Spirochaeta cellobiosiphila sp. nov., a facultatively anaerobic, marine spirochaete

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A facultatively anaerobic, marine spirochaete, designated strain SIP1T, was isolated from interstitial water from a cyanobacteria-containing microbial mat. Cells of strain SIP1T were 0.3–0.4×10–12 μm in size, helical with a body pitch of approximately 1.4 μm and motile by means of two to four periplasmic flagella (one, or occasionally two, being inserted near each end of the cell). Cells were catalase-negative and used a variety of monosaccharides and disaccharides and pectin as energy sources, growing especially well on cellobiose. Neither organic acids nor amino acids were utilized as energy sources. One or more amino acids in tryptone and one or more components of yeast extract were required for growth. Growth was observed at 9–37 °C (optimally at or near 37 °C), at initial pH 5–8 (optimally at initial pH 7.5) and in media prepared with 20–100 % (v/v) seawater (optimally at 60–80 %) or 0.10–1.00 M NaCl (optimally at 0.30–0.40 M). The products of cellobiose fermentation were acetate, ethanol, CO2, H2 and small amounts of formate. Aerated cultures performed incomplete oxidation of cellobiose to acetate (and, presumably, CO2) plus small amounts of ethanol and formate, but exhibited a Ycellobiose that was only slightly greater than that of cellobiose-fermenting anoxic cultures. The G+C content of the genomic DNA of strain SIP1T was 41.4 mol%, the lowest among known spirochaetas. On the basis of its 16S rRNA gene sequence, strain SIP1T was grouped among other members of the genus Spirochaeta, but it bore only 89 % similarity with respect to its closest known relatives, Spirochaeta litoralis and Spirochaeta isovalerica, two marine obligate anaerobes. On the basis of its phenotypic properties and phylogenetic position, strain SIP1T represents a novel species of the genus Spirochaeta, for which the name Spirochaeta cellobiosiphila sp. nov. is proposed. The type strain is SIP1T (= ATCC BAA-1285T = DSM 17781T).

The genus Spirochaeta consists of more than a dozen species of obligately or facultatively anaerobic, saccharolytic, generally free-living spirochaetes found in aquatic habitats. Included are freshwater, marine and halophilic species, as well as alkaliphiles and thermophiles (Canale-Parola, 1984, 1992; Aksinova et al., 1992; Hoover et al., 2003; and references therein). Although most known spirochaetas are coiled or undulate in shape and are motile by means of periplasmic flagella, non-motile coccoid types have also been recognized as free-living forms (Ritalahti & Löfler, 2002, 2003, 2004) and as symbionts in the hindguts of termites (Dröge et al., 2006) and in the excretory organ of a Nautilus species (Pernice et al., 2007).

During the Microbial Diversity Summer Course at the Marine Biological Laboratory, Woods Hole (MA, USA) in 2004, selective isolation of marine spirochaetas was included as a laboratory exercise. This process was performed using serial dilution of inocula in anoxic, rifampicin-containing agar medium (Weber & Greenberg, 1981), as well as a technique involving placement of filter discs on agar plates (Canale-Parola, 1973). Inasmuch as all known marine spirochaetas were obligate anaerobes, special attention was paid to the possible development of spirochaete colonies on plates of a non-reduced isolation medium incubated in air, in that such colonies might represent novel organisms. Such colonies appeared and a facultatively anaerobic, marine member of the genus Spirochaeta was isolated that was sufficiently different from other members of the genus to be considered as representing a novel species. This paper describes the
isolation and properties of this strain, which was designated SIP<sub>T</sub>.

