracted using the Instagene Matrix DNA isolation kit following the manufacturer’s instructions (Bio-Rad). The D1/D2 region of the 26S rRNA gene was amplified using primers NL1 and NL4 as outlined previously (Jespersen et al., 2005).

Following purification (Qiagen PCR Purification kit), the fragments were sequenced in both directions by a commercial sequencing facility (DNA Technology, Aarhus, Denmark). Sequences were manually corrected and aligned using Vector NTI Suite 10 software (Informax). The closest phylogenetic relatives were determined by aligning the corrected sequences to 26S rRNA gene sequences in the GenBank database using the BLAST algorithm (Altschul et al., 1997).

Comparison of the similarity of the 26S rRNA gene sequence (D1/D2 region) of isolate G3T with type strains from the GenBank database revealed that isolate G3T was most closely related to Saturnispora mendoncae [26S rRNA gene (D1/D2 region) gene sequence similarity 92.4 %], Saturnispora besseyi (88.8 %), Saturnispora saitoi (88.8 %) and Saturnispora ahearnii (88.3 %). Isolates G6T and G17 were most closely related to Dipodascus albidus (86.2 %), Dipodascus geniculatus (85.3 %) and Dipodascus australiensis (84.7 %). Isolates G15T, G68, G71 and G174 were most closely related to Candida rugopelliculosa (92.3 %), Pichia occidentalis (91.6 %) and Pichia exigua (91.9 %). Until recently Pichia occidentalis and Pichia exigua were considered to belong to the genus Issatchenka. However, convincing phylogenetic evidence that the genus Issatchenka should be transferred to the genus Pichia was recently published (Kurtzman et al., 2008); this nomenclature is used in the present manuscript.

The 26S rRNA gene sequences (D1/D2 region) of the new isolates and their closest phylogenetic relatives retrieved from the GenBank database were aligned and phylogenetic trees were constructed by the neighbour-joining method.
Table 2. Key phenotypic characteristics of Geotrichum ghanense sp. nov. and its closest phylogenetic relatives

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Using BioNumerics version 4.5 (Applied Maths). Unknown bases were discarded for the analysis. The statistical reliability of the topology of the phylogenetic trees was evaluated using bootstrap resampling of the data (1000 resamplings) (Figs 2, 3 and 4).

Additionally, the isolates and their closest phylogenetic neighbours were genotypically investigated by Repetitive Element Palindromic (rep)-PCR using the primer GTG5 (5’-GTGTTGTTGTTGTTGTTG-3’) as previously described (Andrade et al., 2006; Nielsen et al., 2007). Isolate G3T (group A) was clearly separated (r=68%) from type strains representing the phylogenetically closest relatives (see Supplementary Fig. S1 in IJSEM Online). Isolates G6T and G17 (group B) clustered closely together (r>90%) and were only distantly related (r=52%) to type strains representing the phenotypically closest relatives (see Supplementary Fig. S2). Isolates G15T, G68, G71 and G174 (group C) formed two closely related (r=82%) but distinct groups with the four isolates clearly clustering (r=61%) away from the closest phylogenetic relatives (see Supplementary Fig. S3).

Based on the above data, it can be concluded that strain G3T is genetically and phenotypically distinct from all currently recognized species with S. mendoncae and S. ahearnii being the closest phylogenetic relatives. Phenotypically, strain G3T can be differentiated from the closest phylogenetic neighbours on the basis of its ability to assimilate cellobiose, D-xylose and maltose (slow reaction) as seen from Table 1. The results obtained in the present study clearly indicate that strain G3T represent a novel species that is genetically closely related to the genus Saturnispora. However, as ascospore production was not observed in the new isolate, the novel species must belong to the anamorphic ascomycetous yeasts. The name Candida halmaiae sp. nov. is proposed for the new isolate, with strain G3T (=CBS 11009T=CCUG 56721T) being the type strain.

Isolates G6T and G17 formed a genetically and phenotypically distinct group that was most closely related to D. albidus, D. geniculatus and D. australiensis but as ascospore formation was not observed, the isolates belong to the anamorphic ascomycetous yeasts. Phenotypically, the ability to ferment galactose and assimilate sucrose and trehalose differentiated strains G6T and G17 from their closest phylogenetic neighbours (Table 2). The results obtained in the present study clearly indicate that isolates G6T and G17 represent a novel species in the anamorphic genus Geotrichum, for which we propose the name Geotrichum ghanense sp. nov. with strain G6T (=CBS 11010T=CCUG 56722T) being the type strain.

Isolates G15T, G68, G71 and G174 formed a genetic and phenotypic distinct group phylogenetically closest related to C. rugopelliculosa, P. occidentalis and P. exigua. Phenotypically, the ability to ferment maltose and assimilate cellobiose and ribitol clearly differentiated strains G15T, G68, G71 and G174 from their closest phylogenetic neighbours (Table 3). In conclusion, the results obtained in the present study clearly indicate that isolates G15T, G68, G71 and G174 represent a novel species in the genus Candida, for which we propose the name Candida awuaii sp. nov. with strain G15T (=CBS 11011T=CCUG 56723T) being the type strain.

