Thioflavicoccus mobilis gen. nov., sp. nov., a novel purple sulfur bacterium with bacteriochlorophyll b

Johannes F. Imhoff¹ and Norbert Pfennig²

Author for correspondence: Johannes F. Imhoff. Tel: +49 431 697 3850. Fax: +49 431 56876.
e-mail: jimhoff@ifm.uni-kiel.de

INTRODUCTION

The first purple sulfur bacterium found to contain bacteriochlorophyll b as the major photosynthetic pigment and with the main in vivo absorption maximum at 1020 nm was described as a Thiococcus species (Eimhjellen al., 1967). This bacterium differed from all other photosynthetic bacteria on the basis of the internal photosynthetic membranes of tubular structure. The cells were non-motile cocci, contained sulfur globules, and did not contain gas vesicles. Strains of Thiococcus were isolated from sediments of lakes and rivers but also from marine habitats containing sulfide. This bacterium was included, later, in the genus Thiocapsa Winogradsky (1888) as Thiocapsa pfennigii (Eimhjellen, 1970). However, on the basis of 16S rDNA sequences, it was significantly different from the type species of this genus, Thiocapsa roseopersicina, and was reclassified as Thiococcus pfennigii (Imhoff et al., 1998). Only a few strains of the Chromatiaceae containing bacteriochlorophyll b have been isolated so far, such as the recently described bacteria from alkaline soda lakes in Siberia, which were described as Thioalkalicoccus limnaeus (Bryantseva et al., 2000).

In sandy intertidal sediments of the Great Sippewissett Salt Marsh on Cape Cod (MA, USA), flat, laminated microbial mats of unusual thickness regularly developed. During the sampling period in the summer of 1986, these mats were about 10 mm thick and comprised four to five distinctly coloured layers (Nicholson et al., 1987). Phototrophic purple sulfur bacteria from the central pink layer and the peach-coloured layer immediately below were analysed. The bacteria of the peach layer consisted of small spirilloid bacteria that were classified as Rhodospira trueperi (Pfennig et al., 1997), as well as motile and non-motile cocci resembling Thiococcus pfennigii (with respect to the presence of bacteriochlorophyll b and bundles of tubular internal membranes). Both types of coccoid bacteria were isolated in pure culture. This paper reports on the properties of the new, motile, coccoid,
purple sulfur bacterium containing bacteriochlorophyll b, and on its genetic relationship to other purple sulfur bacteria.

METHODS

Source of organisms. The coccoid bacteria were isolated from deep-agar dilution series that had been inoculated with a peach-coloured sample of the laminated microbial mats from Great Sippewissett Salt Marsh, Cape Cod, MA, USA. A detailed description of these microbial mats has been given by Nicholson et al. (1987).

Isolation and cultivation. Pure cultures were obtained using repeated deep-agar dilution series (Pfennig, 1978). The basal medium used for isolation and further cultivation of the bacterium contained (per litre distilled water): 0.25 g KH₂PO₄, 0.4 g NH₄Cl, 0.35 g KCl, 20.0 g NaCl, 2.8 g MgSO₄·7H₂O, 0.25 g CaCl₂, 2H₂O, 1.5 g NaHCO₃, 0.3 g Na₂S·9H₂O, 1 ml vitamin solution (Pfennig & Trüper, 1981) and 1 ml trace-element solution SL 12 (Overmann et al., 1992). The basal medium was sterilized, the pH was adjusted to 7.2–7.3 and then the medium was aseptically distributed into culture vessels as described by Pfennig & Trüper (1981). The purity of cultures was checked microscopically and by using growth tests in AC medium (Diffco).

 Cultures were grown phototrophically in 100 ml screw-capped bottles with autoclavable rubber seals. Cultures were incubated at 20–22 °C at a light intensity of 300–500 lx from a tungsten lamp. The light intensity was checked in the 400–700 nm range with a Metrolux K light sensor (Metrwatt). The addition of sterile pyruvate solution to a final concentration of 5 mM was used to obtain high cell yields. Stock cultures were stored at 4 °C in the dark.

Growth experiments were performed in duplicate in 20 ml screw-capped tubes with autoclavable rubber seals. Electron donors and carbon sources were aseptically added to each tube, from sterile stock solutions, to the final concentrations indicated. The utilization of organic carbon sources was tested in the presence of hydrogen carbonate and sulfide in the media. Growth was followed by measuring the optical density at 650 nm with a Bausch & Lomb Spectronic 70 photometer.