Isolation was performed on plates containing SCTY isolation medium [comprising, l<sup>-1</sup>: 700 ml seawater, 2.0 g cellobiose, 2.0 g tryptone (Difco), 1.0 g yeast extract (Difco), 10.0 g agar (Difco) and 300 ml distilled water] adjusted to pH 7.5 prior to autoclaving. Four sterile filter discs (mixed esters of cellulose; 0.45 µm mean pore diameter; Millipore) were placed on each plate and the centre of each disc was inoculated with approximately 0.5 ml marine sample. Plates were incubated (filter side up) for 8 h in air at 30 °C, after which the filter discs were removed aseptically; the plates were then turned upside down and incubated again. After 5 days, subsurface, semi-transparent, pale-yellow and white spreading colonies were observed on a plate inoculated with water that had been expressed, by gentle pressure, from a portion of a cyanobacteria-containing microbial mat collected from Little Sippewissett salt marsh, Woods Hole (MA, USA). Phase-contrast microscopy of a thin slice of agar from the advancing margin of a pale-yellow subsurface colony revealed cells with spirochaete-like morphology and motility; similar samples from the white colonies revealed thin, rapidly motile rods. Additional samples from the pale-yellow colony were excised, macerated on the surfaces of fresh plates and streaked for the purposes of isolation. After three successive subcultures had been performed using well-isolated colonies (each being composed of spirochaetes possessing uniform cell morphology), the culture was considered pure and was designated strain SIP<sub>T</sub>. Analogous isolation plates inoculated with surficial sediment water from Eel Pond, Oyster Pond Inlet and the Great Sippewissett salt marsh (all at Woods Hole) did not yield spirochaete colonies.

Strain SIP<sub>T</sub> was routinely maintained at room temperature (22–23 °C), under air, in 15 mm screw-capped tubes approximately two-thirds full with SCTY broth (as above, but lacking agar) or CTY broth. CTY broth contained the following (l<sup>-1</sup>): 2.0 g each of cellobiose and tryptone, 1.0 g yeast extract, 20.45 g NaCl (equivalent to 0.35 M); 0.52 g KCl, 4.31 g MgSO<sub>4</sub>·7H<sub>2</sub>O, 3.56 g MgCl<sub>2</sub>·6H<sub>2</sub>O, 0.18 g K<sub>2</sub>HPO<sub>4</sub>, 2.38 g HEPES buffer and 1.03 g CaCl<sub>2</sub>·2H<sub>2</sub>O. All of the ingredients in CTY medium (except CaCl<sub>2</sub>·2H<sub>2</sub>O) were dissolved in 990 ml distilled water and the solution was adjusted to pH 7.5 prior to being sterilized by autoclaving. When cooled, 10 ml (l<sup>-1</sup>) 100 × sterile stock solution of CaCl<sub>2</sub>·2H<sub>2</sub>O (i.e. 102.9 g l<sup>-1</sup>) was added. The final pH of the CTY medium was 7.3–7.4. Routine subculture was achieved by means of loop transfer every 3–4 days. Growth studies designed to determine the physiological properties and nutritional requirements of cells were performed using 10 ml medium in 18 mm diameter tubes incubated upright, in air, without shaking. Inocula (0.1 %, v/v) usually consisted of cells growing in CTY medium at room temperature. Salt requirements and salt tolerance were evaluated by using SCTY broth or CTY broth and varying the seawater or NaCl content in 10 % (v/v) or 0.10 M increments, respectively. The pH optimum and range were determined by adjusting CTY broth to various initial pH values and filter-sterilizing the medium prior to inoculation. Nutritional requirements were evaluated by using CTY broth in which cellobiose, tryptone or yeast extract were omitted or replaced with other compounds. Growth was quantified by determining the OD<sub>600</sub> with a Spectronic 20 colorimeter (Bausch & Lomb) and the readings were converted into cell number, cell protein or cell dry mass equivalents by using standard curves correlating these quantities (Graber et al., 2004). Aerated (oxic) cultures were grown in Erlenmeyer flasks (each equipped with a side arm allowing OD<sub>600</sub> readings to be taken) one-fifth filled with medium; the flasks were shaken on a rotary shaker operating at 120 r.p.m. Anoxic cultures were grown in 18 mm anaerobe tubes (or in bottles each fitted with an 18 mm anaerobe tube as a side arm) sealed under N<sub>2</sub> with butyl rubber stoppers as described previously (Wertz & Breznak, 2007). Soluble metabolic products were determined by means of HPLC with refractive index and UV detection, while H<sub>2</sub> was determined by GC; both procedures were performed as described previously (Graber et al., 2004). CO<sub>2</sub> was determined gravimetrically (Breznak & Canale-Parola, 1969). HPLC determination of the G+C content of genomic DNA and phase-contrast photomicroscopy were performed as described previously (Graber et al., 2004; Wertz & Breznak, 2007). Transmission electron micrographs were prepared from cells negatively stained with 1 % ammonium molybdate and examined with a JEOL JEM-100C XII electron microscope operating at an accelerating voltage of 100 kV.