Latin diagnosis of Candida halmaiae Nielsen, Jakobsen et Jespersen sp. nov.

In liquido YM, post dies 1–3 ad 25 °C, cellulæ vegetativæ ovoideae (2.5–5.0 μm x 3.8–7.5 μm), singulae aut binæ. Per gemmationem multipolare reproducentes. In agaro MYGP post dies 4–6 ad 25 °C cultura 2–3 mm, candida et cremea,
butyrosa, glabra, infima-convexa et margo integra. Ascosporae non formantur. Glucosum et maltosum fermentatur. Sucrosum, D-galactosum, D-trehalosum, D-lactosum, D-raffinosum non fermentatur. Assimilantur D-glucosum, ribitolum (exigue), D-xylulosum (exigue), cellobiosum (exigue), D-mannitolum, maltosum (exigue), trehalosum (exigue), et DL-acidum lacticum. Non assimilatur L-sorbusum, sucrosum, D-galactosum, D-sorbitolium, D-ribosum, N-acetylglucosaminum, L-rhamnosum, L-arabinosum, erythritolium, D-raffinosum, D-melibiosum, D-melezitosum, 2-ketogluconatum, gluconatum, methyl D-pyranosidium, D-lactosum, inositolum et glycerolium. Non assimilatur nitratum. Aesculinum hydrolysatur. Crescentiae (exigue) 37°C. Typus G3\textsuperscript{T} (=CBS 11009\textsuperscript{T}=CCUG 56721\textsuperscript{T}).

**Description of Candida halmiae Nielsen, Jakobsen & Jespersen sp. nov.**

*Candida halmiae* (hal’mi.ae. N.L. fem. gen. n. *halmiae* of Halm, named in honour of Dr Mary Halm, a Ghanaian microbiologist who has contributed significantly to our understanding of the importance of yeasts in indigenous African fermented foods).

In YM and YGP broth after 1–3 days at 25°C, cells are ovoidal (2.5–5.0 μm × 3.8–7.5 μm), occur singly and in pairs, and divide by multilateral budding. On YPG and MYGP agar (4–6 days, 25°C) colonies are circular with a diameter of 2–3 mm, tannish-white, butyrous, smooth, glistering, low convex and with an entire margin. Formation of ascospores is not observed. Glucose and maltose are fermented. Sucrose, D-galactose, trehalose, lactose and raffinose are not fermented. Glucose, ribitol (weak), D-xylose (slow), cellobiose (slow), D-mannitol, maltose (slow), trehalose (weak) and DL-lactic acid are assimilated. L-Sorbose, sucrose, D-galactose, D-sorbitol, D-ribose, N-acetylglucosamine, L-rhamnose, L-arabinose, erythritol, raffinose, melibiose, melezitose, 2-ketogluconate, gluconate, methyl D-glucopyranoside, D-lactose, inositol and glycerol are not assimilated. Aesculin is hydrolysed. Nitrate is not assimilated. No growth in the presence of 0.01 % cycloheximide. Grows at 37°C, but growth is weak and delayed.

The type strain, G3\textsuperscript{T} (=CBS 11009\textsuperscript{T}=CCUG 56721\textsuperscript{T}), was isolated from cocoa fermentations in Tafo, Ghana. The description of the type strain corresponds to the description of the species.

**Latin diagnosis of Geotrichum ghanense Nielsen, Jakobsen et Jespersen sp. nov.**

In liquido YM, post dies 1–5 ad 25°C pellicula formantur et cellularae ovoidae aut elongatae (3.5–6.3 × 6.3–12.5 μm), singulae authyphae. Hyphae ramosae cum arthroconidiis formantur. In agaro MYGP post dies 4 ad 25°C cultura 5–10 mm, candida, farinosus, margine ciliata. Ascosporae non
formantur. Glucosum et D-galactosum fermentatur. Sucrosum, maltosum, D-trehalosum, D-lactosum, D-raffinosa-
num non fermentatur. Assimilantur D-glucosum, L-sorbosum, ribitolum (exigue), sucrosum (exigue), maltosum (exigue), D-
xylosum, cellobiosum, D-mannitolum, trehalosum (exique), D-
galactosum glyceralum et levulitum. Non assimilatatur D-
ribosum, N-acetylglucosaminum, L-rhamnosum, L-arabinosum,
D-raffinosum, D-melibiosum, gluconatum, D-melezitosum,
2-ketogluconatum, methyl D-pyranosidum, D-lactosum et
inositolum. Assimilantur DL-acidum lacticum variabile.
Non assimilatur nitratum. Aesculinum hydrolysatur.
Crescentiae 37 °C. Typus G6(=CBS 11010 T=CCUG 56722 T) non assimilatur DL-acidum lacticum.

Description of Geotrichum ghanense Nielsen,
Jakobsen & Jespersen sp. nov.

