Selenomonas bovis sp. nov., isolated from yak rumen contents

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Two strictly anaerobic, crescent-shaped bacterial strains, designated WG⁷ and Ycb08, were isolated from a cellulose-degrading mixed culture enriched from yak rumen contents. The strains were Gram-negative, non-spore-forming and motile, with four to six flagella situated at the centre of the concave side of the cell. The cells were 0.9–1.1×4–6 µm. Growth was observed at 27–46 °C (optimum 39 °C) and pH 4.2–8.3 (optimum pH 7.0–7.2). Arabinose, glucose, mannose, cellobiose, lactose, sucrose, trehalose, melibiose, raffinose, salicin and aesculin were fermented. The end products of glucose fermentation were acetate, propionate and CO₂. The G+C contents of strains WG⁷ and Ycb08 were respectively 63.9±0.2 and 62.5±0.2 mol% (Tm). Phylogenetic analysis based on 16S rRNA gene sequences revealed that the two strains were related to the genera Mitsuokella and Selenomonas at similarity levels below 97%; however, they differed from members of the genus Mitsuokella in their flagellar arrangement. On the basis of phenotypic, genotypic and physiological evidence, strains WG⁷ and Ycb08 are identified as members of a novel species of the genus Selenomonas, for which the name Selenomonas bovis sp. nov. is proposed. The type strain is WG⁷ (=CGMCC 1.5073T =JCM 15470T).

The yak (Bos grunniens) is a ruminant found mainly on the Qinghai–Tibet Plateau, China, at altitudes over 3000 m above sea level, with grasses as their main food. Previous work in our laboratory (An et al., 2005) based on a culture-independent approach determined that more than 60 % of the bacteria in the yak rumen clustered in not-yet-cultured groups, displaying <90 % 16S rRNA gene sequence similarity with cultured species. In addition, a relatively large proportion of the species detected in the yak rumen grouped with fibrolytic bacteria isolated previously from rumen contents, suggesting that the yak rumen could harbour novel types of fibrolytic bacteria. Therefore, by using filter paper as the sole carbon source, we obtained a cellulose-degrading mixed-culture enrichment with yak rumen content as inoculant; a few fibrolytic and non-fibrolytic bacterial strains were isolated from the mixed culture. In this study, we describe two non-fibrolytic bacteria strains that represent a novel species of the genus Selenomonas.

Yak rumen content was inoculated into a modified basal medium (Hungate, 1966) with filter paper as the sole carbon source and grown under a gas phase of N₂/CO₂ (80:20). The medium contained 20 % rumen fluid and 80 % basal medium. A fibrolytic mixed culture was obtained by subculturing the enrichment in the same medium for eight to ten transfers. After serial dilution in peptone-yeast extract-glucose (PYG) broth (Holdeman et al., 1977) and the use of the Hungate roll-tube technique (Hungate, 1969), single colonies were observed after incubation at 39 °C for 2 days. Colonies were picked and transferred to the same medium. The roll-tube procedure was repeated several times before strains WG⁷ and Ycb08 were obtained. The purity of the isolates was examined by light microscopy. All inoculations and transfers were done with syringes and needles, and the cultures were incubated at 39 °C unless indicated. Selenomonas ruminantium subsp. ruminantium DSM 2150T, S. ruminantium subsp. lactilytica DSM 2872T, Selenomonas sputigena DSM 20758T and Mitsuokella jalaludinii DSM 13811T were purchased from the Deutsche Sammlung von Mikroorganismen und Zellkulturen (Braunschweig, Germany) and served as reference strains in physiological characterization tests.

Cell morphology was examined under a light microscope (Olympus BH-2) and an electron microscope (Hitachi H-600A). For electron microscopy studies, bacterial cells were negatively stained with uranyl acetate. Spore formation was examined by phase-contrast microscopy at the end of growth as well as by heat treatment in a water bath at 80 °C for 10 min.

The GenBank/EMBL/DDBJ accession numbers for the 16S rRNA gene sequences of strains WG⁷ and Ycb08 are respectively EF139191 and EU821596.

A transmission electron micrograph of a cell of strain WG⁷ is available as supplementary material with the online version of this paper.