Cells of strain SIP<sub>T</sub> were 0.3–0.4 µm in diameter and 10–12 µm long (range, 5–40 µm) with slightly tapered ends (Fig. 1a, c); they appeared to be helical, with a mean body pitch of approximately 1.4 µm (as measured from phase-contrast micrographs). Cells also possessed ultrastructural features typical of most known spirochaetes, including the following: a protoplasmic cylinder; two to four periplasmic flagella (one, occasionally two, being inserted near each end of the protoplasmic cylinder); and an outer membranous sheath (Fig. 1b). Cells were motile, with spinning and flexing movements, and gave rise to spherical bodies (Breznak & Canale-Parola, 1975) that were either free or attached to the spiral cells and which became more numerous when cells entered stationary phase and as cultures aged and lost viability.

In SCTY medium, strain SIP<sub>T</sub> grew best if seawater comprised 60–80 % (v/v; balance, distilled water) of the liquid phase (range, 20–100 % seawater). In CTY medium, cells grew best with 0.30–0.40 M NaCl (range, 0.10–1.00 M), at initial pH 7.5 (range, pH 5.0–8.0) and at, or near, 37 °C (range, 9–37 °C). No growth was observed at 42 °C. Under optimum conditions, the doubling time (t<sub>d</sub>) of cells in unshaken tubes of CTY medium under air was 1.9 h (mean of two determinations). At 30 °C (the temperature used for most experiments), cells grew only
slightly more slowly, whether in unshaken tubes \( (t_d = 2.2 \pm 0.2 \text{ h}; \text{mean} \pm \text{SD for four determinations}) \) or in shake flasks \( (t_d = 2.1 \text{ h}; \text{mean of two determinations}) \). Cells of strain SIP1\(^T\) harvested from aerated cultures in CTY medium were catalase-negative and pale yellow in colour, but gave a negative reaction in the carotenoid spot test (Breznak & Canale-Parola, 1969). Likewise, acetone/methanol (70:30, v/v) extracts of cell pellets did not possess a visible wavelength spectrum typical of that of carotenoids (Zechmeister, 1962; data not shown). The NaCl requirement for optimum growth of strain SIP1\(^T\) was similar to that for other mesophilic, neutrophilic, marine spirochaetas, e.g. *Spirochaeta litoralis, Spirochaeta isovalerica* and *Spirochaeta bajacaliforniensis*, all of which are obligate anaerobes, but it was about half that required for optimum growth of *Spirochaeta halophila*, a carotenoid-pigmented, facultative anaerobe isolated from a high-salinity pond, and *Spirochaeta smaragdinae*, an obligate anaerobe isolated from an offshore oilfield (Table 1). In contrast, the facultatively anaerobic, freshwater spirochaete *Spirochaeta aurantia* subsp. *aurantia* J1\(^T\) (=ATCC 25082\(^T\); Breznak & Canale-Parola, 1969), used here as a control, grew best in CTY medium with no added NaCl (range, 0–0.10 M NaCl). Similar results were reported previously for *S. aurantia*, as well as for the freshwater, anaerobic spirochaete *Spirochaeta stenostrepta* (Hespell & Canale-Parola, 1970).