Geotrichum ghanense (ghan.en’se. N.L. neut. adj. ghanense pertaining to Ghana where the organism was first isolated).

A pellicle is formed in YM and YGP broth after 1–5 days at 25 °C. Cells are ovoid to elongate (3.5–6.3 × 6.3–12.5 μm)
occurring singly and as hyphae disarticulating into arthroconidia. On YPG and MYGP agar (4 days, 25 °C),
true mycelium with branching hyphae is produced. Colonies are 5–10 mm in diameter, whitish, dry, farinose and
with creeping hyphae. Formation of ascospores is not observed. Glucose and galactose are fermented. Sucrose,
trehalose, lactose, maltose and raffinose are not fermented. Glucose, L-sorbose, ribitol, sucrose (weak), maltose (weak),
D-xylode, cellobiosum, D-mannitolum, trehalosum (weak), galac-
tose, levulitum and glycerol are assimilated. D-Ribo, N-
acetylglucosamine, L-rhamnosum, L-arabinose, raffinose,
melibiosum, glyconate, melezitosum, 2-ketogluconate, methyl
D-glucopyranoside, lactose, inositol and glucosamine are
not assimilated. Assimilantur DL-lactic acid is strain
dependent. Nitrate is not assimilated. Aesculin is hydro-
lysed. Able to grow in the presence of 0.01 % cyclohex-
imide. Growth occurs at 37 °C.

The type strain, G6(=CBS 11010 T=CCUG 56722 T), and
all other known strains were isolated from cocoa fermentiones in Tafó, Ghana. The description of the type
strain corresponds to the description the species, except
that the type strain does not assimilate DL-lactic acid.

Latin diagnosis of Candida awuái Nielsen,
Jakobsen & Jespersen sp. nov.

In liquido YM, post dies 2–5 ad 25 °C, cellulae vegetativae
ovoidae (2.5–3.8 μm × 3.8–6.3 μm), singulae, binae et
agregatae. Per genmationem multipolarum reproductenses.
In agaro MYGP post dies 4 ad 25 °C cultura 2–3 mm,
candida et creame, butyrosa, convexa et margo non integra.
Ascosporae non formantur. Glucosum et maltosum fermenta-
tur. Sucrosum, D-galactosum, D-trehalosum, D-lactosum,
D-raffinosum non fermentatur. Assimilantur D-glucosum,
glycoroleum, D-xylolsum (exigue), cellobiosum (exigue),
L-sorbo, sucrosum (exigue), sucrosum (exique). Non assimilatatur D-
galactosum, N-acetylglucosaminum, D-sorbitolum, L-rham-
nonum, L-arabinosum, erythritolium, D-raffinosum, D-meli-
biosum, D-melezitosum, 2-ketogluconatum, gluconatum,
methygl-D-pyranosidum, D-lactosum, inositolum et glucosa-
minium. Assimilantur maltosum (exigue), ribitolum, D-
mannitolum et trehalosum variabile. Non assimilantur
nitratum. Aesculinum hydrolysatur. Crescentiae (exigue)
37 °C. Typus G15(=CBS 11011 T=CCUG 56723 T)
assimilantur ribitolum et trehalosum; D-mannitolum non
assimilatur.

Description of Candida awuái Nielsen, Jakobsen & Jespersen sp. nov.

Candida awuái (a.wuái’i. N.L. masc. gen. n. awuái of
Awua, named in honour of Dr Wisdom K. Amoa-Awua, a
Ghanaian microbiologist who has contributed significantly
to our understanding of the importance of yeasts in
indigenous African fermented foods).

In YM and YGP broth after 2–5 days at 25 °C, cells are ovoid
(2.5–3.8 μm × 3.8–6.3 μm), divide by multilateral budding,
occur singly, in pairs and form chains and star-like
aggregates. On MYGP agar (4 days, 25 °C) colonies are 2–3
mm in diameter, tannahish-white, circular, butyrous, convex
and with a slightly fringed margin. After 14 days the colony
edge is fringed and the centre of the colonies becomes
yellowish. Formation of ascospores is not observed. Glucose
and maltose are fermented. Sucrose, galactose, trehalose,
sorbose (weak), sucrose (slow), D-xylode (weak) and
glycerol are assimilated. Galactose, D-sorbitol, N-acetylglu-
cosamine, 1-rhamnose, 1-arabinose, erythritol, raffinose, melibiose, melezitose, 2-ketogluconate, gluconate, methyl D-glucopyranoside, lactose, inositol and glucosamine are not assimilated. Assimilation of maltose (slow), ribitol, D-mannitol (slow) and trehalose (slow) are strain dependent. Nitrate is not assimilated. No growth in the presence of 0.01 % cycloheximide. Growth occurs at 37 °C.

The type strain, G15T (=CBS 11011T=CCUG 56723T), and all other known strains of the species have been isolated from cocoa fermentations in Tafo, Ghana. The description of the type strain corresponds to the description the species except that ribitol (slow) and trehalose (slow) are assimilated and D-mannitol is not assimilated.

Acknowledgements

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References