Electron microscopy. For negative staining, cells were treated with a 3 % aqueous solution of uranyl acetate. For ultrathin sections, cells were prefixed in glutaraldehyde, fixed in OsO₄ solution, stained with uranyl acetate in 75 % acetone, embedded in Spurr medium, sectioned and then post-stained with lead acetate. Electron microscopy was carried out by Dr H. Lünsdorf (GBF, Braunschweig, Germany).

Pigment and sulfur analyses. Absorption spectra were recorded with a Lambda 2 spectrophotometer (PerkinElmer). The identification of carotenoids was carried out by means of TLC, as described by Eichler & Pfennig (1986), using carotenoids of Thiococcus pfennigii as a reference (Schmidt, 1978). Sulfide was measured colorimetrically (Trüper & Schlegel, 1964) and sulfate was determined turbidimetrically (Dodgson, 1961).

DNA analysis. DNA was isolated using the method of Marmur (1961). The G + C content (mol %) of the DNA was determined by Dr J. Floßdorf (Braunschweig, Germany) according to Floßdorf (1983). Cell material for 16S rDNA sequencing was taken from 1–2 ml well-grown liquid cultures. DNA was extracted and purified by using the QIAGEN genomic DNA buffer set. The PCR amplification and the 16S rDNA sequencing were done as described earlier (Imhoff et al., 1998). Recombinant Taq polymerase was used for the PCR, which was started with the primers 5′-GTTTGATCCTGGCTCAG-3′ and 5′-TACCTTGTTAC-GACTTCA-3′ (positions 11–27 and 1489–1506, according to the Escherichia coli 16S rRNA numbering of the International Union of Biochemistry). Sequences were obtained by cycle sequencing with the SequiTherm sequencing kit (Biozym and the chain-termination reaction (Sanger et al., 1977), using an automated laser fluorescence sequencer (Pharmacia). Sequences were aligned using the CLUSTAL W program (Thompson et al., 1994). The alignment length was from position 29 to position 1381 (according to E. coli numbering). The distance matrix was calculated on the basis of the algorithm according to Jukes & Cantor (1969) with the DNADIST program within the PHYLIP package (Felsenstein, 1989). The FITCH program in the PHYLIP package fitted a tree to the evolutionary distances.

RESULTS

Natural habitat and isolation

During the 1986 ‘Microbial Diversity’ summer course at the Marine Biological Laboratory, Woods Hole, MA, USA, samples were taken from the peach-coloured layer of a laminated microbial mat at Great Sippewissett Salt Marsh, Cape Cod, MA, USA. The material was suspended in sterile culture medium and used as the inoculum for a deep-agar dilution series with basal medium plus 3 mM acetate and 1 % washed agar. After two weeks of incubation in the light,
Thioflavicoccus mobilis gen. nov., sp. nov.

Fig. 2. Electron photomicrograph of negatively stained cells of strain 8321T showing monopolar–monotrichous flagella and pili. Bar, 0.5 µm. Courtesy of H. Lünsdorf, GBF, Braunschweig, Germany.

various types of pinkish, reddish and yellowish-beige colonies developed. Individual colonies were picked and studied microscopically. The majority of the yellowish-beige colonies contained coccoid non-motile cells. Only a few colonies consisted of highly motile cocci. Single colonies were used as inocula for second and third deep-agar dilution series to obtain pure cultures of strains 8320 (coccoid, non-motile) and strain 8321T (coccoid, motile), which were maintained as stock cultures and used for further studies. As strain 8320 resembled *Thiococcus pfennigii*, strain 8321T was studied in more detail.

**Morphology and fine structure**

In well-growing cultures, individual cells of strain 8321T were 0.8–1.0 µm in diameter. Before division by binary fission, rod-shaped to diplococcus-shaped division stages occurred (Fig. 1). The cells were highly motile and stained Gram-negative. In negatively stained preparations, monopolar monotrichous flagella were observed (Fig. 2). Electron microscopic examination of ultrathin sections revealed an intracellular photosynthetic membrane system of the tubular type (Fig. 3). The membrane system resembled that of *Thiococcus pfennigii* (Eimhjellen et al., 1967) and *Thioalkalicoccus limnaeus* (Bryantseva et al., 2000). Ultrathin sections also revealed an outer membrane and a cytoplasmic membrane characteristic of Gram-negative bacteria (Fig. 3).