The yields of strain SIP1\(^T\) in tubes of CTY medium were 5–7 \times 10^8 \text{ cells ml}^{-1}. In CTY medium lacking tryptone, the cell yields were only about 25% of that normally attained, whether or not 5 mM NH4Cl was included in the medium. The cell yields in CTY medium lacking yeast extract were only about 10% of that normally attained; little or no growth was observed in CTY medium lacking cellobiose. These results implied that cellobiose was used as a primary energy source by strain SIP1\(^T\) and that one or more of the amino acids in tryptone was required for growth, as was one or more components present in yeast extract. Cellobiose, when present at 5 mM in static culture tubes of CTY medium at 30 °C, supported a doubling time of 2.2 h. A variety of other monosaccharides and disaccharides (tested at 10 and 5 mM, respectively), as well as pectin (Sigma-Aldrich; tested at 0.2%, w/v), were also used as energy sources by strain SIP1\(^T\), but the doubling times at 30 °C were substantially longer than those obtained with cellobiose, e.g. glucose, 4.5 h; galactose, 5.2 h; d-xylene, 7.4 h; l-arabinose, 7.4 h; sucrose, 17.0 h; mannose, 17.8 h; trehalose, 18.8 h; maltose, 19.2 h; and pectin, 162.0 h. Little or no growth above that of ‘no cellobiose’ controls occurred with fructose, l-fucose, l-rhamnose, mannitol, D-ribose, arabininoxylan (0.2%, w/v; Megazyme), cellulose (0.5 x 2 cm strip of Whatman No. 4 filter paper), acetate, lactate, citrate or succinate (organic acids added as sodium salts, 5.0 mM final concentration; tested under both anoxic and aerated culture conditions). Hence, strain SIP1\(^T\) was typical of most other known spirochaetas in being saccharolytic and unable to use organic acids or amino acids as energy sources for growth. The ability of strain...
Table 1. Characteristics of strain SIP1T that serve to distinguish it from mesophilic, neutrophilic, marine and halophilic spirochaetas

<table>
<thead>
<tr>
<th>Characteristic</th>
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<tbody>
<tr>
<td>Cell size (μm)</td>
<td>0.3–0.4 × 10–12</td>
<td>0.4–0.5 × 6–7</td>
<td>0.4 × 10–15</td>
<td>0.3 × 30</td>
<td>0.4 × 15–30</td>
<td>0.3–0.5 × 5–30</td>
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<td>Optimum growth conditions</td>
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<td>NaCl concentration (M)</td>
<td>0.3–0.4</td>
<td>0.35</td>
<td>0.2–0.3</td>
<td>0.48</td>
<td>0.75</td>
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<td>pH</td>
<td>7.5</td>
<td>7.0–7.5</td>
<td>NR</td>
<td>7.5</td>
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<tr>
<td>Temperature (°C)</td>
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<td>36</td>
<td>35–40</td>
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<td>Substrate utilization</td>
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<td>Sucrose</td>
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<td>O2 metabolism*</td>
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<td>OA</td>
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<tr>
<td>DNA G + C content (mol%)</td>
<td>41.4</td>
<td>50.5a</td>
<td>44.2b†</td>
<td>63.6–65.6</td>
<td>50.1</td>
<td>62</td>
</tr>
</tbody>
</table>

*FA, Facultative anaerobe; OA, obligate anaerobe.
†Results obtained using the buoyant density method by Hespell & Canale-Parola (1970) (a) and using the Tm method by Breznak & Canale-Parola (1975) (b).

SIP1T to use amino acids such as l-leucine, l-isoleucine and l-valine as energy sources for maintenance and survival (but not growth), as demonstrated by S. isovalerica (Harwood & Canale-Parola, 1983), was not explored. However, no branched-chain fatty acids resulting from the metabolism of such amino acids were detected in cellbiose-limited anoxic or aerated cultures (below).