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Cells were Gram-negative, typical curved rods, 0.9–1.1 × 4–6 μm, and occurred singly or in pairs. Spores were never observed, and the strains could not survive heat treatment at 80 °C for 10 min. Negatively stained cells revealed the presence of four to six flagella in a tuft situated near the centre of the concave side (Supplementary Fig. S1, available in IJSEM Online).

Growth was determined by monitoring the OD600 of the culture in PYG medium at 39 °C. The strains were strictly anaerobic; no growth occurred when exposed to air. The generation time of strain WGT was determined as 2.3 h by monitoring the OD600 at 30 min intervals for 24 h. The temperature profile of strain WGT was determined in a water bath from 20 to 50 °C at 1 °C intervals (pH 7.0) and growth was observed between 27 and 46 °C, with optimal growth at 39 °C. The pH range for growth was pH 4.2–8.3, with optimal growth around pH 7.0–7.2, as determined in PYG broth at various pH values adjusted with Na2HPO4 and NaH2PO4.

The end products from glucose fermentation were determined by growing strains WGT and Ycb08 in PYG medium; short-chain fatty acids were then analysed by using a gas chromatograph (GC-14B; Shimadzu) equipped with a flame-ionization detector (column temperature 230 °C, injector 250 °C, detector 280 °C). The carrier gas was N2 at a flow rate of 30 ml min⁻¹. It was determined that strain WGt and Ycb08 produced acetate, propionate and CO2 from glucose at a molar ratio of 0.7:1.1:1.2.

Substrate utilization was tested by growing the two strains in PY medium (Holdeman et al., 1977) separately containing each of the following substrates: lactate, glycerol, dulcitol, mannitol, sorbitol, xylose, arabinoise, glucose, mannose, cellobiose, lactose, sucrose, trehalose, melibiose, raffinose, inulin, salicin and aesculin. Stock solutions of the test compounds were prepared anaerobically, sterilized by filtration and injected into PY medium to the final concentration of 1 % (w/v). Other physiological characteristics including the Voges–Proskauer test, nitrate reduction, indole production, H2S production, methyl red test and gelatin liquefaction were determined according to Holdeman et al. (1977).

The two strains exhibited almost identical physiological and biochemical profiles except that strain Ycb08 could not ferment inulin or raffinose. Other physiological characteristics are listed in the species description.

Genomic DNA was extracted and purified according to Marmur (1961). The G+C content of the DNA was determined by the thermal denaturation method (Marmur & Doty, 1962) using a DU800 spectrophotometer (Beckman) with Escherichia coli K-12 DNA as the reference. The G+C contents of the genomic DNA for strains WGt and Ycb08 were determined as 63.9 ± 0.2 and 62.5 ± 0.2 mol%, respectively, from three parallel measurements.

The 16S rRNA gene was amplified using the bacterial universal primers 27F and 1541R (Weisburg et al., 1991), Purified PCR products approximately 1.5 kbp in length were cloned to Escherichia coli DH5α and sequenced by the Sangon Biological Engineering Technology Service (Shanghai, China). The sequences were submitted to GenBank to search for similar sequences using BLAST algorithm. The 16S rRNA gene sequences of strain WGt and reference strains retrieved from GenBank were aligned using the CLUSTAL X program v. 1.83 (Thompson et al., 1997). Phylogenetic trees were constructed using the neighbour-joining, UPGMA, minimum evolution and maximum-parsimony methods as implemented in MEGA3 (Kumar et al., 2004). Stability of groupings was evaluated by bootstrap analysis of 1000 datasets (Felsenstein, 1985). Strains WGt and Ycb08 displayed 16S rRNA gene sequence similarity of 99.7 %; they were therefore considered to represent a single species. To ascertain the phylogenetic position of the new isolates, the complete 16S rRNA gene sequences were compared with the most similar sequences retrieved from GenBank. On the basis of a consensus 1379 bp 16S rRNA gene sequence, a phylogenetic tree rooted with Bacillus subtilis DSM 10T was constructed (Fig. 1). It was shown that the two strains clustered with the members of the genera Selenomonas and Mitsuokella and were closest to M. jalaluddini M 9T (95.5 % 16S rRNA gene sequence similarity), followed by S. ruminantium subsp. ruminantium GA192T (95.4 %), S. ruminantium subsp. lactilytica HD4 (95.2 %) and Mitsuokella multacida NCTC 10934T (95.1 %). According to the well-accepted bacterial species delimitation of less than 97 % 16S rRNA gene sequence similarity (Wayne et al., 1987), strains WGt and Ycb08 could represent a novel species affiliated either to the genus Selenomonas or Mitsuokella.