**Pigments**

Phototrophically grown cultures of strain 8321T appeared yellowish-beige to orange-brown in colour. The absorption spectrum of intact cells was similar to that of *Thiococcus pfennigii* (Eimhjellen et al., 1967). The main absorption maximum at approximately 1025 nm quite clearly indicates the presence of bacteriochlorophyll b. In pigment extracts, bacteriochlorophyll b was identified by co-chromatography with pigments from *Thiococcus pfennigii*. The in vivo maxima at 530, 492 and 462 nm resemble those of the carotenoids of *Thiococcus pfennigii* (Aasen & Liaaen-Jensen, 1967) and indicate the presence of 3,4,3',4'-tetrahydrospirilloxanthin. Co-chromatography of pigment extracts with those from *Thiococcus pfennigii* allowed identification of the main carotenoid as 3,4,3',4'-tetrahydrospirilloxanthin (Schmidt, 1978).

**Physiological properties**

Photolithoautotrophic growth of strain 8321T occurred under anoxic conditions in the light with hydrogen sulfide and elemental sulfur as electron donors. Thiosulfate was not used. The bacterium was strictly anaerobic and obligately phototrophic. No growth occurred in the dark under oxic or microoxic conditions.

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Fig. 3. Electron photomicrograph of an ultrathin section of strain 8321T showing tubular internal membrane structures in longitudinal section (t) and in cross-section (inset). om, Outer membrane; cm, cytoplasmic membrane. Bar, 0.5 µm. Courtesy of H. Lünsdorf, GBF, Braunschweig, Germany.
**Table 1.** Substrate utilization and characteristic properties of strain 8321<sup>T</sup> in comparison to the most closely related bacteria, *Thiococcus pfennigii* and *Thioalkalicoccus limnaeus*

<table>
<thead>
<tr>
<th>Property/substrate</th>
<th>Thioflavivoccus mobilis strain 8321&lt;sup&gt;T&lt;/sup&gt;</th>
<th>Thiococcus pfennigii strain DSM 226</th>
<th>Thioalkalicoccus limnaeus strain A26</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cell shape</td>
<td>Coccus</td>
<td>Sphere</td>
<td>Sphere</td>
</tr>
<tr>
<td>Cell size</td>
<td>0·8–1·0</td>
<td>1·2–1·5</td>
<td>1·3–1·8</td>
</tr>
<tr>
<td>Motility</td>
<td>Monopolar flagella</td>
<td>Non-motile</td>
<td>Non-motile</td>
</tr>
<tr>
<td>Salt optimum (%)</td>
<td>2</td>
<td>0·5–2·0</td>
<td>5</td>
</tr>
<tr>
<td>pH optimum</td>
<td>7·2–7·4</td>
<td>7·2–7·4</td>
<td>8·8–9·5</td>
</tr>
<tr>
<td>DNA G + C content (mol %)</td>
<td>66·5</td>
<td>69·4–69·9</td>
<td>63·6–64·8</td>
</tr>
<tr>
<td>Substrates used:</td>
<td>Propionate</td>
<td>–</td>
<td>+</td>
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<tr>
<td></td>
<td>Fructose</td>
<td>–</td>
<td>+</td>
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<td></td>
<td>Ascorbate</td>
<td>+</td>
<td>–</td>
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<tr>
<td></td>
<td>Peptone</td>
<td>–</td>
<td>(+)</td>
</tr>
<tr>
<td></td>
<td>Casamino acids</td>
<td>–</td>
<td>(+)</td>
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<tr>
<td></td>
<td>Lactate</td>
<td>–</td>
<td>–</td>
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<td></td>
<td>Succinate</td>
<td>–</td>
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<td></td>
<td>Malate</td>
<td>–</td>
<td>+</td>
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<tr>
<td></td>
<td>Fumarate</td>
<td>–</td>
<td>(+)</td>
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</tbody>
</table>

NA, No data available.

All strains had tubular internal membranes, bacteriochlorophyll b, and 3,4,3',4'-tetrahydrospirilloxanthin as the main carotenoid; all strains used sulfide, S<sub>2</sub> and acetate and pyruvate as substrates; none required vitamins and none was able to use thiosulfate. The following additional substrates were tested (concentrations 5 mM), but were not used by strain 8321<sup>T</sup>: arginine, butyrate, benzoate, valerate, glycerol, glycolate, glucose, gluconate, glutamate, caproate, caprylate, crotonate, malonate, mannitol, methanol, ethanol, propanol, tartrate, formate, citrate, 2-oxoglutarate, acetoin, 2,3-butanediol and ethylene glycol.