Products formed during anoxic growth of strain SIP1T in CTY medium containing cellbiose at 2.5 mM (a concentration determined empirically to be growth-limiting for both anoxic and aerated cultures) were as follows (mol per 100 mol glucosyl unit of cellbiose): acetate, 80.1; ethanol, 69.4; CO2, 152.3; H2, 183.7; and formate, 29.9 (carbon recovery 80.2%; oxidation/reduction index 1.04). These fermentation products are among those typically produced by spirochaetes of the genus Spirochaeta (Canale-Parola, 1984, 1992; Hoover et al., 2003 and references therein; Dröge et al., 2006). No other soluble or gaseous products were detected by HPLC or GC, respectively. These results imply that nearly 20% of the cellbiose carbon may have been assimilated by the cells. This degree of assimilation, although apparently high for the conditions employed (i.e. during energy-source–limited anoxic growth of cells in a complex medium), is nevertheless that of maltose-carbon assimilation by S. aurantia during anoxic growth under comparable conditions (16.1–17.5%; Breznak & Canale-Parola, 1972). Soluble products formed by strain SIP1T during aerobic growth on 2.5 mM cellbiose were as follows (mol per 100 mol glucosyl unit): acetate, 122.4; ethanol, 11.0, and formate, 2.9. These results suggest that strain SIP1T, like aerated cultures of S. aurantia (Breznak & Canale-Parola, 1972) and S. halophila (Greenberg & Canale-Parola, 1976), uses oxygen as an electron acceptor to perform incomplete oxidation of carbohydrate, mainly to acetate (and presumably CO2, which was not measured). Consistent with this interpretation was the observation that strain SIP1T, like S. aurantia and S. halophila, did not use acetate, lactate or tricarboxylic acid cycle intermediates as energy sources for aerobic growth (above), implying that it, too, lacks a complete tricarboxylic acid cycle. However, the Ysubstrate values for S. aurantia and S. halophila grown under aerated conditions were 2- to 3.4-fold greater than those obtained under anoxic conditions, and both of these species appear to be capable of conserving energy by oxidative phosphorylation during incomplete aerobic oxidation of carbohydrates. In contrast, Ycellbiose values for aerated cultures of strain SIP1T (approx. 98.3 g dry mass per mol cellbiose) were only 1.2-fold greater than those for anoxic cultures (approx. 79.2 g dry mass per mol cellbiose). This may simply reflect a modest gain in net ATP yield from substrate-level phosphorylation (via the acetate kinase reaction) accompanying the incremental increase in acetate produced (per mol cellbiose) under aerated conditions, rather than a genuine capacity for oxidative phosphorylation. Nevertheless, even modest gains in energy resulting from the presence of oxygen may be of competitive or survival value to strain SIP1T in situ and could be obtained, in part, during oxygenic photosynthesis of cyanobacteria and algae in the salt-marsh microbial mats it inhabits.

The 16S rRNA gene of strain SIP1T was amplified by using a PCR with primers 27F (5′-AGAGTTTGATCCTGGC-
The results of this study indicate that strain SIP1T is readily distinguishable from recognized spirochaetas on the basis of physiological and nutritional properties, as well as from the G+C content of its genomic DNA, which is the lowest among known spirochaetas (Table 1; Canale-Parola, 1984, 1992; Aksinova et al., 1992; Hoover et al., 2003; Dröge et al., 2006; and references therein). Moreover, the 16S rRNA gene sequence of strain SIP1T is phylogenetically distinct from those of all recognized spirochaetas (Fig. 2) and bears only 89% similarity with the sequences of its closest known relatives, S. litoralis and S. isovalerica. While such low levels of similarity might prompt the proposal of a novel genus for strain SIP1T, not merely a novel species, there are several reasons why this is not advisable. Doing so would imply that other species within the tree shown in Fig. 2 should also be assigned to a novel genus/genera, as the mean similarity of all Spirochaeta strains and clones depicted in Fig. 2 is only 86 ± 3% (mean ± SD), the lowest similarity among named species (81%) occurring between S. litoralis and Spirochaeta cocoides. Even among the free-living neutrophilic marine and halophilic spirochaetas (Table 1), the mean interspecies similarity is only 88 ± 3%. Moreover, if such reassignment were contemplated it would still be difficult to decide which species should correctly remain in the genus Spirochaeta, as no 16S rRNA gene sequence is available for the uncultured species Spirochaeta plicatilis, the type species of the genus (Canale-Parola, 1984).