Based on the genus descriptions, members of Mitsuokella are characterized by a cell morphology of stout rods without flagella (Mitsuoka et al., 1974; Holdeman et al., 1984; Lan et al., 2002), while cells of the genus Selenomonas are curved to helical rods, motile with a typical flagellar arrangement as a tuft near the centre of the concave side (Bryant, 1984; Kingsley & Hoeniger, 1973). Strain WGt presented crescent-shaped cells with flagella on the concave side; together with other differential characteristics listed in Table 1, the two strains can be classified in the genus Selenomonas. Furthermore, the two strains resembled other members of the genus Selenomonas in metabolic features, such as the use of a variety of carbohydrates as substrates and the production of acetate, propionate and CO2 from glucose, as well as production of H2S from L-cysteine but the absence of nitrate reduction (Table 1). However, strains WGt and Ycb08 displayed <97 % 16S rRNA gene sequence similarity with described Selenomonas species and differed from their closest relative, S. ruminantium subsp. ruminantium DSM 2150T, in their higher G+C content and by liquefying gelatin but not fermenting dulcitol, mannitol, sorbitol and xylose. Thus, on the basis of DNA G+C content and the physiological and phylogenetic traits described above, strains WGt and Ycb08 represent a novel species.
species of the genus *Selenomonas*, for which the name *Selenomonas bovis* sp. nov. is proposed.

**Description of Selenomonas bovis sp. nov.**

*Selenomonas bovis* (bo'vis. L. gen. n. bovis of a cow, of a bovine, referring to the isolation of the first strains from yak rumen contents).

Gram-negative, crescent-shaped rods, 4–6 μm long and 0.9–1.1 μm wide. No spore formation is observed. Flagella (four to six) are arranged linearly as a tuft near the centre of the concave side of the cell. The temperature range for growth is 27–46 °C; optimum growth at 39 °C. Grows at pH 4.2–8.3, with optimum growth at pH 7.0–7.2. The generation time is 2.3 h when grown on glucose at 39 °C. The final pH in PYG medium is 4.5. Utilizes arabinose, glucose, mannose, cellobiose, lactose, sucrose, trehalose, melibiose, raffinose, salicin and aesculin but not lactate, xylose, starch, dulcitol, mannitol or sorbitol. The end products of glucose fermentation are acetate, propionate and CO₂ according to the equation 1 glucose → 0.7 acetate + 1.1 propionate + 1.2 CO₂. liquefies gelatin and

**Table 1. Differential characteristics of strains WGᵀ and Ycb08 and phylogenetically related type strains**

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<td>Isolation source</td>
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<td>Yak</td>
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<td>Cattle</td>
<td>Human</td>
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<tr>
<td>Morphology*</td>
<td>CR</td>
<td>CR</td>
<td>CR</td>
<td>CR</td>
<td>CR</td>
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<td>Voges–Proskauer test</td>
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<td>Nitrate reduction</td>
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<td>H₂S production</td>
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<td>Gelatin liquefaction</td>
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<td>Acid production from:</td>
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<tr>
<td>DNA G+C content (mol%)</td>
<td>63.9 ± 0.2</td>
<td>62.5 ± 0.2</td>
<td>54.2 ± 0.2</td>
<td>55.2 ± 0.4</td>
<td>61</td>
<td>56.8</td>
<td>57.3</td>
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*CR, Curved rods; SR, stout rods.
†LA, Linearly arranged as a tuft near the centre of the concave side; NF, no flagella.
‡Data from this study.
produces H₂S but not indole. Does not reduce nitrate. Voges–Proskauer and methyl red tests are negative. The G+C content of the genomic DNA of the type strain is 63.9 ± 0.2 mol%.

The type strain, WC₂ (＝CGMCC 1.5073T = JCM 15470T), and a second strain, Ycb08, were isolated from yak rumen contents.

Acknowledgements

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References