**DISCUSSION**

In deep-agar dilution tubes from the peach-coloured layer of a laminated microbial mat, yellowish-beige colonies were dominant in all dilution steps. In most of the cases, suspensions of individual colonies of this type contained non-motile cocoid cells with internal sulfur globules characteristic of *Thiococcus pfennigii*. Surprisingly, a few colonies contained motile cocoid cells. Therefore, initially the question arose as to whether there are motile variants of the non-motile *Thiococcus pfennigii* or whether the motile cocci represented a separate and new taxon. The results presented in this paper and summarized in Table 1 demonstrated that, apart from motility, many other traits of strain 8321<sup>T</sup> are different from those of the typical *Thiococcus pfennigii* strain DSM 226 (8013). Apparent similarities in cell shape, colour of colonies, pigment content and internal membrane structure were counteracted by other significant differences. Only acetate, pyruvate and ascorbate were used as organic substrates by strain 8321<sup>T</sup>, whereas *Thiococcus pfennigii* strains were generally able to utilize a number of additional organic compounds. Also, the G + C content (mol %) of the DNA of strain 8321<sup>T</sup> was significantly lower than that of *Thiococcus pfennigii*. Furthermore, the genetic relationship between the motile strain, 8321<sup>T</sup>, and strains of *Thiococcus pfennigii* (strains DSM 226 and 8320) and *Thioalkalicoccus limnaeus*.
**Thioflavicoccus mobilis gen. nov., sp. nov.**

*Thioflavicoccus mobilis* (mo’bi.lis. L. adj. mobilis mobile).

Cells are coccoid, short rod-shaped to diplococcus-shaped before cell division. Cocci are 0.8–1.0 μm in diameter. Motile by monopolar monotrichous flagella. Possess an internal photosynthetic membrane system of tubular type. The colour of cell suspensions is yellowish-beige to orange-brown. The absorption spectrum of living cell suspensions exhibits maxima at 410, 462, 492, 530 and 1025 nm, with shoulders at 602 nm.

**Description of Thioflavicoccus gen. nov.**

*Thioflavicoccus* (Thi’o.flä’vi.co’c’us. Gr. n. thios sulfur; L. masc. adj. flavus golden-yellow, beige; L. masc. n. coccus sphere; M.L. masc. n. Thioflavicoccus beige-yellow coccus with sulfur).

Cells are spherical with diplococcus-shaped division stages, motile by flagella, and multiply by binary fission. Gram-negative; belong to the γ-Proteobacteria. Contain internal photosynthetic membranes of tubular shape. Photosynthetic pigments are bacteriochlorophyll b and carotenoids. Obligately phototrophic and strictly anaerobic. Photolithoautotrophic growth under anoxic conditions in the light, with sulfide and elemental sulfur as electron donors. During oxidation of sulfide, elemental sulfur is transiently stored inside the cells in the form of highly refractile globules; the final oxidation product is sulfate. In the presence of sulfide and bicarbonate, simple organic substrates are photoassimilated. Mesophilic bacteria, growing well at 20–30 °C and neutral pH (pH range 6.5–7.5) and requiring sodium chloride for optimum growth. The G+C content of the DNA is 66.5 mol% (Bd). The type species is *Thioflavicoccus mobilis* sp. nov.

**Description of Thioflavicoccus mobilis sp. nov.**

*Thioflavicoccus mobilis* (mo’bi.lis. L. adj. mobilis mobile).

Fig. 4. Genetic relatedness of strain 8321T to *Thiococcus pfennigii* (strains DSM 226 and 8320) and *Thioalkalicoccus limnaeus* (strains A26 and A31) as well as to other purple sulfur bacteria, based on 16S rDNA sequence similarities. Numbers at the nodes give confidence values of bootstrap analyses with 100 resamplings.
and 835 nm. Photosynthetic pigments are bacteriochlorophyll b and 3,4,3′,4′-tetrahydrospirilloxanthin as the main carotenoid. The metabolism is obligately phototrophic and strictly anaerobic. Photolithoautotrophic growth occurs in the light with hydrogen sulfide as electron donor. Globules of elemental sulfur are accumulated inside the cells. The final oxidation product is sulfate. Thiosulfate is not used. Sodium chloride is required and ascorbate are used as organic substrates. Growth factors are not required. Sodium chloride is required for growth. Good growth occurs at 25–30 °C, pH 7.2–7.4, with 1–3 % NaCl (the optimum is at 2 %), and at a light intensity of 500 lx from a tungsten lamp. The habitat is laminated microbial mats of salt marshes.

ACKNOWLEDGEMENTS

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REFERENCES