Clearly, the genus Spirochaeta is a good candidate for revision, an effort that would be best achieved by using a comprehensive polyphasic taxonomic approach facilitated by the acquisition of 16S rRNA gene sequence data for the type species, S. plicatilis. However, that is beyond the scope of the present communication. Accordingly, and until any such future revision dictates otherwise, strain SIP1T represents a novel species of the genus Spirochaeta, for which the name Spirochaeta cellulosiphila sp. nov. is proposed.

Description of Spirochaeta cellulosiphila sp. nov.

Spirochaeta cellulosiphila (cel.lo.bi.o.si’ phi.la. N.L. neut. n. cellulosium cellibiose; N.L. fem. adj. phila from Gr. fem. phi (phi) love)

![Fig. 2. Maximum-likelihood phylogenetic tree, based on 16S rRNA gene sequences, for strain SIP1T. Bootstrap percentages are shown at nodes, as follows: filled circles, 75–100%; open circles, 50–74%. GenBank accession numbers are shown in parentheses. The black shape at the bottom represents the phylogenetic breadth and depth of 16S rRNA gene sequences from selected Brevinema andersonii strains (GenBank accession numbers M69179.1, L31544.1 and L31543.1) and Brevinema environmental clones (DQ340184.1 and AF027047.1), which were used collectively as an outgroup. Bar, 10% estimated sequence divergence.](Image)
adj. philè friendly to, loving; N.L. fem. adj. cellobiosiphila
loving cellobiose).

Cells are pale yellow, 0.3–0.4 × 10–12 μm in size and helical, with a body pitch of 1.4 μm. Motile by means of
two (occasionally four) periplasmic flagella, of which one
(or two) is inserted near each end of the cell. Facultatively
anaerobic and catalase-negative. Growth occurs at 9–37 °C
(optimally at or near 37 °C), at initial pH 5–8 (optimally at
initial pH 7.5) and in media prepared with 20–100 % (v/v)
seawater (optimally at 60–80 %) or containing 0.10–
1.00 M NaCl (optimally at 0.30–0.40 M). A variety of
monosaccharides and disaccharides and pectin (but not
cellobose or arabinoxylan) are used as energy sources; the
most rapid growth occurs on cellobiose. Neither organic
acids nor amino acids are utilized as energy sources. One or
more amino acids in tryptone and one or more components
of yeast extract are required for growth. The products of
cellobiose fermentation are acetate, ethanol, CO2, H2 and small amounts of formate. Aerated cultures
oxidize cellobiose incompletely to acetate (and, presum-
ably, CO2) plus small amounts of ethanol and formate; they exhibit a YC\textsubscript{cellobiose} that is 1.2-fold greater than that of
cellobiose-fermenting cultures. The G+C content of the
genomic DNA of the type strain is 41.4 mol% (HPLC).

The type strain, SIP1\textsuperscript{T} (=ATCC BAA-1285\textsuperscript{T} =DSM 17781\textsuperscript{T}), was isolated from interstitial water of a cyano-
bacteria-containing microbial mat collected from Little
Sippewissett salt marsh, Woods Hole, MA, USA.

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